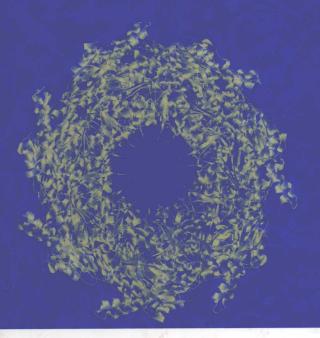


WILEY-INTERSCIENCE SERIES ON MASS SPECTROMETRY

Series Editors: Dominic M. Desiderio and Nico M. Nibbering

Principles of MASS SPECTROMETRY APPLIED TO BIOMOLECULES



EDITED BY JULIA LASKIN AND CHAVA LIFSHITZ



PRINCIPLES OF MASS SPECTROMETRY APPLIED TO BIOMOLECULES

Edited by

JULIA LASKIN, PhD

Pacific Northwest National Laboratory Richland, Washington

CHAVA LIFSHITZ, PhD

The Hebrew University Jerusalem, Israel





Copyright © 2006 by John Wiley & Sons, Inc. All rights reserved

Published by John Wiley & Sons, Inc., Hoboken, New Jersey Published simultaneously in Canada

No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording, scanning, or otherwise, except as permitted under Section 107 or 108 of the 1976 United States Copyright Act, without either the prior written permission of the Publisher, or authorization through payment of the appropriate per-copy fee to the Copyright Clearance Center, Inc., 222 Rosewood Drive, Danvers, MA 01923, (978) 750-8400, fax (978) 750-4470, or on the web at www.copyright.com. Requests to the Publisher for permission should be addressed to the Permissions Department, John Wiley & Sons, Inc., 111 River Street, Hoboken, NJ 07030, (201) 748-6011, fax (201) 748-6008, or online at http://www.wiley.com/go/permission.

Limit of Liability/Disclaimer of Warranty: While the publisher and author have used their best efforts in preparing this book, they make no representations or warranties with respect to the accuracy or completeness of the contents of this book and specifically disclaim any implied warranties of merchantability or fitness for a particular purpose. No warranty may be created or extended by sales representatives or written sales materials. The advice and strategies contained herein may not be suitable for your situation. You should consult with a professional where appropriate. Neither the publisher nor author shall be liable for any loss of profit or any other commercial damages, including but not limited to special, incidental, consequential, or other damages.

For general information on our other products and services or for technical support, please contact our Customer Care Department within the United States at (800) 762-2974, outside the United States at (317) 572-3993 or fax (317) 572-4002.

Wiley also publishes its books in a variety of electronic formats. Some content that appears in print may not be available in electronic formats. For more information about Wiley products, visit our web site at www.wiley.com.

Library of Congress Cataloging-in-Publication Data:

Principles of mass spectrometry applied to biomolecules/edited by Julia Laskin, Chava Lifshitz. p. cm.

Includes bibliographical references and index.

ISBN-13 978-0-471-72184-0 (cloth)

ISBN-10 0-471-72184-0 (cloth)

1. Mass spectrometry. 2. Biomolecules–Analysis. I. Laskin, Julia, 1967-II. Lifshitz, Chava.

QP519.9.M3P77 2006

543'.65-dc22 2006043900

Printed in the United States of America

10 9 8 7 6 5 4 3 2 1

CONTRIBUTORS

- **Richard L. Beardsley**, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- **Kathrin Breuker**, Institute of Organic Chemistry and Center for Molecular Biosciences Innsbruck (CMBI), University of Innsbruck, Innrain 52a, A-6020 Innsbruck, Austria
- **Guilong Cheng**, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- R. Graham Cooks, Department of Chemistry, Purdue University, 560 Oval Drive, West Lafayette, IN 47907-2038
- Robert C. Dunbar, Chemistry Department, Case Western Reserve University, Cleveland, OH 44106
- You-Jun-Fu, Department of Physics, Washington State University, 2710 University Drive, Richland, WA 99352; W. R. Wiley Environmental Molecular Sciences Laboratory and Chemical Sciences Division, Pacific Northwest National Laboratory, MS K8-88, P.O. Box 999, Richland, WA 99352
- **R. Benny Gerber**, Department of Chemistry, University of California, Irvine, CA 92697; Department of Physical Chemistry and the Fritz Haber Research Center, The Hebrew University, Jerusalem 91904, Israel
- Markus Gerhards, Heinrich-Heine Universität Düsseldorf, Institut für Physikalische Chemie I, Universitätstrasse 26.33.O2, 40225 Düsseldorf, Germany

- **Bogdan Gologan**, Department of Chemistry, Purdue University, 560 Oval Drive, West Lafayette, IN 47907-2038
- M. Kirk Green, McMaster Regional Centre for Mass Spectrometry, Department of Chemistry, McMaster University, Hamilton, Canada
- William L. Hase, Department of Chemistry and Biochemistry, Texas Tech University, Lubbock, TX 79409-1061
- **Kristin A. Herrmann**, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- Amy E. Hilderbrand, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- Alan C. Hopkinson, Centre for Research in Mass Spectrometry and the Department of Chemistry, York University, 4700 Keele Street, Toronto, Ontario, Canada M3J 1P3
- Julia Laskin, Fundamental Sciences Division, Pacific Northwest National Laboratory, P.O. Box 999 K8-88, Richland, WA 99352
- Carlito B. Lebrilla, Department of Chemistry, University of California, Davis, CA 95616
- Chava Lifshitz, Department of Physical Chemistry and The Farkas Center for Light Induced Processes, The Hebrew University of Jerusalem, Jerusalem 91904, Israel
- Scott A. McLuckey, Department of Chemistry, Purdue University, 560 Oval Drive, West Lafayette, IN 47907-2084
- **Samy O. Meroueh**, Department of Chemistry and Biochemistry, University of Notre Dame, Notre Dame, IN 46556-5670
- **Asif Rahaman**, Department of Chemistry and Biochemistry, Texas Tech University, Lubbock, TX 79409-1061
- **Carol V. Robinson**, The University Chemical Laboratory, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, United Kingdom
- **Dorit Shemesh**, Department of Physical Chemistry and The Fritz Haber Research Center, The Hebrew University, Jerusalem 91904, Israel
- K. W. Michael Siu, Centre for Research in Mass Spectrometry and the Department of Chemistry, York University, 4700 Keele Street, Toronto, Ontario, Canada M3J 1P3
- **Frank Sobott**, Structural Genomics Consortium, University of Oxford, Botnar Research Centre, Oxford OX3 7LD, United Kingdom
- **Kihyung Song**, Department of Chemistry, Korea National University of Education, Chongwon, Chungbuk 363-791, Korea

- **Jiangping Wang**, Department of Chemistry, Wayne State University, Detroit, MI 48202
- Lai-Sheng Wang, Department of Physics, Washington State University, 2710 University Drive, Richland, WA 99352; W. R. Wiley Environmental Molecular Sciences Laboratory and Chemical Sciences Division, Pacific Northwest National Laboratory, MS K8-88, P.O. Box 999, Richland, WA 99352
- Ping Wang, Department of Chemistry, The University of Akron, Akron, OH 44325
- **Xue-Bin Wang**, Department of Physics, Washington State University, 2710 University Drive, Richland, WA 99352; W. R. Wiley Environmental Molecular Sciences Laboratory and Chemical Sciences Division, Pacific Northwest National Laboratory, MS K8-88, P.O. Box 999, Richland, WA 99352
- **Chrys Wesdemiotis**, Department of Chemistry, The University of Akron, Akron, OH 44325
- **Justin M. Wiseman**, Department of Chemistry, Purdue University, 560 Oval Drive, West Lafayette, IN 47907-2038
- Vicki H. Wysocki, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- Xin Yang, Department of Physics, Washington State University, 2710 University Drive, Richland, WA 99352; W. R. Wiley Environmental Molecular Sciences Laboratory and Chemical Sciences Division, Pacific Northwest National Laboratory, MS K8-88, P.O. Box 999, Richland, WA 99352
- **Qingfen Zhang**, Department of Chemistry, Box 210041, University of Arizona, 1306 East University Avenue, Tucson, AZ 85721-0041
- **Roman Zubarev**, Laboratory for Biological and Medical Mass Spectrometry Uppsala University, Box 583, Uppsala S-751 23, Sweden

PREFACE

The introduction of biological molecules into the gas phase by matrix-assisted laser desorption/ionization (MALDI) and electrospray ionization (ESI) has led to a revolution in biological mass spectrometry. The analytical aspects are a success story. Molecular weights can be determined with a high precision, peptide sequencing is now done with great success, and even higher-order structures of peptides and proteins can be accessed using mass spectrometry. Exceptionally high sensitivity, high mass resolution, and inherent speed are the key factors that positioned mass spectrometry at the forefront of analytical techniques for identification and characterization of biomolecules.

This success is based largely on the principles of mass spectrometry that have been developed since the mid-1970s for small organic molecules. However, studies of biomolecules in the gas phase have also revealed a number of challenges associated with the flexibility and the size of these species. For example, it was difficult to achieve efficient fragmentation of large molecules using traditional mass spectrometric approaches. Understanding of fundamental limitations of the existing ion activation techniques resulted in development of novel analytical approaches for studying fragmentation of large molecules in the gas phase. Improved identification of biomolecules in real-world applications is facilitated by understanding of their fragmentation mechanisms and the effect of the primary and the secondary structure on the observed fragmentation patterns.

Because of the large size, conformational flexibility, and the ability of biomolecules to hold multiple charges, studies of biomolecular gas-phase ion chemistry have opened a number of new and exciting areas of research. Multiply charged biomolecules are excellent targets for studying ion—ion chemistry and processes following capture of low-energy electrons. Various approaches are being

xvi PREFACE

developed to gain phenomenological understanding of the formation and fragmentation of hydrogen-rich radical cations, molecular radical cations, and radical anions of peptides and proteins. Development of new approaches for studying thermochemistry of gas-phase biomolecules and their dissociation energetics is at the forefront of the field. Vibrational spectroscopy of biomolecular ions is another area of research that is currently undergoing an explosive growth. In parallel, new high-resolution spectroscopic techniques have been successfully applied to larger systems, providing feedback to mass spectrometric studies. Reactivity of mass-selected biomolecules with solid targets has a potential for preparation of novel surfaces relevant for a variety of applications in biology and biotechnology.

In addition, there are several basic aspects related to the physics of the various problems that have remained unanswered. For example, the question of ergodicity and/or statistical versus nonstatistical behavior in the breakup of biomolecules has been raised in connection with several methods, including electron capture dissociation (ECD) or photodissociation. The old questions that were raised many years ago concerning organic molecules are again at the forefront—do gas phase biomolecules undergo intramolecular vibrational redistribution (IVR) prior to dissociation? Are all vibrational modes involved in IVR? Is there site selectivity and charge-directed reactivity? The mere fact that a large protein fragments on the short timescale of mass spectrometry, which is an absolute necessity in terms of analysis and sequencing, is somewhat surprising in view of our previous knowledge of dissociation of relatively small organic molecules in the gas phase and its description using statistical theories [Rice-Ramsperger-Kassel-Marcus/quasiequilibrium theory (RRKM/QET) and the like].

This book is a collection of reviews on fundamental aspects underlying mass spectrometry of biomolecules. The various selected topics have been arranged in three parts: (1) structures and dynamics of gas-phase biomolecules; (2) activation, dissociation, and reactivity; and (3) thermochemistry and energetics.

Fundamental mass spectrometry has always been strongly linked to a variety of gas-phase spectroscopic techniques, which provide unique insights on the structure and dynamics of ions and molecules in the gas phase. High-resolution UV and IR spectroscopy discussed in Chapter 1 allows study of the structure and dynamics of individual conformers of neutral biomolecules, exploring the effect of the solvent on the intrinsic properties of these molecules, and molecular recognition by examining the behavior of gas-phase clusters of biomolecules. Chapter 2 gives an example of high-resolution photodetachment phoelectron spectroscopy studies of electron transfer in iron–sulfur (Fe–S) clusters. In particular, this technique is used to explore the effect of solvents and protein environment on the electronic properties of the cubane-type [4Fe–4S] cluster—the most common agent for electron transfer and storage in metalloproteins.

Ion-molecule reactions and H/D (hydrogen/deuterium) exchange studies have traditionally been used in mass spectrometry for structure determinations. Chapter 3 gives an overview of the application of these techniques to studies of structures and conformations of gas-phase biomolecules. While spectroscopic techniques are

PREFACE xvii

currently limited to relatively small systems, mass spectrometry has been used to investigate quaternary structures of large protein complexes. Experimental approaches utilized in such studies are summarized in Chapter 4. Protein structures and folding in the gas phase is discussed in Chapter 5. Understanding protein dynamics in the absence of solvent—the driving force and the timescale of protein folding in the gas phase—is important for separating the effect of solvent from the effect of the intrinsic properties of proteins on their dynamics in solution.

The dynamics of the intramolecular vibrational energy redistribution (IVR) in gas-phase biomolecules is discussed in Chapters 6 and 7. Classical trajectory simulations using semiempirical PM3 potential energy surfaces described in Chapter 6 are instrumental for understanding ultra fast dynamics following photoionization of biomolecules and the validity of statistical theories of dissociation of these large floppy molecules. Studies of gas-phase ion chemistry of peptides and proteins revealed a variety of very interesting phenomena, some of which (e.g., electron capture dissociation and photodissociation) were described as nonergodic processes that circumvent IVR. The pros and cons of IVR and ergodic behavior in biomolecules based on the available experimental findings are discussed in Chapter 7.

Gas-phase fragmentation of protonated peptides is an important prerequisite for peptide and protein identification using tandem mass spectrometry (MS/MS). Understanding mechanistic aspects of peptide fragmentation as a function of peptide sequence and conformation summarized in Chapter 8 plays a central role in the interpretation of MS/MS spectra and refining strategies for database searching. Most mass spectrometric studies utilize closed-shell biomolecules (protonated or cationized on metals) generated using soft ionization techniques. Formation and dissociation of peptide radical cations described in Chapter 9 is a new rapidly growing field in gas-phase ion chemistry of biomolecules. These ions are formed by gas-phase fragmentation of complexes of the corresponding neutral peptide with transition metals and various organic ligands.

Collisional activation and multiphoton excitation are conventionally used for identification of biomolecules in a variety of mass spectrometric applications. Current status of multiphoton excitation, spectroscopy, and photodissociation of gasphase biomolecules is summarized in Chapter 10. Chapter 11 presents classical trajectory simulations of the energy transfer in collisions of ions with atomic neutrals and surfaces. The phenomena observed following ion–surface collisions and the instrumentation involved in such studies are presented in Chapter 12 with particular emphasis on soft landing of biological molecules on a variety of surfaces. Soft landing can be utilized for a very specific modification of surfaces using a beam of mass-selected ions of any size and composition or for separating and preparing biomolecules on substrates in pure form for subsequent analysis.

Another method of ion activation in biological mass spectrometry relies on capture of low-energy electrons by multiply charged ions. Electron capture dissociation (ECD), discussed in Chapter 13, opens up a variety of unique dissociation pathways and provides information on the structure of the ion that is complementary to collisional or multiphoton excitation. Chapter 14 presents the

xviii PREFACE

fundamental principles of ion—ion chemistry of biomolecules. Ion—ion reactions provide a means of manipulating charge states of multiply charged peptides and proteins. Charge reduction by reactions of multiply charged biomolecules with singly charged ions of opposite polarity has developed as a powerful tool for structural elucidation of peptides and proteins.

Mass spectrometry has been widely utilized for thermochemical determinations. However, studying thermochemistry and dissociation energetics of peptides and proteins is challenging because most of the well-developed experimental approaches that have been successfully employed in the studies of small and medium-size ions are simply not applicable to the fragmentation of large molecules. Chapter 15 presents an overview of mass spectrometric approaches that have been utilized for thermochemical determinations of biomolecules and discusses the current status and limitations of these techniques, focusing on determination of proton affinities and alkali metal affinities of biomolecules. Chapter 16 describes the experimental approaches developed for studying the energetics and entropy effects in peptide and protein dissociation reactions.

Finally, we would like to acknowledge the authors of the chapters, who have invested a considerable amount of time and effort and prepared high-quality reviews for this book. Special thanks go to Jean Futrell for his generous help on various stages of this project and insightful feedback on the contents of several chapters. We are also thankful to many other colleagues who provided their comments and suggestions on the contents of this book.

JULIA LASKIN AND CHAVA LIFSHITZ

CONTENTS

CONTRIBUTORS	
PREFACE	XV
PART I STRUCTURES AND DYNAMICS OF GAS-PHASE BIOMOLECULES	1
1 Spectroscopy of Neutral Peptides in the Gas Phase: Structure, Reactivity, Microsolvation, Molecular Recognition Markus Gerhards	3
2 Probing the Electronic Structure of Fe–S Clusters: Ubiquitous Electron Transfer Centers in Metalloproteins Using Anion Photoelectron Spectroscopy in the Gas Phase Xin Yang, Xue-Bin Wang, You-Jun Fu, and Lai-Sheng Wang	63
3 Ion-Molecule Reactions and H/D Exchange for Structural Characterization of Biomolecules M. Kirk Green and Carlito B. Lebrilla	119
4 Understanding Protein Interactions and Their Representation in the Gas Phase of the Mass Spectrometer Frank Sobott and Carol V. Robinson	147
	vii

此为试读,需要完整PDF请访问: www.ertongbook.com

viii CONTENTS

5	Protein Structure and Folding in the Gas Phase: Ubiquitin and Cytochrome c Kathrin Breuker	177
6	Dynamical Simulations of Photoionization of Small Biological Molecules Dorit Shemesh and R. Benny Gerber	213
7	Intramolecular Vibrational Energy Redistribution and Ergodicity of Biomolecular Dissociation Chava Lifshitz	239
PA	RT II ACTIVATION, DISSOCIATION, AND REACTIVITY	277
8	Peptide Fragmentation Overview Vicki H. Wysocki, Guilong Cheng, Qingfen Zhang, Kristin A. Herrmann, Richard L. Beardsley, and Amy E. Hilderbrand	279
9	Peptide Radical Cations Alan C. Hopkinson and K. W. Michael Siu	301
10	Photodissociation of Biomolecule Ions: Progress, Possibilities, and Perspectives Coming from Small-Ion Models *Robert C. Dunbar**	337
11	Chemical Dynamics Simulations of Energy Transfer and Unimolecular Decomposition in Collision-Induced Dissociation (CID) and Surface-Induced Dissociation (SID) Asif Rahaman, Kihyung Song, Jiangping Wang, Samy O. Meroueh, and William L. Hase	379
12	Ion Soft Landing: Instrumentation, Phenomena, and Applications Bogdan Gologan, Justin M. Wiseman, and R. Graham Cooks	443
13	Electron Capture Dissociation and Other Ion–Electron Fragmentation Reactions Roman Zubarev	475
14	Biomolecule Ion–Ion Reactions Scott A. McLuckey	519

CONTENTS	ix
PART III THERMOCHEMISTRY AND ENERGETICS	565
15 Thermochemistry Studies of Biomolecules Chrys Wesdemiotis and Ping Wang	567
16 Energy and Entropy Effects in Gas-Phase Dissociation of Peptides and Proteins Julia Laskin	619
INDEX	667

PART I

STRUCTURES AND DYNAMICS OF GAS-PHASE BIOMOLECULES



SPECTROSCOPY OF NEUTRAL PEPTIDES IN THE GAS PHASE: STRUCTURE, REACTIVITY, MICROSOLVATION, MOLECULAR RECOGNITION

Markus Gerhards

Heinrich-Heine Universität Düsseldorf Institut für Physikalische Chemie I Düsseldorf, Germany

- 1.1. Introduction and Historical Background
- 1.2. Experimental Setups and Methods
 - 1.2.1. Laser Spectroscopic Methods and Microwave Spectroscopy
 - 1.2.2. Some Experimental Setups: Mass Spectrometry, Double-Resonance Spectroscopy, and Sources
- 1.3. Spectroscopy on Selected Amino Acid Model Systems
- 1.4. Double-Resonance and Microwave Spectroscopy on Amino Acids
 - 1.4.1. Phenylalanine
 - 1.4.2. Tryptophan
 - 1.4.3. Applications of Microwave Spectroscopy
- 1.5. Spectroscopic Analysis of Peptide Structures
- 1.6. Molecular Recognition
- 1.7. Calculations and Assignment of Vibrational Frequencies
- 1.8. Summary and Outlook

Principles of Mass Spectrometry Applied to Biomolecules, edited by Julia Laskin and Chava Lifshitz Copyright \odot 2006 John Wiley & Sons, Inc.

1.1. INTRODUCTION AND HISTORICAL BACKGROUND

As reported in previous chapters of the book, it has been a great challenge to transfer large molecules in the gas phase without dissociation. The investigations focus on a pure mass spectrometric analysis, but no spectroscopic information on the analyzed species is available. To obtain more information on the energy of different electronic states as well as the structure and dynamical changes of the investigated isolated species, the pure mass spectrometry has to be combined with different spectroscopic techniques. The motivation is strongly triggered by the following questions:

- (1) What are the driving forces for protein folding or aggregation of peptides?
- (2) How does solvation change the secondary structure of peptides, and how can this process be influenced, i.e. in our investigations can we perform experiments on mass-selected peptides and can we add, for instance, one water molecule after the other in order to determine how the structures will change?

By answering these questions on a molecular level, we may contribute to explanations of how structures and dynamics of peptides can be understood or predicted. The main focus of this chapter is a review on the most important combined spectroscopic and mass spectrometric analyses. This chapter focuses only on neutral amino acids and peptides; the spectroscopic investigation of ionic species is another rapidly growing field and will not be discussed here.

As mentioned in other chapters, large charged molecules can be transferred into the gas phase by applying MALDI (Karas and Hillenkamp 1988), ESI (Fenn et al. 1989), or LILBID (laser-induced liquid beam ion description) (Kleinekofort et al. 1996) and other sources. Neutral molecules can be transferred by heating sources, but in the case of pure amino acids or peptides, the molecules can easily fragment by elimination of CO2. Different sources for transferring neutral species into the gas phase are discussed in this chapter. A major breakthrough was the introduction of laser desorption sources (see Section 1.2) in combination with supersonic cooling and laser ionization (of the neutral desorbed species). The combination of this pure mass spectrometry on selected neutral species (which are ionized for detection as cations) with spectroscopic techniques was triggered by the pioneering work of Levy and coworkers (Cable et al. 1987, 1988a,b; Rizzo et al. 1985, 1986b). Starting from the analysis of amino acids by a combination of laser desorption and fluorescence spectroscopy or resonant multiphoton ionization, the Levy group increases the size of the investigated species up to tripeptides (Cable et al. 1987, 1988a,b). The spectroscopic results yield information on the vibrations of the S_1 state, especially in the low-frequency region up to several hundred wavenumbers. The amide I or amide II region as well as NH stretching modes could not be investigated. Although the work of Levy's group lead to phantastic spectroscopic results, the main drawback was that spectra could not be clearly interpreted: (1) it could not be excluded that the spectra result from an overlay of different isomers, and (2) the computer power available in the late 1980s made it impossible to get any reliable prediction of vibrational spectra of different isomers. Furthermore, the structures of S_1 states can