

# **Research in Food Science and Nutrition**

**Volume 2**

## **BASIC STUDIES IN FOOD SCIENCE**

**EDITORS**

**J.V. McLoughlin**

**B.M. McKenna**

**Proceedings of the Sixth International  
Congress of Food Science and Technology,  
Dublin, September 18 – 23, 1983**

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# **RESEARCH IN FOOD SCIENCE AND NUTRITION**

*Editors*

**J.V. McLoughlin**

**B.M. McKenna**

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Several large strains (lines 1) of various types of pneumococci were examined for their ability to synthesize various enzymes. The results are given in Table 1. The following are the results for the strains listed in Table 1. The results for the other strains are given in the accompanying tables. The results for the other strains are given in the accompanying tables. The results for the other strains are given in the accompanying tables.

TABLE 1. Enzymatic activities of various strains of pneumococci.

Strain	Enzyme	Activity	Activity	Activity	Activity
1	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
2	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
3	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
4	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
5	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
6	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
7	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
8	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
9	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000
10	Streptococcus pneumoniae	0.0000	0.0000	0.0000	0.0000

# BIOCHEMISTRY

Enzymatic activity (units/ml) of various strains of pneumococci. The results are given in Table 1. The results for the other strains are given in the accompanying tables. The results for the other strains are given in the accompanying tables.

Based on the results of the enzymatic activities, strains 1, 2 and 3 were selected for further study. The temperature and pH characteristics of each were determined. The results are given in Table 1 and Fig. 1-3. The results for the other strains are given in the accompanying tables. The results for the other strains are given in the accompanying tables.

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SCREENING OF DEXTRANASE PRODUCING FUNGI

J.M. LEE and P.F. FOX,  
Department of Food Chemistry, University College, Cork.

Eleven fungal strains (Table 1) obtained from culture collections were screened for dextranase production. Each strain was grown in Kosasics screening medium (Biotechnol. Bioeng. XV, 729-741, 1973) at 24°C for 5 d in a reciprocating water bath operating at 100 rpm. Dextranase activities (Table 1) in cell-free filtrates of the cultures were determined on dextran agar diffusion plates. One dextranase unit was defined as that amount of activity which yielded reducing sugars equivalent to 1  $\mu$  mole of iso-maltose  $\text{min}^{-1}$  at pH 6.0 and 40°C.

TABLE 1 Production and some characteristics of selected fungal dextranases

Strain	Activity <sup>1</sup>	pH opt	temp. opt, °C	pH <sup>2</sup> stability	temp. <sup>3</sup> stability, °C	temp. <sup>4</sup> stability, °C
1) <u>Aspergillus carneus</u>	0.0121	-	-	-	-	-
2) <u>A. carneus</u>	0.0004	-	-	-	-	-
3) <u>Chartomium gracile</u>	0.4810	5.0	60	4.0-11.5	≤ 60	65
4) <u>Fusarium moniliforme</u>	1.3371	4.0	60	4.5-5.5	≤ 50	65
5) <u>Paecilomyces lilacinus</u>	0.3881	4.5	50	4.5-7.5	≤ 65	70
6) <u>Penicillium aculeatum</u>	0.0287	-	-	-	-	-
7) <u>Penicillium funiculosum</u>	0.0257	-	-	-	-	-
8) <u>P. funiculosum</u>	0.0876	-	-	-	-	-
9) <u>P. funiculosum</u>	0.0664	-	-	-	-	-
10) <u>P. funiculosum</u>	0.0151	-	-	-	-	-
11) <u>P. funiculosum</u>	-	-	-	-	-	-

<sup>1</sup>dextranase activity (units ml<sup>-1</sup>) of cell-free filtrates

<sup>2</sup>> 90% residual activity following 24 h exposure at 37°C

<sup>3</sup>90% residual activity following heating for 1 min at reported temperature

<sup>4</sup>temperature causing 50% inactivation in 1 min

Based on the results of this primary screening, strains 3, 4 and 5 were selected for further study. The temperature and pH characteristics of cell-free filtrates (dialysed and lyophilized) from these strains, assayed by the blue dextran method of Mattiasson (Anal. Lett. 13(B10), 851-860, 1980), at 40°C and pH 5.0 unless otherwise stated, are summarized in Table 1 and Figs. 1-4. The optimum pH (Fig. 1) was determined on blue dextran dissolved in universal buffer (Analyst. 64, 490, 1939) over the pH range 3.0-7.5. The 3 enzymes lost activity abruptly at pH values slightly on the acid side of the optimum but showed activity over a wide pH range on the alkaline side.

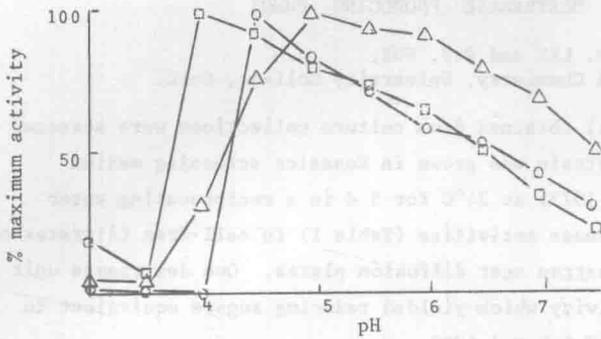


Fig. 1

Activity-pH profile on blue dextran at 40°C

Δ, strain 3  
 □, strain 4  
 o, strain 5

pH stability (Fig. 2) was determined by measuring the residual activity after incubation of the enzyme, dissolved in universal buffer at pH values from 3.0 to 11.5, for 24 h at 37°C; the dextranases secreted by strains 3 and 5 were very stable under strongly alkaline conditions. The 3 enzymes were maximally active at 50-60°C but lost activity rapidly > 60°C (Fig. 3). For the determination of heat stability (Fig. 4) the enzymes were dissolved in universal buffer pH 5.0, sealed in capillary tubes, heated for 1 min and the residual activity measured; the 3 enzymes were unstable > 65°C.

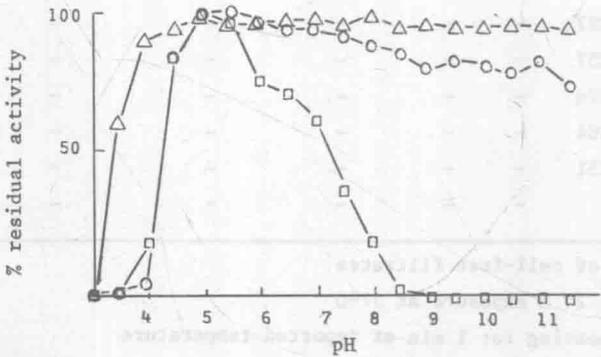


Fig. 2

pH-stability of the 3 dextranases at 37°C for 24 h

Δ, strain 3  
 □, strain 4  
 o, strain 5

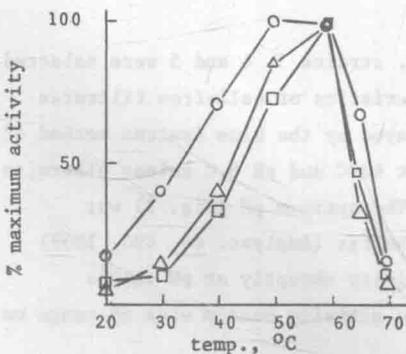


Fig. 3 Temperature-dependence of dextranase activity on blue dextran at pH 5.0

Δ, strain 3; □, strain 4;  
 o, strain 5.

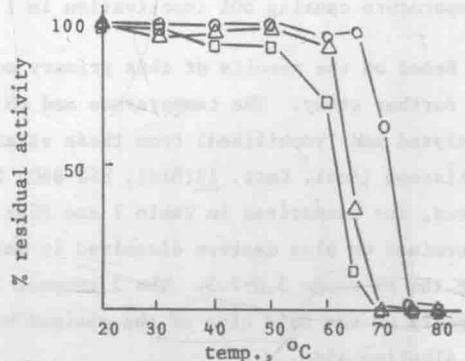


Fig. 4 Thermal inactivation of the dextranases on heating at pH 5.0 for 1 min: Δ, strain 3; □, strain 4; o, strain 5.