SYNTHESIS, STORAGE AND SECRETION OF ADRENAL CATECHOLAMINES

Editors: F. IZUMI M. OKA K. KUMAKURA

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Proceedings of a satellite symposium to the 8th International Congress of Pharmacology, Tokyo, July 1981

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PERGAMON PRESS

OXFORD · NEW YORK · TORONTO · SYDNEY · PARIS · FRANKFURT

U.K.

Pergamon Press Ltd., Headington Hill Hall,

Oxford OX3 0BW, England

U.S.A.

Pergamon Press Inc., Maxwell House, Fairview Park,

Elmsford, New York 10523, U.S.A.

CANADA

Pergamon Press Canada Ltd., Suite 104,

150 Consumers Rd., Willowdale, Ontario M2J 1P9, Canada

AUSTRALIA

Pergamon Press (Aust.) Pty. Ltd., P.O. Box 544,

Potts Point, N.S.W. 2011, Australia

FRANCE

Pergamon Press SARL, 24 rue des Ecoles,

75240 Paris, Cedex 05, France

FEDERAL REPUBLIC OF GERMANY

Pergamon Press GmbH, 6242 Kronberg-Taunus, Hammerweg 6, Federal Republic of Germany

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First edition 1982

Library of Congress Cataloging in Publication Data

Symthesis, storage & secretion of adrenal catecholamines.

(Advances in the biosciences; v. 36)

Catecholamines—Metabolism—congresses.

2. Adrenal medulla—Congresses. 3. Chromaffin cells-Congresses. I. Izumi, F. (Futoshi),

. II. Oka, M. (Motoo), 1931-III. Kumakura, K. IV. International Congress of Pharmacology (8th: 1981: Tokyo, Japan) V. Title: Synthesis, storage, and secretion of adrenal catecholamines. VI. Series [DNLM:

1. Catechnolamines—Congresses. 2. Adrenal medulla-Physiology-Congresses. W3 AD244 v. 36

82-520

1981 / WK 725 S993 1981]

QP801.C33S96 599.01'88 1982

British Library Cataloguing in Publication Data

Synthesis, storage & secretion of adrenal catecholamines. — (Advances in the biosciences;

1. Adrenergic mechanisms — Congresses

2. Catecholamines — Congresses

I. Izumi, F. II. Oka, M. III. Kumakura, K.

IV. Series

599.01'9245 QP365.5

ISBN 0-08-028012-9

In order to make this volume available as economically and as rapidly as possible the authors' typescripts have been reproduced in their original forms. This method unfortunately has its typographical limitations but it is hoped that they in no way distract the reader.

Printed in Great Britain by A. Wheaton & Co. Ltd., Exeter

PREFACE

Since the memorial observation by Oliver and Schäfer (1895) that suprarenal extracts contained cardioactive pressor substance, adrenal medulla have been the central subject in endocrinology. A variety of physiological concepts in cellular and synaptic events have been much developed from this organ. The role of catecholamines spread out to a number of autonomically innervated tissues and central nervous systems as well. Today, obviously, the adrenal medullary cells are the most distinctive prototype for neuronal and neurosecretory cells.

The major pathways for the synthesis and metabolism of catechol-amines were fully understood in cellular, subcellular and molecular levels. Substantial recognition of catecholamines has been made very clearly. At the same time, recent developments in biochemical, physiological and pharmacological techniques have enabled investigation of membrane receptors linked to the action of catecholamines. However, when we retrospect in the primary function of adrenal medulla, - the synthesis, storage and secretion of catecholamines-, we will realize the fact that functional recognitions of this organ are so rigid and fragmental.

Adrenal medullary cells are neural crest origin and their functions are wholly under the control of first order preganglionic fibers. Each function could be well studied in these cells, since they show the highest activities of catecholamine synthesizing enzymes, have the well developed catecholamine storing granules and readily secrete their contents upon stimulation of cholinergic receptor. And in fact, knowledges in separate function have been successfully accumulated to a great amount. But question arises whether the mere summation of each separate function could give the comprehensive figure of the adrenal medullary cells. What is the central event in the receptormediated control of cell functions? Is there any integrating mechanism in the functions of adrenal medulla?

It seemed, therefore, to be important to have a meeting for scientists working with different aspects of adrenal catecholamines and to seek for the fundamental and integrating mechanisms which function in adrenal medulla. The 8th International Congress of Pharmacology in Tokyo was an opportune time to have such meeting. The Satellite Symposium of Adrenal Catecholamine followed to the Congress and lasted for three days at Hakone. The proceeding of the meeting

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are presented in this book. We really hope that all the readers will recognize the present frontier of adrenal catecholamines and realize that adrenal medullary cells are unrivaled one in studies of neuroendocrinology as well as neurotransmitter mechanism.

Such a meeting would not have been possible without the help of many peoples. We are grateful for the help of M.Sandler, H.O.Viveros, E.Costa and C.Gagnon who worked with us on the Programme Committee. We wish to thank the University of Occupational and Environmental Health, Japan, School of Medicine for repeated encouragements. We also wish to thank J. Lavender for his efforts in bringing this book together. Last, but not least, we are grateful for the excellent secretarial help of Y. Toyohira, K. Okuda and K. Hanada.

Hakone, Japan, July 1981

- F. Izumi
- M. Oka
- K. Kumakura

ACKNOWLEDGEMENTS

The financial support of the following is gratefully acknowledged:

The Japan Medical Association

Ciba-Geigy, Japan
Dainippon Pharmaceutical Co., Ltd., Japan
Nakarai Chemicals, Ltd., Japan
Otsuka Pharmaceutical Co., Ltd., Japan
Toyo Jozo Co., Ltd., Japan
Yoshitomi Pharmaceutical Ind., Co., Ltd., Japan

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The Chromaffin Granule: A Challenge in the Past, Present and Future

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ABSTRACT

In this paper we will review the major aspects of research on chromaffin granules: isolation methods, chemical composition and molecular function. Our own recent studies on nucleotide transport and on the morphological substrate of the ${\rm Mg}^{2+}-{\rm ATP}-{\rm ase}$ will also be discussed.

KEYWORDS

Adrenal medulla; chromaffin granules; catecholamines; nucleotide transport; ATP-ase; exocytosis.

INTRODUCTION

In 1953 Blaschko and Welch and Hillarp, Lagerstedt and Nilson reported that centrifugation of homogenates of bovine adrenal medulla led to the sedimentation of the major part of the hormones of this organ. These authors rightly concluded that the medullary hormones, the catecholamines, are stored in subcellular organelles. This classical observation finally led to the isolation and detailed characterization of the catecholamine-storing vesicles of the adrenal medulla, the chromaffin granules.

Twenty-eight years of research on chromaffin granules have yielded numerous publications. In this paper we will review the major challenges which had to be met by the investigators in this field. Due to the limited space only a selected number of references will be given.

ISOLATION OF CHROMAFFIN GRANULES

One of the first problems which had to be tackled was the establishment of isolation methods for these cell organelles. As it can be seen from Table 1 methods employing linear or simplified sucrose den-

sity gradients were introduced first. These methods proved essential for the analysis of the composition of chromaffin granules. More recently isoosmotic gradients were described. Chromaffin granules purified by these methods could be incubated afterwards under isotonic conditions which was a prerequisite for various uptake studies.

TABLE 1 The Introduction of Isolation Methods for Chromaffin Granules in the Period from 1953 to 1981

```
DIFFERENTIAL CENTRIFUGATION: Blaschko and Welch (1953);
Hillarp and co-workers (1953)

SUCROSE DENSITY GRADIENT CENTRIFUGATION:
SEPARATION FROM MITOCHONDRIA: Blaschko and co-workers (1957)

SEPARATION FROM LYSOSOMES: Smith and Winkler (1966)
SIMPLIFIED GRADIENT: Smith and Winkler (1967a)

MILLIPORE FILTER: Oka and co-workers (1966)

ISOTONIC GRADIENTS: FICOLL: Trifaró and Dworkind (1970)
METRIZAMIDE: Morris and Schovanka (1977)
PERCOLL: Meyer and Burger (1979)
```

THE COMPOSITION OF CHROMAFFIN GRANULES

After the establishment of isolation methods considerable work was invested to study the composition of chromaffin granules (see Table 2). In the first period the major contributions were made by English and Swedish scientists. However, already in the sixties several new groups joined the field and quite a few of them are still active today. It seems unlikely that future studies will discover new granule components which are important from a quantitative point of view. However, as demonstrated by the recent finding of enkephalins, new components, which are minor in quantity, but have important functional properties, may still be found. Thus most of the protein components of chromaffin granules have not yet been assigned a function.

TABLE 2 Studies on the Composition of Chromaffin Granules in the Period from 1953 to 1981

```
CATECHOLAMINES: Blaschko and Welch (1953); Hillarp and co-workers (1953)

SOLUBLE PROTEINS: Hillarp (1958b)

Mg<sup>2+</sup>-ATP-ASE: Hillarp (1958a)

ADP, AMP: Hillarp and Thieme (1959)

PHOSPHOLIPIDS, CHOLESTEROL: Hillarp (1959)

CYTOCHROME b-561: Spiro and Ball (1961); Ichikawa and Yamano (1965)

DOPAMINE β-HYDROXYLASE: Oka and co-workers (1967)
```

The Chromaffin Granule

```
LYSOLECITHIN: Blaschko and co-workers (1967)
     Ca<sup>2+</sup>: Borowitz, Fuwa and Weiner (1965)
     CHROMOGRANIN A: Helle (1966); Smith and Winkler (1967b);
       Smith and Kirshner (1967)
1970
     GTP, UTP: Goetz, Da Prada and Pletscher (1971)
     NADH: acceptor oxidoreductase: Flatmark, Terland and Helle
       (1971)
     PHOSPHATIDYLINOSITOLKINASE: Buckley and co-workers (1971)
     CHROMOMEMBRIN B: Hörtnagl and co-workers (1971)
     (= CYTOCHROME b-561: Apps, Pryde and Phillips, 1980)
     MUCOPOLYSACCHARIDES: Fillion, Nosal and Uvnäs (1971); Margolis
       and Margolis (1973)
     ASCORBIC ACID: Terland and Flatmark (1975)
     ACTIN: Phillips and Slater (1975)
     GLYCOPROTEINS: Eagles and co-workers (1975); Huber and co-workers
       (1979)
     ACTININ: Jockusch and co-workers (1977)
     SYNAPTIN: Bock and Helle (1977)
     GANGLIOSIDES: Geissler and co-workers (1977); Dreyfus and co-
     ENKEPHALINS: Lundberg and co-workers (1979); Viveros and co-
      workers (1979)
1981
```

MOLECULAR FUNCTION OF CHROMAFFIN GRANULES

Molecular Mechanism of Catecholamine Storage

Since chromaffin granules contain an extremely high concentration of catecholamines the question of the mechanism of storage was already discussed by the first investigators. ATP was soon recognized as a possible partner in catecholamine storage (see Table 3). The claim that the membrane of chromaffin granules was freely permeable to catecholamines at 0°C together with the fact of the stability of the catecholamine store at this temperature led to the postulate of a stable storage complex involving catecholamines and ATP. This seemed finally established when such complexes involving these two components could be demonstrated under in vitro conditions. However, during the last five years this concept has changed. The impermeability of the membranes of chromaffin granules at 0° for small molecules like ions was established. Thus the stability of the catecholamine storage at this temperature could be explained by this property without postulating a storage complex. In fact the absence of a stable storage complex at this temperature was demonstrated by nuclear magnetic resonance studies (see Sen and Sharp, 1981). However, the absence of a stable storage complex does not exclude a certain amount of interaction between the granule components. Indeed, the nuclear magnetic resonance studies indicate that catecholamines and ATP, although free in solution within the granule, interact with each other and possibly also with the acidic chromogranins.

Since chromaffin granules do not contain a stable storage complex of catecholamines additional mechanisms must exist to maintain the high concentration of amines and nucleotides within these organelles.

TABLE 3 Studies on the Molecular Mechanism of Catecholamine Storage in the Period from 1953 to 1981

```
ATP: A STORAGE COMPONENT: Hillarp and co-workers (1955);
Blaschko and co-workers (1956)
o.6 M CONCENTRATION OF CATECHOLAMINES: Hillarp (1959)

GRANULE MEMBRANE FREELY PERMEABLE TO CATECHOLAMINES: Kirshner,
Holloway and Kamin (1966)
IN VITRO COMPLEXES OF CATECHOLAMINES AND ATP: Berneis, Pletscher and Da Prada (1969)

GRANULE MEMBRANES: LOW PERMEABILITY AT O°: Perlman (1976);
Johnson and Scarpa (1976)
NMR-STUDIES: Daniels, Williams and Wright (1978); Sen and co-workers (1979)
```

Molecular Mechanisms of Catecholamine and Nucleotide Uptake
References for the major breakthroughs in the field are given in
Table 4.

TABLE 4 Studies on Catecholamine and Nucleotide Uptake in the Period from 1962 to 1981

```
1962
     UPTAKE OF CATECHOLAMINES:
       IN INTACT GRANULES: Kirshner (1962); Carlsson and co-workers
         (1962)
       DEPENDENCE ON ATP-ASE: Banks (1965)
       IN GRANULE GHOSTS: Taugner (1971); Phillips (1974)
1970
       DRIVEN BY PROTON GRADIENT: Bashford and co-workers (1975)
       DRIVEN BY \triangle pH and \triangle \psi: Johnson and Scarpa (1979)
       RECONSTITUTION IN VITRO: Maron and co-workers (1979)
     ISOLATION OF ATP-ASE: Apps and Glover (1978)
     UPTAKE OF NUCLEOTIDES:
       CARRIER-MEDIATED TRANSPORT: Kostron and co-workers (1977)
       DRIVEN BY Au: Aberer and co-workers (1978); Weber and
         Winkler (1981)
1981 L
```

The uptake of both catecholamines and nucleotides is driven by a proton-gradient maintained by the proton pumping ATPase of the granule membrane. This enzyme, which has been isolated, was shown to be very similar to the proton pumping ATPase of mitochondria (Apps and Schatz, 1979). Based on these data we calculated (Winkler and Westhead, 1980) that a single granule should possess about 10 molecules of this enzyme. We have now found the morphological substrate of this enzyme by negative staining and by freeze-etching (Schmidt,

Winkler and Plattner, 1981). About 20 particles of 10 nm diameter are present on the outer surface of chromaffin granules. They resemble closely the stalked particles observed previously in mitochondria.

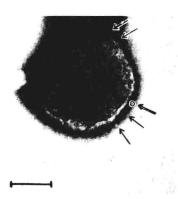


Fig. 1. Negative staining of chromaffin granules. Isolated chromaffin granules were subjected to negative staining (2 % ammonium heptamolybdate). The arrows indicate 10 nm particles on the surface of a chromaffin granule. The double arrow indicates a particle which has been encircled by ink for better demonstration. The bar represents 100 nm.

Recent studies on the nucleotide transport (Weber and Winkler, 1981, and unpublished observations) indicate that the carrier has a relatively broad specificity. In addition to ATP other nucleotides, e.g. UTP and GTP, are transported with similar affinity. However, even phosphoenolpyruvate, PO4 $^{3-}$ and SO4 $^{2-}$ are transported. On the other hand, acetate, glutamic acid or pyruvate have no affinity for the carrier.

Future studies will probably soon lead to the isolation of the carriers for catecholamines and nucleotides. The exact molecular transport of these compounds can then be established.

Molecular Mechanism of Catecholamine Release

Although exocytosis as the mechanism of catecholamine secretion was already established fifteen years ago (see Smith and Winkler, 1972 for references) the molecular mechanism of this process still eludes us. The only established fact is the involvement of calcium, whereas the role of contractile proteins (see Trifaró, 1978), of granule membrane methylation (Gagnon and Heisler, 1979), of phosphorylation (Amy and Kirshner, 1981), of lysolecithin (see Smith and Winkler, 1972) and of synexin (Creutz and co-workers, 1979) are still a matter of speculation.

Japanese scientists (Oka and co-workers, 1965) first showed that incubation of chromaffin granules with ATP-Mg²⁺ led to the release of catecholamines. It was speculated that this release reaction was involved in catecholamine secretion in vivo (Poisner and Trifaró, 1967; see also Pazoles and Pollard, 1978). However, experimental proof is still lacking and evidence not supporting this suggestion has recently been presented (Englert and Perlman, 1981). It will therefore be one of the greatest challenges in the adrenergic field

to elucidate the molecular mechanism of catecholamine secretion. This will not only be relevant for the adrenal medulla but also for other secretory systems where knowledge on this mechanism is still lacking.

ACKNOWLEDGEMENT

The original work of the authors quoted in this review was supported by the Dr.Legerlotz-Stiftung and by the Fonds zur Förderung der wissenschaftlichen Forschung.

REFERENCES

- Aberer, W., H. Kostron, E. Huber, and H. Winkler (1978). A characterization of the nucleotide uptake by chromaffin granules of bovine adrenal medulla. <u>Biochem.J.</u>, 172, 353-360.
- Amy, C. M., and N. Kirshner (1981). Phosphorylation of adrenal medulla cell proteins in conjunction with stimulation of catecholamine secretion. J. Neurochem., 36, 847-854.

 Apps, D. K., and L. A. Glover (1978). Isolation and characterization
- Apps, D. K., and L. A. Glover (1978). Isolation and characterization
 of magnesium adenosinetriphosphatase from the chromaffin granu le membrane. FEBS Letters, 85, 254-258.
 Apps, D. K., J. G. Pryde, and J. H. Phillips (1980) Cytochrome b-561
- Apps, D. K., J. G. Pryde, and J. H. Phillips (1980) Cytochrome b-561 is identical with chromomembrin B, a major polypeptide of chromaffin granule membranes. <u>Neuroscience</u>, <u>5</u>, 2279-2287.
 Apps, D. K., and G. Schatz (1979). An adenosine triphosphatase iso-
- Apps, D. K., and G. Schatz (1979). An adenosine triphosphatase isolated from chromaffin granule membranes is closely similar to F1-adenosine triphosphatase of mitochondria. <u>Eur. J. Biochem.</u>, 100, 411-419.
- Banks, P. (1965). The adenosine-triphosphatase activity of adrenal chromaffin granules. Biochem.J., 95, 490-496.
- Bashford, C. L., R. P. Casey, G. K. Radda, and G. A. Ritchie (1975). The effect of uncouplers on catecholamine incorporation by vesicles of chromaffin granules. Biochem.J., 148, 153-155.
- sicles of chromaffin granules. <u>Biochem.J.</u>, <u>148</u>, 153-155.

 Berneis, K. H., A. Pletscher, and M. Da Prada (1969). Metal-dependent aggregation of biogenic amines: a hypothesis for their storage and release. Nature, 224, 281-283.
- Blaschko, H., G. V. R. Born, A. D'Iorio, and N. R. Eade (1956). Observations on the distribution of catecholamines and adenosine-triphosphate in the bovine adrenal medulla. J. Physiol. Lond., 133, 548-557.
- Blaschko, H., H. Firemark, A. D. Smith, and H. Winkler (1967). Lipids of the adrenal medulla: lysolecithin, a characteristic constituent of chromaffin granules. Biochem. J., 104, 545-549.
- ent of chromaffin granules. <u>Biochem.J.</u>, <u>104</u>, 545-549.

 Blaschko, H., H. M. Hagen, and P. Hagen (1957). Mitochondrial enzymes and chromaffin granules. <u>J. Physiol. Lond.</u>, <u>138</u>, 316-322.

 Blaschko, H., and A. D. Welch (1953). Localization of adrenaline in
- Blaschko, H., and A. D. Welch (1953). Localization of adrenaline in cytoplasmic particles of the bovine adrenal medulla. Naunyn Schmiedebergs Arch. exp. Path. Pharmacol., 219, 17-22.
- Bock, E., and K. B. Helle (1977). Localization of synaptin on synaptic vesicle membranes, synaptosomal plasma membranes and chromaffin granule membrane. FEBS Letters, 82, 175-178.
- Borowitz, J. L., K. Fuwa, and N. Weiner (1965). Distribution of metals and catecholamines in bovine adrenal medulla subcellular fractions. Nature, 205, 42-43.

- Buckley, J. T., Y. A. Lefebvre, and J. N. Hawthorne (1971). Identification of an actively phosphorylated component of adrenal me-
- dulla chromaffin granules. <u>Biochim. biophys. Acta</u>, <u>239</u>, 517-519. Carlsson, A., N. A. Hillarp, and B. Waldeck (1962). A Mg²⁺ ATP dependent storage mechanism in the amine granules of the adrenal medulla. <u>Med. Exptl.</u>, <u>6</u>, 47-53.
- Creutz, C. E., C. J. Pazoles, and H. B. Pollard (1979). Self-association of synexin in the presence of calcium. J. Biol. Chem., 254, 553-558.
- Daniels, A. J., R. J. P. Williams, and P. E. Wright (1978). The character of the stored molecules in chromaffin granules of the adrenal medulla: a nuclear magnetic resonance study. Neuroscien-<u>ce,</u> <u>3</u>, 573-585.
- Dreyfus, H., D. Aunis, S. Harth, and P. Mandel (1977) Gangliosides and phospholipids of the membranes from bovine adrenal medullary chromaffin granules. Biochim. biophys. Acta, 489, 89-97.
- Eagles, P. A. M., L. N. Johnson, and C. Van Horn (1975). The distribution of concanavalin A receptor sites on the membrane of chromaffin granules. J. Cell Sci., 19, 33-54.
- Englert, D. F., and R. L. Perlman (1981). Permeant anions are not required for norepinephrine secretion from pheochromocytoma
- cells. Biochim. biophys. Acta, 674, 136-143. Fillion, F. G., R. Nosal, and B. Uvnäs (1971). The presence of a sulphomucopolysaccharide-protein complex in adrenal medullary cell
- granules. Acta physiol. scand., 84, 286-288.
 Flatmark, T., O. Terland, and K. B. Helle (1971). Electron carriers of the bovine adrenal chromaffin granules. Biochim. biophys. Acta, 226, 9-19.
- Gagnon, C., and S. Heisler (1979). Protein-methylation: role in exo-
- cytosis and chemotaxis. <u>Life Sciences</u>, <u>25</u>, 993-1000. Geissler, D., A. Martinek, R. U. Margolis, R. K. Margolis, J. A. Skrivanek, R. Ledeen, P. König, and H. Winkler (1977).Composition and biogenesis of complex carbohydrates of ox adrenal chromaffin granules. Neuroscience, 2, 685-693.
- Goetz, U., M. Da Prada, and A. Pletscher (1971). Adenine-, guanineand uridine 5'-phosphonucleotides in blood platelets and storage organelles of various species. J. Pharm. exp. Ther., 178, 210-215.
- Helle, K.B. (1966). Some chemical and physical properties of the soluble protein fraction of bovine adrenal chromaffin granules. Molec. Pharmac., 2, 298-310.
- Hillarp, N. R. (1958a). Enzymatic systems involving adenosinephosphates in the adrenaline and noradrenaline containing granules of the adrenal medulla. Acta physiol. scand., 42, 144-165.
- Hillarp, N. A. (1958b). Isolation of some biochemical properties of the catecholamine granules in the cow adrenal medulla. Acta physiol. scand., 43, 82-96.
 Hillarp, N. R. (1959) Further observations of the state of the cate-
- cholamines stored in the adrenal medullary granules. Acta physiol. scand., 47, 271-279.
- Hillarp, N. R., S. Lagerstedt, and B. Nilson (1953) The isolation of granular fraction from the suprarenal medulla containing the sympathomimetic amines. Acta physiol. scand., 29, 251-263. Hillarp, N. A., B. Nilson, and B. Högberg (1955). Adenosine triphos-
- phate in the adrenal medulla of the cow. Nature, 176,1032-1033.
- Hillarp, N. A., and G. Thieme (1959). Nucleotides in the catecholamine granules of the adrenal medulla. Acta physiol. scand., 45, 328-338.

- Hörtnagl, H., H. Winkler, J. A. L. Schöpf, and W. Hohenwallner (1971) Membranes of chromaffin granules. Isolation and partial charac-
- terization of two proteins. <u>Biochem.J.</u>, <u>122</u>, 299-304. Huber, E., P. König, G. Schuler, W. Aberer, H. Plattner, and H. Winkler (1979). Characterization and topography of the glycoproteins
- of adrenal chromaffin granules. <u>J. Neurochem.</u>, <u>32</u>, 35-47. Ichikawa, Y. and T. Yamano (1965). Cytochrome b-561 in the microsomes of the adrenal medulla. Biochem.biophys.Res.Commun., 20,263-268.
- Jockusch, B. M., M. M. Burger, M. Da Prada, J. G. Richards, C. Chaponnier, and G. Gabbiani (1977). < -actinin attached to membranes of secretory vesicles. Nature, 270, 628-629.
- Johnson, R. G. and A. Scarpa (1976). Ion permeability of isolated chromaffin granules. J. gen. Physiol., 68, 601-631.
- Johnson, R. G. and A. Scarpa (1979). Protonmotive force and catecholamine transport in isolated chromaffin granules. J. Biol. Chem., 254, 3750-3760.
- Kirshner, N. (1962). Uptake of catecholamines by a particulate frac-
- tion of the adrenal medulla. <u>J. biol. Chem.</u>, 237, 2311-2317.

 Kirshner, N., C. Holloway, and D. L. Kamin (1966). Permeability of catecholamine granules. <u>Biochim. biophys. Acta</u>, 112, 532-537.

 Kostron, H., H. Winkler, L. J. Peer, and P. König (1977) Uptake of adenosine triphosphate by isolated adrenal chromaffin granules: A carrier-mediated transport. Neuroscience, 2, 159-166.
- Lundberg, J. M., B. Hamberger, M. Schultzberg, T. Hökfelt, P. O. Granberg, S. Efendic, L. Terenius, M. Goldstein, and R. Luft (1979) Enkephalin- and somatostatin-like immunoreactivities in human adrenal medulla and pheochromocytoma. Proc. Natl. Acad. Sci. USA, 76, 4079-4083.
- Margolis, R. U., and R. K. Margolis (1973) Isolation of chondroitin sulfate and glycopeptides from chromaffin granules of adrenal
- medulla. <u>Biochem. Pharmac.</u>, <u>22</u>, 2195-2197.

 Maron, R., H. Fishkes, B. I. Kanner, and S. Schuldiner (1979) Solubilization and reconstitution of the catecholamine transporter from bovine chromaffin granules. Biochemistry, 18,4781-4785.
- Meyer, D. I. and M. M. Burger (1979). Isolation of a protein from the plasma membrane of adrenal medulla which binds to secretory vesicles. J. biol. Chem., 254, 9854-9859.
- Morris, S. J., and I. Schovanka (1977). Some physical properties of adrenal medulla chromaffin granules isolated by a new continuous iso-osmotic density gradient method. Biochim. biophys. Acta, 464,
- Oka, N., K. Kajikawa, T. Ohuchi, H. Yoshida, and R. Imaizumi (1967). Distribution of dopamine &-hydroxylase in subcellular fractions
- of adrenal medulla. <u>Life Sciences</u>, <u>6</u>, 461-465. Oka, M., T. Ohuchi, H. Yoshida, and R. Imaizumi (1965). Effect of adenosine triphosphate and magnesium on the release of catecholamines from adrenal medullary granules. Biochim. biophys. Acta, 97, 170-171.
- Oka, M., T. Ohuchi, H. Yoshida, and R. Imaizumi (1966). The isolation of catecholamine storage granules from adrenal medulla by the membrane filter technique. Life Sciences, 5, 427-432.
- Pazoles, C.J. and H. B. Pollard (1978). Evidence for stimulation of anion transport in ATP-evoked transmitter release from isolated secretory vesicles. J. Biol. Chem., 253, 3962-3969.
- Perlman, R. L. (1976). The permeability of chromaffin granules to non-electrolytes. Biochem. Pharmacol., 25, 1035-1038.
- Phillips, J. H. (1974). Transport of catecholamines by resealed chromaffin granule ghosts. Biochem. J., 144, 311-318.