


植物品种 特异性、一致性和稳定性测试 15周年论文集

ZHIWU PINZHONG
TEYIXING YIZHIXING HE WENDINGXING CESH
15 ZHOUNIAN LUNWENJI



农业部科技发展中心
农业部植物新品种测试中心  编

植物品种特异性、一致性和 稳定性测试15周年论文集

农业部科技发展中心 编
农业部植物新品种测试中心

中国农业出版社



图书在版编目 (CIP) 数据

植物品种特异性、一致性和稳定性测试 15 周年论文集 /
农业部科技发展中心, 农业部植物新品种测试中心编. —
北京: 中国农业出版社, 2016. 9
ISBN 978-7-109-21998-4

I. ①植… II. ①农…②农… III. ①植物—品种—
中国—文集 IV. ①Q94-53

中国版本图书馆 CIP 数据核字 (2016) 第 189446 号

中国农业出版社出版

(北京市朝阳区麦子店街 18 号楼)

(邮政编码 100125)

策划编辑 闫保荣

文字编辑 李 晓

北京印刷一厂印刷 新华书店北京发行所发行
2016 年 9 月第 1 版 2016 年 9 月北京第 1 次印刷

开本: 889mm×1194mm 1/16 印张: 18.5

字数: 700 千字

定价: 60.00 元

(凡本版图书出现印刷、装订错误, 请向出版社发行部调换)

编 委 会

主 任 杨雄年

副主任 周云龙

委 员 (以姓氏笔画为序)

王 威	王凤华	邓 超	刘 洋	孙连发
李汝玉	李硕碧	杨 坤	杨旭红	余 毅
沈 奇	张如莲	张建华	陈 红	陈海荣
夏来坤	徐 岩	徐振江	唐 浩	堵苑苑
崔野韩	韩瑞玺	魏兴华		

主 编 杨旭红

副主编 唐 浩 崔野韩

编 委 (以姓氏笔画为序)

王 威	王凤华	王艳平	卢 新	刘志勇
刘丽娟	刘迪发	刘艳芳	李华勇	李祥羽
杨 扬	杨 坤	杨旭红	杨江龙	应东山
张 晗	张建华	张春宵	张浙峰	陈 红
陈海荣	郑永胜	徐振江	高 玲	唐 浩
黄志城	崔野韩	赖运平	解艳华	褚云霞
管俊娇	熊国富	颜国荣	魏兴华	

植物品种的特异性 (distinctness)、一致性 (uniformity) 和稳定性 (stability), 简称为 DUS。DUS 测试是指依据相应植物属 (种) 的测试技术与标准, 通过田间种植试验或室内分析对待测试品种的 DUS 进行评价的过程。

我国于 1999 年 4 月 23 日加入国际植物新品种保护联盟 UPOV, 标志着植物新品种保护制度的正式实施。截至 2015 年 12 月 31 日, 我国品种权申请量居 UPOV 成员第二位, 仅农业部受理的申请达 15 552 件, 授予品种权 6 258 件。为保证授权的客观、公正和科学, 农业部于 2001 年在全国一级农业生态区成立了植物品种 DUS 测试机构, 包括 1 个测试中心和 14 个测试分中心, 2014 年和 2015 年分别新建了 4 个分中心, 现在分中心已达 22 家。DUS 测试机构成立以来, 共完成 69 个植物属 (种) 10 000 多个品种测试、100 多个品种真实性鉴定, DUS 测试为植物品种的授权、市场监管和司法鉴定等提供了强有力的技术支撑。

2016 年 1 月 1 日实施的《中华人民共和国种子法》规定审定、登记和保护的植物品种必须具备特异性、一致性和稳定性, 首次明确了 DUS 测试的法律地位, 使 DUS 测试在引领育种技术创新、种业管理等方面将发挥越来越重要的作用。

目前 DUS 测试体系团队共有人员 159 人。根据测试体系二期项目建设规划, 到 2020 年, 将新建 15 个测试分中心、改 (扩) 建 1 个测试中心和 14 个测试分中心, 挂牌建立 28 个多年生无性繁殖植物新品种测试站, DUS 测试人员和测试品种数量必将大幅度增加。为加强植物品种测试技术研究和人才培养, 2015 年 5 月, 农业部科技发展中心 (农业部植物新品种测试中心) 成立了测试技术专业委员会, 下设信息技术、生物技术、大田作物、蔬菜、观赏植物和果树 6 个技术组。为充分发挥各技术组的作用, 鼓励 DUS 测试体系年轻人在扎实做好 DUS 测试工作的同时, 加强测试技术的学习、研究与交流, 促进优秀人才脱颖而出, 打造一支与国际接轨、专业技术过硬的审查测试人员队伍, 农业部科技发展中心在各测试分中心支持下, 收集、梳理了自测试体系成立以来至 2015 年 12 月 31 日 15 年间, 测试人员和管理人员以第一作者发表的 DUS 测试技术研究论文 146 篇。由于汇编篇幅限制, 这些论文交由 DUS 测试技术委员会评选后, 仅对评选出的 44 篇优秀论文列入汇编。

本汇编论文研究内容全面, 涉及大田作物、蔬菜、观赏植物和果树等植物品种的表



型测试技术、分子测试技术和信息技术等方面，吸收和借鉴了国外 DUS 测试技术最新成果，并结合我国 DUS 测试实际情况，实用性强，对从事 DUS 测试技术研究以及新入职的 DUS 测试人员具有较高参考价值。

本汇编论文是测试体系成立以来，对国内外 DUS 测试技术经验的总结和探索，凝结了测试体系全体人员的智慧和汗水，在此向大家表示感谢。同时，由于时间仓促，难免存在错误疏漏之处，欢迎大家批评指正。此外，随着 DUS 测试技术的发展和测试方法的改进，希望大家再接再厉，继续对 DUS 测试技术进行深入研究，提高我国 DUS 测试技术和水平，为现代种业发展做出新的贡献。

编委会

2016 年 3 月 23 日

目 录

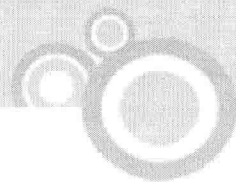
前言

测试技术研究 (1)

Impact of Light-induced Changes in Anthocyanin Coloration Intensity on DUS Test	Yanfang Liu et al (3)
Sensitivity of Panicle Related Quantitative Characteristics of Rice DUS Test to Environmental Changes	Yanfang Liu et al (8)
SRAP Markers and Morphological Traits Could Be Used in Test of Distinctiveness, Uniformity, and Stability (DUS) of Lettuce (<i>Lactuca sativa</i>) Varieties	Lijuan Liu et al (14)
Statistic Analysis on Quantitative Characteristics for Developing the DUS Test Guideline of <i>Ranunculus asiaticus</i> L.	Yanfang Liu et al (24)
2 种栽培模式对辣椒 DUS 测试性状表达的影响	高玲等 (33)
78 份高粱种质的遗传关系和遗传结构解析	张春宵等 (39)
SRAP 标记与形态学标记在西瓜 DUS 测试中的比较	刘丽娟等 (51)
不同生态环境对郑单 958 部分数量性状的影响研究	黄志城等 (59)
大白菜高通量 SSR 标记鉴定体系的建立和应用	张晗等 (66)
大豆 DUS 测试标准品种测试性状表达差异性分析	解艳华 (76)
大豆 DUS 测试主要数量性状的变异及概率分布的研究	王威等 (80)
大麦 DUS 测试标准品种的遗传多样性分析及指纹图谱的构建	王艳平等 (83)
非洲菊新品种 DUS 测试数量性状分级及形态性状多样性研究	褚云霞等 (89)
甘蓝型油菜核心 SSR 引物筛选及指纹图谱构建	赖运平等 (96)
基于 DUS 测试的标准品种形态性状稳定性和重要性分析	唐浩等 (107)
基于 DUS 测试的上海粳稻地方品种遗传多样性分析	黄志城等 (118)
基于 DUS 测试的水稻标准品种形态性状多样性分析	唐浩等 (129)
基于 DUS 测试性状的玉米标准品种形态多样性分析	陈海荣等 (137)
基于荧光检测技术的小麦品种 SSR 鉴定体系的建立	郑永胜等 (144)
设施栽培对玉米 DUS 测试性状及产量相关性状表达的影响	刘迪发等 (152)
适宜南京地区水稻新品种 DUS 测试标准品种的筛选	李华勇等 (159)
水稻 DUS 测试新品种品质性状的差异性分析及应用	徐振江等 (165)
水稻新品种 DUS 测试技术	魏兴华等 (170)
水稻新品种 DUS 测试数量性状特异性统计分析判别研究	徐振江等 (173)
水稻新品种测试的标准品种 DNA 指纹图谱多样性分析	唐浩等 (177)
甜瓜种质资源若干果实数量性状差异研究及分级探讨	颜国荣等 (185)
同名甘蓝型油菜品种比较分析	赖运平等 (189)
豌豆新品种 DUS 测试指南的研制	张浙峰等 (199)



我国芒果主栽品种遗传多样性分析及指纹图谱构建	王明等 (205)
与水稻品种特异性、一致性、稳定性 (DUS) 测试性状相关联的分子标记分析	王艳平等 (214)
玉米 DUS 测试标准品种的 SSR 分子指纹图谱的构建	张金渝等 (223)
玉米 DUS 测试标准品种在云南的差异性分析	张建华等 (230)
玉米 DUS 测试性状在春播与夏播间的表达差异研究	杨江龙等 (234)
玉米 DUS 测试主要数量性状分级方法的研究	王凤华等 (238)
玉米新品种 DUS 测试中数量性状的适宜样本容量研究	李祥羽 (242)
栽培措施对玉米 DUS 测试数量性状表达的影响	管俊娇等 (246)
 国内外品种保护	 (251)
对 UPOV 公约 91 文本的分析与思考	卢新等 (253)
建立我国非主要农作物品种国家登记制度的探讨	杨旭红等 (257)
美国植物新品种保护审查制度	崔野韩等 (260)
青海省植物新品种保护工作现状与思考	熊国富 (263)
我国建立实质性派生品种制度的必要性讨论	陈红等 (266)
新西兰植物新品种保护及其对中国的启示	杨扬等 (270)
 信息技术研究	 (273)
植物已知品种数据库构建方法的研究	杨坤等 (275)
植物资源图像采集初报	刘志勇等 (281)



测试技术研究

植物品种特异性、一致性和稳定性测试15周年论文集



Impact of Light-induced Changes in Anthocyanin Coloration Intensity on DUS Test

Yanfang Liu^{1,2}, Shiquan Shen^{3,1}, Jianhua Zhang^{1,2}, Xiaohong Yang^{1,2},
Jiangmin Wang^{1,2}, Hui Zhang^{1,2}, Qingmei Huang^{1,2}, Peng Zhang^{1,2}, Junjiao Guan^{1,2}

(1. Quality Standard and Testing Technology Research Institute, Yunnan Academy of Agricultural Sciences, Kunming 650205, China; 2. Kunming DUS Test Station, Ministry of Agriculture, Kunming, China; 3. Division of Scientific Research Management, Yunnan Academy of Agricultural Sciences, Kunming 650205, China)

Abstract: Intensity of anthocyanin coloration is a frequently-used characteristics in DUS tests, and its expressions are susceptible to the change of light intensity. Based on the requirement that characteristics used in DUS tests should be consistent and repeatable in a particular environment, research related to the impact of light-induced changes in anthocyanin coloration intensity on DUS test is essential to reveal the problems of current DUS tests. Results gained in this study showed that plants of the same rice variety tested in the given environment had identical expressions in the same characteristic of anthocyanin coloration intensity, indicating the characteristics were consistent; if plants were tested in environments with different light intensity, the characteristics had various expressions, especially the results of DUS test differed from each other. The fluctuation would bring great troubles to DUS test, and it would also obviously lead to problems for harmonization of variety descriptions. This study proposed approaches for this situation, including deleting characteristics of anthocyanin coloration intensity, making the growing conditions fixed, or making DUS tests of the same plant genera or species concentrated.

Keywords: DUS test; light intensity; intensity of anthocyanin coloration; characteristics

1 Introduction

In recent years, more and more attention has been paid to intellectual property rights due to the increasing international cooperation and exchanges. Plant breeder's right (PBR), or plant variety protection (PVP) is one form of intellectual property rights. PBR has drawn public interest with the development of marketing of plant varieties^[1,2]. To protect the legal rights of plant breeders, Chinese government has issued "Regulation of the People's Republic of China on the Protection of New Varieties of Plants" (hereinafter referred to as "Regulation") in 1997. China became the 39th member of International Union for the Protection of New Varieties of Plants (UPOV) in 1999, and accordingly, new varieties of plants can be applied for protection in China since then^[3]. In accordance with the "Regulation", PBR is granted to the plant variety which can be distinguished (D) from any other variety commonly known when the application is filed, and which is considered as sufficiently uniform (U) and stable (S)^[4,5]. It is noted in UPOV Convention that a variety is defined by its characteristics and therefore those characteristics are the basis for DUS^[4,5]. Besides examining whether a variety meets DUS criteria, DUS test can also generate a description of variety. Each plant genera or species has an exclusive set of characteristics in corresponding DUS test guideline^[6]. These characteristics are generally morphological characteristics, e. g., plant height, plant width, or physiological characteristics, e. g., growing period^[6,7]. The range of expression of each characteristic in the test guidelines is divided into a number of states for the benefit of description, and



the wording of each state is attributed a numerical "Note" [4]. Characteristics can be grouped as qualitative characteristics, quantitative characteristics, and pseudo-qualitative characteristics based on their different expression ways. Qualitative characteristics are expressed in discontinuous ways, and these states are self-explanatory and independently meaningful [e. g., sex of plant: dioecious female (1), dioecious male (2), monoecious unisexual (3), monoecious hermaphrodite (4)]. Expressions of quantitative characteristics differ from one extreme to the other, and such differences are continuous, and thus their expression can be recorded on a linear scale [e. g., width of leaf: very narrow (1), narrow (3), medium (5), wide (7), very wide (9)] [4,5]. As a rule, qualitative characteristics are usually independent from environment, while quantitative characteristics are often polygenically inherited and can be influenced by environmental conditions [8-10].

Anthocyanin coloration is one of the most common characteristic frequently used in DUS test. Above 85% DUS test guidelines adopt one or more characteristic (s) of anthocyanin coloration [11]. Anthocyanin coloration can appear in different parts of plant, e. g., root, stem, leaf, flower, fruit, seed, etc. Characteristics of anthocyanin coloration include two kinds: absence or presence of anthocyanin coloration, and intensity of anthocyanin coloration. The first kind is generally described as "plant part: anthocyanin coloration", and the states include "absent" (Note 1) and "present" (Note 9). This kind of characteristics is absolutely qualitative characteristics. Description for the second kind is "plant part: intensity of anthocyanin coloration." This kind of characteristics is quantitative characteristic, and the states include "very weak (1), weak (3), medium (5), strong (7), very strong (9)." The states can also be "very weak (1), weak (2), medium (3), strong (4), very strong (5)" or "weak (1), medium (2), strong (3)" if the span of intensity fluctuation is small.

Intensity of anthocyanin coloration can be influenced by cultivation conditions, especially light and nutrient, e. g., growth hormone NAA, carbohydrate [12-14]. Light plays an important role in the formation of anthocyanin. Strong light can induce more accumulation of anthocyanin [12,13]. According to requirements of DUS test, any characteristic listed in DUS Test Guideline should be sufficiently consistent and repeatable in a particular environment [4]. Since characteristics related to anthocyanin coloration are frequently used in DUS test, and intensity of anthocyanin coloration can be influenced by light changes, it is of great importance to research the impact of light-induced changes in anthocyanin coloration intensity on DUS test. Studies on the influence of light changes to the changes of anthocyanin coloration intensity are involved in lots of papers. In contrast, report related to the impact of those changes on DUS test is yet unavailable.

This study conducts research on rice, the most important crop with the largest application and authorization in China. In DUS test of rice, there are eight characteristics related to intensity of anthocyanin coloration. Based on the "Guideline for the Conduct of Tests for Distinctness, Uniformity and Stability—Rice" (hereinafter referred to as "Guideline") [15], the impact of light-induced changes in anthocyanin coloration intensity on DUS test was analyzed here.

2 Subject and Methods

Two rice varieties were selected in this study. They were Liheizimi and Yunlu 99, marked as A1 and A2, respectively. Both varieties are typical representatives of anthocyanin coloration.

The test was carried out in 2009 and 2011. It began in April and ended in October. It was conducted in Yunnan Academy of Agriculture Science. Seeds were sown in trays for wet seedling nursing in April. One month later, the four-leaved seedlings were transferred to 30-cm-size pots. Four plants were grown in one pot.

After transferring, plants of the two varieties were grown in three conditions with different light intensity. The three conditions were marked as B1 (greenhouse condition), B2 (shelter condition), and B3 (outdoor condition). Light intensity of the three conditions increases in sequence. Three repeats were set for each treatment.

DUS test was carried out on the eight characteristics of anthocyanin coloration intensity in strict accordance with "Guideline." The eight characteristics are "leaf sheath: intensity of anthocyanin coloration," "lemma: intensity of anthocyanin coloration of keel (early observation)," "lemma: intensity of anthocyanin coloration of area below apex (early observation)," "lemma: intensity of anthocyanin coloration of apex (early observation)," "stem: intensity of anthocyanin

coloration of nodes,” “lemma: intensity of anthocyanin coloration of keel (late observation),” “lemma: intensity of anthocyanin coloration of area below apex (late observation),” “lemma: intensity of anthocyanin coloration of apex (late observation)”, respectively. The eight characteristics were marked from C1 to C8 in order. Observations were recorded, and each expression state was attributed a numerical “Note” .

Table 1 Treatments

Variety	B1 (Greenhouse)	B2 (Shelter)	B3 (Outdoor)
A1 (Liheizimi)	A1B1	A1B2	A1B3
A2 (Yunlu99)	A2B1	A2B2	A2B3

3 Results

The observations of the eight characteristics were translated into corresponding “Note” . The “Note” was used to judge the expression states of the eight characteristics of the same variety under the above mentioned environments. The results indicate that plants of three repeats of the same treatment had the same expression in the same characteristic. In contrast, for the same variety, the same characteristic had different expressions in environments with different light intensity (Table 2) . Changes in light intensity were positively correlated with changes in anthocyanin coloration intensity. Environment of B3 had stronger light than B1 and B2, and expressions of anthocyanin coloration intensity in B3 were the strongest in each characteristics. Figures 1 and 2 also show that intensity of anthocyanin coloration increased from left to right, or from B1 to B3. Characteristic expressions differed greatly (Table 2, Fig. 1 and 2) . Taking characteristic of “lemma: intensity of anthocyanin coloration of keel (late observation)” (C6) as an example, expression of Liheizimi (A1) in B1 was very weak (Note1), while expression in B3 was very strong (Note 9) (Table 2 and Fig. 2) .

Table 2 DUS test of the eight characteristics

Characteristics	Expressions and Notes					
	A1B1	A1B2	A1B3	A2B1	A2B2	A2B3
C1	vw (3)	vw (3)	w (5)	vw (1)	vw (1)	w (3)
C2	w (3)	m (5)	s (7)	vw (1)	vw (1)	m (5)
C3	w (3)	m (5)	vs (9)	vw (1)	w (3)	s (7)
C4	s (7)	s (7)	vs (9)	s (7)	s (7)	vs (9)
C5	m (5)	m (5)	s (7)	vw (1)	vw (1)	w (3)
C6	vw (1)	m (5)	vs (9)	w (3)	w (3)	m (5)
C7	m (5)	s (7)	vs (9)	vw (1)	w (3)	s (7)
C8	s (7)	vs (9)	vs (9)	s (7)	vs (9)	vs (9)

(1) C1 - C8 are “leaf sheath: intensity of anthocyanin coloration,” “lemma: intensity of anthocyanin coloration of keel (early observation),” “lemma: intensity of anthocyanin coloration of area below apex (early observation),” “lemma: intensity of anthocyanin coloration of apex (early observation),” “stem: intensity of anthocyanin coloration of nodes,” “lemma: intensity of anthocyanin coloration of keel (late observation),” “lemma: intensity of anthocyanin coloration of area below apex (late observation),” “lemma: intensity of anthocyanin coloration of apex (late observation)”, respectively.

(2) In “expressions and Notes,” figures between brackets are Notes; outside brackets, vw means very weak, w: weak, m: media, s: strong, vs: very strong.

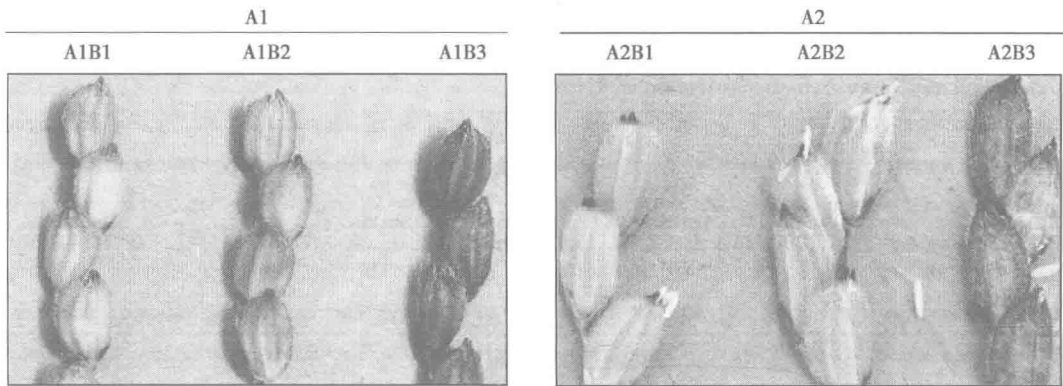


Fig. 1 Anthocyanin coloration intensity of lemma (early observation)



Fig. 2 Anthocyanin coloration intensity of lemma (late observation)

4 Discussion

Difference in light intensity will cause fluctuation in the intensity of anthocyanin coloration, and thus has impact on DUS test. Besides light, other factors, e. g. , nutrients can also have impact on accumulation of anthocyanin^[14]. For field crop, natural light is difficult to control compared with other factors, and that is why light was chosen as a variable here.

DUS test is the basis for the granting of plant breeder's right. Any characteristic used in DUS test should be sufficiently consistent and repeatable in a particular environment^[4,5]. In this study, results from three repeats of one treatment were the same, indicating the eight characteristics of anthocyanin coloration intensity were consistent in a given environment. However, results of the same characteristic from the same variety in environments with different light intensity were various. The fluctuation would bring great troubles to DUS test, and it would also obviously lead to problems for harmonization of variety descriptions. In China, there are 14 stations responsible for the DUS tests of different plant genera or species. The 14 stations are located in different ecological regions, and have different light intensity. Therefore, the same variety tested in different DUS test stations may have different results, and the results of different varieties of the same plant genera or species tested in different DUS test stations are not comparable with each other. To solve these problems, three approaches are proposed here: ① Characteristics of anthocyanin coloration intensity could be deleted in the DUS test guidelines of field crops. ② Growing condition with fixed light intensity and other environment factors can be used for certain plant genera or species, e. g. horticulture crops, vegetables, etc. ③ For field crops, all the varieties of the same crop can be tested in one fixed DUS test station to avoid the absence of comparability of results gained in different stations.



>>> Acknowledgment

This study was assisted by Special Fund for Agro-scientific Research in the Public Interest of Ministry of Agriculture, China (200903008 - 14) .

>>> References

- [1] Jördens R. Progress of plant variety protection based on the International Convention for the protection of new varieties of plants (UPOV Convention) . World Patent Information, 2005, 27 (3): 232 - 243.
- [2] Tripp R, Louwaars N, Eaton D. Plant variety protection in developing countries. A report from the field. Food Policy, 2007, 32 (3): 354 - 371.
- [3] Liu YF, Zhang JH, Yang XH, et al. Analysis on the functions and properties of the preliminarily determined characteristics listed in DUS Test Guideline of *Tagetes L.* Agricultural Science and Technology, 2012, 13 (8): 1789 - 1792.
- [4] UPOV. TG/1/3. General introduction to the examination of distinctness, uniformity and stability and the development of harmonized descriptions of new varieties of plants. Geneva, Switzerland, 2002.
- [5] UPOV. TGP/8/1. Trial design and techniques used in the examination of distinctnessm uniformity and stability. Geneva, Switzerland, 2010.
- [6] Liu YF, Zhang JH, Lv B, et al. Statistic analysis on quantitative characteristics for developing the DUS Test Guideline of *Ranunculus asiaticus L.* Journal of Integrative Agriculture, 2013, 12 (6): 971 - 978.
- [7] Furones-Pérez P, Fernández-López J. Usefulness of 13 morphological and phenological characteristics of sweet chestnut (*Castanea sativa Mill.*) for use in the DUS test. Euphytica, 2009, 167 (1): 1 - 21.
- [8] Liu YF, Yang XH, Zhang JH, et al. Distinctness determination of DUS test on some quantitative characteristics of rice. Advances in Intelligent and Soft Computing, 2012, 134: 943 - 951.
- [9] Borys J, Kowalczyk B, Waszak J. Distinctness, uniformity and stability testing of tomato varieties in Poland. Acta Physiologiae Plantarum, 2000, 22 (3): 225 - 229.
- [10] Liu YF, Zhang JH, Yang XH, et al. Application of statistic analysis on the selection of quantitative characteristics listed in the DUS Test Guideline of *Tagetes L.* Agricultural Science and Technology, 2012, 13 (9): 2021 - 2023, 2033.
- [11] http://www.upov.int/test_guidelines/en/.
- [12] Zhang W, Curtin C, Kikuchi M, et al. Integration of jasmonic acid and light irradiation for enhancement of anthocyanin biosynthesis in *Vitis vinifera* suspension cultures. Plant Science, 2002, 162 (3): 459 - 468.
- [13] Tsurunaga Y, Takahashi T, Katsube T, et al. Effects of UV - B irradiation on the levels of anthocyanin, rutin and radical scavenging activity of buckwheat sprouts. Food Chemistry, 2013, 141 (1): 552 - 556.
- [14] Simões C, Bizarri CHB, Cordeiro LDS, et al. Anthocyanin production in callus cultures of *Cleome rosea*: modulation by culture conditions and characterization of pigments by means of HPLC - DAD/ESIMS. Plant Physiology and Biochemistry, 2009, 47 (10): 859 - 903.
- [15] UPOV. TG/16/8. Guidelines for the conduct of tests for distinctness, uniformity and stability. Geneva, Switzerland, 2004.

Sensitivity of Panicle Related Quantitative Characteristics of Rice DUS Test to Environmental Changes

Yanfang Liu^{1,2,a}, Jiangmin Wang^{1,2}, Qingmei Huang^{1,2}, Jianhua Zhang^{1,2,b},
Xiaohong Yang^{1,2}, Hui Zhang^{1,2}, Peng Zhang^{1,2}, Junjiao Guan^{1,2}

(1. Quality Standard and Testing Technology Research Institute, Yunnan Academy of Agricultural Sciences, Kunming 650205, China; 2. Kunming DUS Test Station, Ministry of Agriculture, Kunming 650205, China;
a. liuyf528@163.com; b. eapvpf2@163.com)

Abstract: Panicle related characteristics of rice have close relationship with rice yield. Among the 48 panicle related characteristics of rice DUS test, 8 quantitative characteristics were used to research whether environmental changes had impact on their expression. Results showed that, ①in general, measurements of the 8 characteristics of the same variety in different environments were visibly different, and when measurements were translated into “notes” according to “DUS Test Guideline of Rice”, such difference could be narrowed, indicating the 8 characteristics were sensitive to environmental changes, while corresponding expression states and “notes” could weaken such influence; ②from the results of SPSS statistic analysis, environment factor were less influential than variety factor to the 8 characteristics; ③compared with previous research, the 8 panicle related quantitative characteristics were less affected by environmental changes than quantitative characteristics of vegetative growing period; ④ “notes” difference of the same variety in different environments were correlated with significance analysis of environment impact on characteristic expression, revealing the potential of statistic analysis in checking the availability of results of DUS tests gained from different test locations. To reduce the impact of environmental changes to characteristic expression, suggestion of fixing the DUS test of the same plant genus and species in one appointed station was also made in this study.

Keywords: DUS test; rice; panicle related characteristics; environment

1 Introduction

Panicle related characteristics of rice are research highlights and attracts great attention worldwide due to their direct bearing on rice yield [1,2]. Panicle related characteristics include quantitative characteristics, e.g. length or width of grain, qualitative characteristics, e.g. phenol reaction of lemma, and pseudo-qualitative characteristics, e.g. colour of glume [3]. Generally, quantitative characteristics are sensitive to environment changes [4-6]. That means, expression of quantitative characteristics can be different in various growing condition. Instead, qualitative characteristics are relatively stable and are less affected by environmental fluctuation [4-6].

Panicle related characteristics of rice are also important characteristics in rice DUS test. Among 78 characteristics of rice DUS test, 48 are panicle related characteristics, accounting for 62% [3]. DUS test is to check whether a bred variety is a new variety in following three aspects; whether the said variety (candidate variety) is distinct (D) in one or more characteristics from any other varieties known or used in market; whether plants of the said variety are uniform (U) in every characteristics; whether characteristics of the said variety are stable (S) after continuous growing cycles [7,8]. Both uniformity and stability are to test the homogeneity of morphological traits of the said variety, where uniformity empha-

原载于: Advomced Materials Research, 2014, 1028: 368-373.

sized the homogeneity among individual plant in the same time, and stability emphasizes the homogeneity among populations in different growing cycles [8]. Obviously, DUS test is the precondition of the authorization of Plant Breeders Right (PBR) where the variety is applied for protection as a new variety [9,10]. Besides, DUS test can establish description for each variety, and provide basis for the exchanges of variety information, especially, the construction of information database of plant variety [11,12].

Rice is the most important food crop and has the largest application for PBR in China [4,13]. This study took rice as the materials, and panicle related quantitative characteristics as the target to research the variation of characteristic expression in environments with difference of temperature and light intensity, and thus, to explore the sensitivity of panicle characteristics to environmental changes.

2 Subject & Methods

Rice varieties of Chugeng 27 (A1), Diantun 502 (A2) and Yunlu 29 (A3) were chosen to conduct this study. The three varieties were bred in Yunnan, and has been used for years in Southern China. Chugeng 27 is suitable to be grown in areas with altitude of 1500 - 1850, and its growing period can last 170 - 175 days. Diantun 502 can be grown in areas with altitude of less than 1500, and its growing period is about 155 days. Among the three varieties, Diantun 502 has the shortest growing period and adapts to warm whether, while Yunlu 29 has the longest growing period and adapts to cool environment, and Chugeng 27 is in between.

Instead of paddy field, the trials were carried out in the form of pot planting, and thus, treatments in different environments (greenhouse, shelter and outdoor) were easy to be achieved. In 2009 and 2011, grains of the three varieties were sown in trays for wet seedling nursing in April. After one-month nursing, four-leaved seedlings were transferred to pots with 30 cm diameter, and each pot could hold 4 plants. Plants were treated in three different environments, namely outdoor (B1), shelter (B2) and greenhouse (B3) (Table 1). The three environments were different in temperature and light intensity. On average, B1 was around 28.2℃ from 8:00 to 18:00, B2 was 25.5℃, and B3 was 33.2℃. For each treatment of each variety, 3 repeats were set to avoid random error. Aside from temperature and light intensity, other growing conditions were the same, including irrigation, fertilization, disease or pest control.

Table 1 Treatments

Variety	Outdoor (B1)	Shelter (B2)	Greenhouse (B3)
Chugeng27 (A1)	A1B1	A1B1	A1B1
Diantun502 (A2)	A2B1	A2B1	A2B1
Yunlu29 (A3)	A3B1	A3B1	A3B1

DUS test were carried out in accordance with “DUS Test Guideline of Rice” (hereinafter referred to as “Guideline”). 8 quantitative characteristics, which are panicle related, were measured (Table 2). Each characteristic was measured in 20 plants of a variety. The measurements were given corresponding “notes” based on the ranges of expression states of each characteristic (Table 3). Expression ranges the 8 characteristics are all divided into 9 states. Taking characteristic of “panicle: length” for example, expression states for the “1 - 9” notes can be described as “very short (Note 1), very short to short (Note 2), short (Note 3), short to medium (Note 4), medium (Note 5), medium to long (Note 6), long (Note 7), long to very long (Note 8), very long (Note 9)”. SPSS, a commonly used statistics software, was also applied to conduct further analysis.

Table 2 8 panicle related quantitative characteristics

No.	Name of Characteristics	Range of Note
C45	panicle: length	1~9
C59	panicle: number of grains	1~9