Bong Hak Hyun, M.D. Gene L. Gulati, Ph.D. John K. Ashton, M.D.

#### Bong Hak Hyun, M.D., D.Sc. (Med.)

Professor of Pathology, University of Medicine and Dentistry of New Jersey—Rutgers Medical School, Piscataway, New Jersey

Director of Laboratories, Muhlenberg Hospital, Plainfield, New Jersey Visiting Professor of Pathology, Yonsei University Medical Center, Seoul, Korea

#### Gene L. Gulati, Ph.D.

Section Chief, Hematology Laboratory, Muhlenberg Hospital, Plainfield, New Jersey Adjunct Assistant Professor of Pathology, University of Medicine and Dentistry of New Jersey—Rutgers Medical School, Piscataway, New Jersey

### John K. Ashton, M.D.

Clinical Associate Professor of Pathology, University of Medicine and Dentistry of New Jersey—Rutgers Medical School, Piscataway, New Jersey Attending Pathologist, Muhlenberg Hospital, Plainfield, New Jersey



IGAKU-SHOIN New York · Tokyo Interior Design by Hudson River Studio
Cover Design by Hudson River Studio
Typesetting by Achorn Graphic Services, Inc. in Times Roman

Published and distributed by IGAKU-SHOIN Ltd., 5-24-3 Hongo, Bunkyo-ku, Tokyo IGAKU-SHOIN Medical Publishers, Inc. 1140 Avenue of the Americas, New York, N.Y. 10036

© First edition 1986 by IGAKU-SHOIN Ltd., Tokyo. All rights reserved. No part of this book may be translated or reproduced in any form by print, photo-print, microfilm or any other means without written permission from the publisher.

#### Library of Congress Cataloging in Publication Data

Hyun, Bong Hak, 1922– Color atlas of clinical hematology.

Includes bibliographies and index.

- 1. Blood—Examination. 2. Hematology—Atlases.
- I. Gulati, Gene L. II. Ashton, John K., 1934-
- III. Title. [DNLM: 1. Hematology—methods—atlases.

WH 17 H999c]

RB45.H98 1985 616.1'5 84-25275

ISBN: 0-89640-110-3 (New York) ISBN: 4-260-14110-4 (Tokyo)

Printed and bound in Japan

10987654321

## Preface

Hematology is a field that has benefited from rapid and continuing advances in basic science and clinical applications. Nevertheless, in spite of constantly changing and expanding technology, the key to hematologic diagnosis remains most often the morphologic evaluation of the peripheral blood and bone marrow. New students and laboratory workers in hematology who will deal with these problems, whether at the bench or in the clinical setting, need to acquire the appropriate level of familiarity with normal and abnormal forms and with the range and variety of disease processes that may be reflected in hematologic reactions.

We have tried to make the coverage of topics in this atlas sufficiently comprehensive to be useful to students at various levels and those engaged in the practice of hematology. We have deliberately utilized a variety of preparations including smears and sections, special stains, histochemical and cytochemical techniques, and immunoperoxidase staining, and have stressed the importance of correlating findings in the peripheral blood with those in the bone marrow. Disorders of the spleen and lymph nodes, although closely related in many instances, are outside the scope of this work. The textual material is designed to supply basic information in a concise form; advanced students will want to supplement it with one or more of the large textbooks of hematology.

We owe a great debt of gratitude to those who have helped make this book possible. We wish to acknowledge the contributions of hematologic specimens made by Drs. Koichi Maeda, Robert W. McKenna, George C. Hoffman, Arkadi M. Rywlin, C. Francis Varga, Vincent Galdi and Stephen Van D. Chandler. The pictures have been selected from photomicrographs taken over a period of years by our residents, fellows and colleagues, too numerous to name. Mr. In Hwan Jo and Ms. Young Yea Ahn skillfully and painstakingly prepared the color prints. Mrs. Cres A. Martinez provided superior secretarial and organizational help. Mr. John Gardner and Ms. Lila G. Maron of Igaku-Shoin were unfailingly patient, encouraging and supportive.

Bong Hak Hyun Gene L. Gulati John K. Ashton

## Contents

Preface v

HEMATOLOGIC DISORDERS 1						
Chapter 1	Hematopoiesis 3 Mechanism of Hematopoiesis 4 Erythropoiesis 5 Granulopoiesis and Production of Monocytes 5 Lymphopoiesis and Production of Plasma Cells 7 Thrombopoiesis 8					
Chapter 2	Collection of Blood 11 Venipuncture 12 Capillary Puncture 15 Anticoagulants 16					
Chapter 3	Preparation of Peripheral Blood Films and Bone Marrow Films and Imprints 19  Manual Procedures 20  Automated Procedures 23  Preparation of Bone Marrow Aspirate Films 27  Preparation of Bone Marrow Biopsy Imprints 27					
Chapter 4	Routine Staining of Blood and Bone Marrow Films and Imprints 29 Wright's Stain 30 Wright-Giemsa Stain 35					
Chapter 5	Routine Examination of Blood and Bone Marrow Films and Imprints 39 Blood Film 40 Bone Marrow Films and Imprints 42					
Chapter 6	Special Stains in Hematology 45 Prussian Blue Stain for Iron 46					

PART I STRUCTURE, FUNCTION, TECHNIQUES, AND

	Leukocyte Alkaline Phosphatase Stain (LAP) 48 Peroxidase Stain (Modified Sigma Kit Method) 50 Sudan Black B Stain (Modified Sigma Kit Method) 52 Staining for Esterases 54 Specific Esterase or Naphthol AS-D Chloroacetate Esterase Nonspecific Esterase or α-Naphthyl Acetate Esterase Periodic Acid-Schiff Stain (PAS) 58 Tartrate-Resistant Acid Phosphatase Stain (Sigma Kit Method) 60							
	Staining for Fetal Hemoglobin (Kleihauer-Betke Method) New Methylene Blue Stain for Reticulocytes 64 Crystal Violet Stain for Heinz Bodies 66							
Chapter 7	Hematolo	gic Disorders 73						
	Red Cell Disorders 74							
White Cell Disorders 82								
Platelet Disorders 92								
PART II	COLOR	PLATES AND DESCRIPTIONS 105						
1	General:	The Peripheral Blood and Bone Marrow 107						
	Plate 1	Examination of the Peripheral Blood Smear 108						
	Plate 2	Examination of Bone Marrow Preparations 110						
	Plate 3	Bone Marrow, Normal Structures 112						
	Plate 4	Mitoses 114						
	Plate 5	The Red Cell Series, Normal 116						
	Plate 6	The Granulocyte-Monocyte Series 118						
	Plate 7	Lymphocytes, Plasma Cells and Lymphoid Follicles 120						
	Plate 8	Plasma Cells, Normal and Abnormal Variations 122						
	Plate 9	The Megakaryocytic Series 124						
	Plate 10	Osteoblasts, Osteoclasts, Tissue Granulocytes 126						
	Plate 11	Histiocytes, Endothelial Cells, Fat Cells 128						
	Plate 12	Iron-Stained Preparations 130						
П	The Red	Cell Series 133						
	Plate 1	Abnormal Red Cells 134						
	Plate 2	Abnormal Red Cells 136						
	Plate 3	Anemias: Iron Deficiency, Combined Iron and Folic						
		Acid Deficiency 138						
	Plate 4	Lead Poisoning, G-6-PD, Pyruvate Kinase and Py-						
		rimidine-5'-Nucleotidase Deficiencies 140						
	Plate 5	Megaloblastic Anemia 142						
	Plate 6	Megaloblastic Anemia 144						
	Plate 7	Sideroblastic Anemia 146						
	Diota V	Anlastic (Hypoplastic) Anemias 148						

Plate 9	Autoimmune Hemolytic Anemia 150				
Plate 10	Hemolytic Anemias 152				
Plate 11	Hemolytic Anemias (Cold Agglutinins) 154				
Plate 12	Hereditary Spherocytosis and Hereditary Elliptocy-				
	tosis 156				
Plate 13	Anemias, Miscellaneous 158				
Plate 14	Hemoglobinopathies 160				
Plate 15	Thalassemias 162				
Plate 16	Dyserythropoiesis 164				
Plate 17	Polycythemia Vera 166				
Plate 18	Polycythemia Vera, Secondary Polycythemias and				
	Stress Polycythemia 168				
TTL - 3371-14	Call Carter 181				
	te Cell Series 171				
Plate 1	Infections, Leukemoid Reaction 172				
Plate 2 Plate 3	Lymphocytosis 174 Reactive Plasmacytosis 176				
Plate 3 Plate 4					
riate 4	Angioimmunoblastic Lymphadenopathy, Eosinophilia, Chloramphenicol and Ethanol Toxicity 178				
Plate 5	Agranulocytosis, Hypersplenism, LE Prepara-				
Tiate 3	tion 180				
Plate 6	Leukocyte Anomalies 182				
Plate 7	Leukocyte Anomalies 184				
Plate 8	Cytochemistry 186				
Plate 9	Cytochemistry 188				
Plate 10	Acute Granulocytic Leukemia (FAB:M1 and				
	M2) 190				
Plate 11	Granulocytic Sarcoma, Acute Promyelocytic Leukemia				
	(FAB:M3) <b>192</b>				
Plate 12	Acute Myelomonocytic Leukemia (FAB:M4) and Acute				
energy and	Monocytic Leukemia (FAB:M5) 194				
Plate 13	,				
D1	FAB:M6) 196				
Plate 14	Chronic Granulocytic Leukemia 198				
Plate 15	Chronic Granulocytic Leukemia, Chronic My-				
DL 4 . 16	elomonocytic Leukemia 200				
Plate 16	Myeloproliferative Disorders, Myelofibrosis 202				
Plate 17	Myeloproliferative Disorders, Myelofibrosis, Osteo-				
Dieta 19	sclerosis 204  Myoloproliforetiyo Disorders, Idiopothic (Essential Pri				
Plate 18	Myeloproliferative Disorders, Idiopathic (Essential, Primary) Thrombocythemia 206				
Plate 19	Dysmyelopoietic Syndrome 208				
Plate 20	Eosinophilic, Basophilic, Megakaryocytic and Con-				
Tate 20	genital Leukemias 210				
Plate 21	Systemic Mast Cell Disease (Systemic Mastocy-				
11110 21	tosis) 212				

Ш

IV

Plate 22	Acute Lymphocytic Leukemia 214
Plate 23	Acute Lymphocytic Leukemia 216
Plate 24	Chronic Lymphocytic Leukemia 218
Plate 25	T-Cell Chronic Lymphocytic Leukemia, Prolymphocy-
	toid Transformation, Prolymphocytic Leukemia and
	Richter's Syndrome 220
Plate 26	Hairy Cell Leukemia, Sézary Syndrome 222
Plate 27	Malignant Lymphomas 224
Plate 28	Malignant Lymphomas 226
Plate 29	Malignant Lymphomas 228
Plate 30	Virus-Associated Hemophagocytic Syndrome, Malig-
	nant Histiocytosis 230
Plate 31	Plasma Cell Myeloma 232
Plate 32	Plasma Cell Myeloma 234
Plate 33	Macroglobulinemia, Cryoproteinemia, Amy-
	loidosis 236
_	akaryocytic Series 239
Plate 1	Reactive Thrombocytosis, Thrombocytopenias 240
Plate 2	Platelets, Miscellaneous 242
Missellon	eous Lesions 245
Plate 1	Metastatic Neoplasms 246
Plate 2	Metastatic Neoplasms 248
Plate 3	Granulomas 250
Plate 4	Granulomas 252
Plate 5	Lipidoses, Histiocytoses 254
Plate 6	Histiocytoses, Thorotrast, Cystinosis 256
Plate 7	Bone Changes, Miscellaneous Conditions 258
Plate 8	Necrobiosis, Fat Necrosis, Infarction 260
Plate 9	Parasites, Fungi 262
Plate 10	Parasites, Fungi, Bacteria, Plastic Sections 264
- 10.0 10	and services, a single, services, a mone services
Indev	267

## PART I

## STRUCTURE, FUNCTION, TECHNIQUES, AND HEMATOLOGIC DISORDERS

## Chapter 1

Hematopoiesis

Blood, defined as intravascular body fluid, consists of red cells (erythrocytes), white cells (leukocytes) and platelets (thrombocytes) suspended in plasma. Normally the red cells and platelets form relatively uniform populations, whereas the white cells comprise several distinct cell lines, namely, the granulocytes (neutrophils, eosinophils and basophils), lymphocytes and monocytes. Blood cells are released into the circulation after their production in the so-called hematopoietic sites in the body.

The process of blood cell production (hematopoiesis) begins early in embryonic life, being first evident in the yolk sac in the first lunar month. The cells produced by the yolk sac are primarily erythroid precursors containing embryonic hemoglobins. By the end of the second lunar month, the activity of the yolk sac ceases with the migration of hematopoietic stem cells to the liver, which starts producing different lines of blood cells. The hepatic hematopoietic activity is at its maximum during the second trimester of pregnancy, declining gradually thereafter to none or an insignificant level by the end of the antenatal period. The spleen's contribution to hematopoiesis is slight and is generally limited to the period of the third through the sixth lunar months. The bone marrow begins to produce blood cells in the fifth lunar month, at which time lymphoid tissues such as the thymus and lymph nodes also start producing lymphocytes. The bone marrow activity continually increases, and at the end of antenatal life, the bone marrow is 100% cellular. At birth and throughout postnatal life, the bone marrow remains the primary supplier of multipotent stem cells and the only site of hematopoiesis other than lymphoid tissues, which contribute lymphocytes and plasma cells. The liver and the spleen, however, maintain the potential for hematopoiesis throughout life. Although at birth the marrow of different bones is considered to be 100% hematopoietic (red), there are continual changes in the distribution of hematopoietic tissue within the bones throughout life. Fatty tissue (yellow marrow) begins to replace red marrow from the periphery of the body toward the axial skeleton, relatively rapidly in some bones (tibia, femur) and gradually in others (sternum, vertebrae and iliac bones). In the adult, most hematopoietic tissue is present in the vertebrae, ribs, sternum and the pelvic bones.

### **Mechanism of Hematopoiesis**

The undifferentiated reticular cells (also known as uncommitted stem cells) in the bone marrow undergo proliferation (mitotic division) and differentiation for self-renewal and for the production of stem cells committed to generate blood cells (hematopoietic stem cells), lymphocytes (lymphoid stem cells), and the connective tissue cells including fibroblasts, osteoblasts, chondrocytes, lipocytes, and endothelial cells.

The hematopoietic stem cells (hemocytoblasts), though morphologically unrecognizable by current methodology, have been identified functionally and named the "colony forming units-spleen" (CFU-S) based on the assay system used. Further proliferation and differentiation of the multipotent hematopoietic stem cells is essential to self-replication and the development of morphologically recognizable unipotent progenitors committed to the production of red cells, white cells including granulocytes and monocytes, and platelets. Similarly, the lymphoid stem cells undergo proliferation and differentiation or maturation to renew their own population and to produce lymphocytes and plasma cells. The production of

differentiated and functionally mature blood cells from their stem cells occurs through various stages of development in a sequential fashion, as described below.

## **Erythropoiesis**

The hematopoietic stem cell under the influence of a humoral factor, erythropoietin, differentiates first into a cell functionally designated as "erythroid colony forming cell" (E-CFC) and then into the pronormoblast (rubriblast), the earliest morphologically recognizable progenitor committed to produce red blood cells. Each pronormoblast undergoes a series of heteromorphogenic mitotic divisions through the stages of early and late basophilic normoblasts (prorubricytes), ultimately producing a total of 16 polychromatophilic normoblasts (rubricytes). With continued maturation without further division, each polychromatophilic normoblast differentiates sequentially into an orthochromic normoblast (metarubricyte), reticulocyte and mature erythrocyte. As the cell divides its size becomes smaller, and as the maturation proceeds, the nucleoli disappear and the nucleus becomes smaller, denser and pyknotic until it is extruded from the orthochromic normoblast. The extruded nucleus is phagocytosed and digested by macrophages. The cytoplasm undergoes changes in its staining characteristics from blue (due to RNA content) in the pronormoblast and basophilic normoblast stages to varying degrees of pink mixed with a bluish tinge in the polychromatophilic normoblast, orthochromic normoblast and reticulocyte, to completely pink in the mature erythrocyte, as a result of the accumulation of hemoglobin.

The synthesis of hemoglobin, though not evident until the polychromatophilic normoblast stage in Wright-stained blood films, probably starts in the late basophilic normoblast stage and continues through the stage of the reticulocyte. The production of reticulocytes from the pronormoblasts takes only about three to five days. The reticulocytes remain in the bone marrow for one or two days prior to being released into the circulation, where they lose their cytoplasmic reticulum and become mature red cells, also within one or two days. The mature red cells in the form of biconcave discs normally remain in the circulation for 120 days, carrying out their sole function of the transport and protection of the oxygencarrying pigment, hemoglobin. The red cells are lost from the circulation by senescence. Under usual steady-state conditions the daily loss of about 1% of circulating red cells is balanced by the production and release of an equivalent number of new red cells. Among several factors (hormones, minerals and vitamins) known to influence the rate of red cell production is the humoral factor, erythropoietin, which is produced mainly but not exclusively by the kidneys. It is considered to be the major regulator of erythropoiesis. The serum erythropoietin level increases as a response to decrease in the oxygen tension of tissues.

## **Granulopoiesis and Production of Monocytes**

The hematopoietic stem cell in the bone marrow under the influence of colony stimulating factor (CSF), a humoral factor apparently produced by monocytemacrophages, differentiates first into a cell functionally defined as a "granulocytemonocyte colony forming cell" (GM-CFC) and then into a cell morphologically identifiable as a myeloblast. Both of these cells, the GM-CFC and the myeloblast,

are committed to producing granulocytes and monocytes. Each myeloblast, presumably under the action of the CSF (also known as granulopoietin), divides and differentiates into two promyelocytes, each of which multiplies and differentiates into two myelocytes. At this stage two or three additional mitotic divisions result in self-replication of myelocytes, and the simultaneous development of specific granulation permits classification of myelocytes into three cell lines, neutrophilic, eosinophilic and basophilic. Further development of granulocytic cells occurs through progressive maturation only, leading to the formation of metamyelocytes, bands and mature segmented cells. Neutrophils, eosinophils and basophils are the mature end products of the maturation of neutrophilic, eosinophilic and basophilic myelocytes, respectively. With each mitotic division between promyelocyte and the late myelocyte stage, the cell size is reduced as are the nuclear size and the number of primary (azurophilic) granules. The promyelocyte is usually larger than the myeloblast and contains the most numerous primary granules, which represent membrane-bound organelles containing mainly lysosomal enzymes (acid hydrolases and peroxidase) with a small amount of muramidase. As the maturation sequence proceeds from the myeloblast to the myelocyte stage, the number of nucleoli decreases until their complete disappearance, the nuclear chromatin becomes progressively more clumped, and the cytoplasm loses its basophilia and acquires pink staining characteristics. The specific (secondary) granules seen in the myelocyte and later stages are also membrane-bound organelles and contain muramidase, collagenase, lactoferrin, phagocytin, alkaline phosphatase and several other enzymes. The maturation of myelocyte to metamyelocyte and band is characterized by progressive condensation of the chromatin and the indentation of the nucleus. Segmentation of the nucleus ultimately results in the formation of mature granulocytes. It takes about 14 days from the myeloblast stage to the release of granulocytes from the marrow into the circulation. Upon entering the circulation, the neutrophils move freely between a circulating granulocyte pool (CGP), which is reflected in the usual total and differential leukocyte count, and a marginal granulocyte pool (MGP), which is marginated along the walls of capillaries and venules. The MGP constitutes about 50% of the total body granulocyte pool and represents a large reserve readily available to the circulating blood upon demand. The estimated average time that neutrophils remain in the blood is six to eight hours, the total life span (in the blood and tissues) being nine to ten days.

There is experimental evidence to suggest that eosinophils may be derived from a progenitor (eosinophilic colony forming cell, EO-CFC) separate from the GM-CFC, under the influence also of a humoral factor (eosinophil colony stimulating factor, EO-CSF) distinct from CSF or granulopoietin. It has also been shown that the EO-CSF, unlike the CSF which is produced by monocyte-macrophages, is produced by lymphocytes when stimulated by pokeweed mitogen or mercaptoethanol.

Though the evidence is lacking, the possibility of a distinct progenitor for basophil production has also been suggested. The eosinophilic granules are similar to the neutrophilic granules in that they are also rich in peroxidase and in lysosomal enzymes with the exception of muramidase. The basophil granules are known to contain heparin, histamine and hyaluronic acid, but generally lack or may have minimal amounts of hydrolytic enzymes (acid phosphatase, alkaline phosphatase). The evidence for the presence of peroxidase in the basophils is controversial. Crystalloid protein structures (Charcot-Leyden crystals) have been shown by electron microscopy within the granules of eosinophils and after de-

granulation in the basophils. Eosinophils and basophils, like neutrophils, remain in the blood only for a few hours and then migrate to the tissues.

Neutrophils are attracted (chemotaxis) to the sites of tissue invasion by microorganisms. These cells are capable of phagocytosis, killing and digesting of bacteria and yeasts. The eosinophils, besides providing some defense against parasitic infestations, are involved in certain hypersensitivity reactions. They may also participate in the phagocytosis and destruction of microorganisms, but do so less readily than neutrophils. The function of basophils is less well understood. They are involved in immediate hypersensitivity reactions such as asthma as well as in delayed hypersensitivity reactions such as contact allergies.

As pointed out earlier, monocytes and neutrophils share a common progenitor, the GM-CFC. The production of monocytes from the GM-CFC proceeds sequentially through the stages of myeloblast, promonocyte and monocyte by way of several mitotic divisions and the process of maturation. The promonocyte, like the promyelocyte, is somewhat larger than the myeloblast. The monocyte, being only slightly smaller than the promonocyte, may also be larger than the myeloblast. As maturation proceeds from the myeloblast to the monocyte stage, the nuclear chromatin becomes progressively but only slightly denser, nucleoli decrease in number and ultimately disappear, and the staining characteristics of the cytoplasm change from blue to blue-gray and then to gray with fine azurophilic granules evident in the promonocyte and the monocyte stages. Upon release from the bone marrow, monocytes are also believed to be distributed between a circulating monocyte pool (CMP) and a marginal monocyte pool (MMP) in a ratio of 1:3.5. The monocytes circulate in the blood also for a short period (16–36 hours) and then migrate to various tissues, where they transform into macrophages or histiocytes. The life span of macrophages is perhaps in the range of several months or even a few years. Mononuclear phagocytes are involved in the disposal of microorganisms, senescent cells and foreign matter, besides participating in the immune response by mechanisms not yet fully elucidated. It is believed that they play a necessary role in the processing of antigens and their recognition by lymphocytes, and act as effector cells ("stimulated macrophages") in certain immune reactions.

Whereas the exact mechanism regulating the production of granulocytes in vivo remains poorly understood, there is a widely held concept that GM-CSF, a glycoprotein derived from monocyte-macrophages, is the chief mediator of proliferation and differentiation of the cells of the granulocytic and monocytic series.

## Lymphopoiesis and Production of Plasma Cells

Lymphoid stem cells in the bone marrow proliferate to self-replicate and to export cells to the thymus (central lymphoid organ) and the peripheral lymphoid tissues—lymph nodes, spleen, gut, tonsils, blood and lymph. The cells that pass through the thymus acquire certain antigenic characteristics and become prothymus cells (pro-T cells), which ultimately mature to become T cells with varied functional attributes, e.g., helper T cells and suppressor T cells. Some of the cells from the thymus also migrate to the peripheral lymphoid tissues, there to proliferate and populate these tissues with immunocompetent T cells. On the other hand, some of the lymphoid stem cells in the bone marrow divide and differentiate into pro-B cells and then into B cells in the marrow. Some of these lymphocytes