



**NEW ATLAS OF
HISTOLOGY**

NEW ATLAS OF HISTOLOGY

***Light microscopy,
histochemistry and
electron microscopy***

MARIANO S. H. DI FIORE

Ex Regular Assistant Professor of Histology and Embriology, Faculty of Medicine, Buenos Aires. Ex Professor of Histology from the Instituto Nacional Superior del Profesorado.

ROBERTO E. MANCINI

Director of the Centro de Investigaciones sobre Reproducción, Faculty of Medicine. Full Professor from the 2nd Chair of Histology, Cytology and Embriology, Faculty of Medicine, Buenos Aires.

EDUARDO D. P. DE ROBERTIS

Director of the Instituto de Biología Celular, Faculty of Medicine, Buenos Aires. Full Professor from the 1st Chair of Histology, Cytology and Embriology, Faculty of Medicine, Buenos Aires.

TRANSLATION

Tomás Alfredo Reader

Head Instructor from the 1st Chair of Histology, Cytology and Embriology, Faculty of Medicine, Buenos Aires.



LEA & FEBIGER

PHILADELPHIA

1978

*First Spanish edition 1971
Second Spanish edition 1973
Third Spanish edition 1976*

*First Portuguese edition 1971
Second Portuguese edition 1973
Third Portuguese edition 1975*

*First English edition 1973
Second English edition 1977
Third English edition 1978*

THIS EDITION HAS BEEN DONE IN COLLABORATION
WITH LIBRERIA "EL ATENEO" EDITORIAL

PRINTED IN ARGENTINE

Queda hecho el depósito que previene la ley N° 11.723 ©1978
"El Ateneo" Pedro García S.A. Librería, Editorial e Inmobiliaria. Florida 340 – Buenos Aires

Se terminó de imprimir el día 30 de octubre de 1978 en Artes Gráficas Planeta, Soc. en Com. p. Acc.
Castro 928/30 – Buenos Aires

**NEW ATLAS
OF HISTOLOGY**

P R E F A C E

In recent years the teaching of a laboratory course in Histology has become more complex because of the new methods that have permitted to expand into the chemical and ultrastructural organization of cells and tissues. While traditionally based on the study of the structures observed with the light microscope, the development of modern histochemical techniques has enabled us to localize many of the main cellular components thus allowing a better correlation with their function. On the other hand, in the last two decades, the technical advances in electron microscopy have permitted to reach the submicroscopic level and even to reveal changes that are produced at a macromolecular level during the functioning of the cell. Such developments are also the basis of modern Pathology since at such levels of organization a disease merely represents a further variation from the normal.

To this increased complexity, which implies the use of techniques and of costly equipment which may be unavailable in some Chairs of Histology, we have to add the recent tendency to shorten the time allowed for teaching in the laboratory. It is evident that although there is no substitute for the direct observation of the slides, with routine or special histochemical techniques, and of the original electronmicrographs, an Atlas in which the various subdivisions of Histology are presented in an orderly manner would be considerably useful both to the student and teacher.

This NEW ATLAS OF HISTOLOGY, based on morphology, histochemistry and ultrastructure was designed to meet the needs we have encountered to teach an ever increasing number of students. Its originality resides in the fact that in every chapter the three basic aspects of Histology are integrated. The presentation of morphology is based on color reproductions of drawings from the usual preparations. They correspond to a combination of different microscopic fields in which the proportions and mutual relationships are maintained. Some plates also give a tridimensional apreciation of the structures. In each case a systematic description of each drawing is given in which are emphasized the main elements used to arrive to a reasoned diagnosis.

The histochemical reactions represented by color micrographs of specifically stained preparations; the techniques of radioautography using labelled isotopes and fluorescence, for the localization of cellular antigens, and the specific enzymic reactions are discussed in detail. These methods localize essential chemical cellular components, such as nucleic acids, lipids, steroids, mucopolysaccharides and enzymes. Brief descriptions give the principles of the techniques, the most important concepts that arise from them, and how these correlate with the histophysiology of each cell or tissue.

The ultrastructural study is represented by electron microscope photographs which illustrate the intracellular structural organization at several magnifications, from the morphology of organelles to that of the macromolecules that compose them. The observation of these cellular components, ranging from 10 to 2,000 angstrom units in size, at a level where Molecular and Cellular Biology converge, make electron microscopy the most powerful instrument in morphological analysis. The description of every electron micrograph points out the most important submicroscopic structures and the functional correlations that they suggest.

Such an ambitious enterprise could not have been completed without generous collaboration. The drawings and diagrams which illustrate the morphologic sections have been done for the most part by Professor Celia M. Ishii de Sato; we would particularly like to point out her skill in accurately reproducing the structures. The retouching of the micrograph reproductions was also done by Professor de Sato. Most of these drawings are taken from the ATLAS OF HUMAN HISTOLOGY by one of us (Dr. Mariano S.H.Di Fiore). The histochemical preparations were mainly made in the Chair of Histology of Professor R.E. Mancini with the help of Prof. O. Vilar and Drs. A. Barket and L. Tres. Some of the-illustrations were also contributed by Prof. E. De Robertis. The electron microscopy, under Prof. E. De Robertis, has benefited from the kind collaboration of Professor David Zambrano and Amanda Pellegrino de Iraldi, with the efficient technical help of Mrs. Lina de Stein. Dr. T.A. Reader, also from this chair, collaborated in the english version of this edition.

We also acknowledge the Publishers "El Ateneo" for the great effort they have put into the publication of this Atlas. It is a pleasure to acknowledge once more the help of those who contributed a great deal to previous editions: Mr. Segundo Domingo Torres, for the diagrams; Mr. José Moreno Soria for the graphic coordination. Finally the lay-out of this book is mainly due to the skill of the staff of Artes Gráficas Planeta, directed by Mr. Nicolás Rubino.

Mariano H. Di Fiore
Roberto E. Mancini
Eduardo De Robertis

PREFACE OF THE THIRD EDITION

The main lines of thought mentioned in the Preface of the first edition of the New Atlas of Histology have been reassured by the excellent reception of this book, not only in the English version, but also in its translations into Spanish, Portuguese and Italian. Thus, it was demonstrated that an integration in the morphological histochemical and ultrastructural study of the cells and tissues is the right approach in modern Histology.

In this new edition additions and replacements of plates have been made which represent a considerable improvement. The figure of the Patches of Peyer of the intestine has been replaced and new drawings have been added which represent: the juxtaglomerular complex (kidney); the tubuli recti, the rete testis and ductuli efferents (testicle); the colliculus seminalis (prostate gland); the corpus luteum and the atretic follicles (ovary); the mammary gland at the seventh month of pregnancy and during lactation; and general fields of the thyroid and parathyroid glands. Plates 6 and 7 corresponding to the ultrastructure of the cell have been replaced by others that illustrate better the various cell organelles. Another addition is an electron micrograph corresponding to the alveolar-capillary wall of the lung, across which the exchange of oxygen and carbon dioxide is produced. Furthermore the entire text has been revised and several errors, which unfortunately were in the former editions, were corrected.

October, 1978

CONTENTS

PLATE	CYTOLOGY	Page
1. Cytology I		2
Fig. 1. Oöcyte in a developing follicle		
Fig. 2. Oöcyte under phase contrast		
Fig. 3. Oöcyte in vitro		
2. Cytology II		4
Fig. 1. Epithelial cells from the oral mucosa examined under the light microscope		
Fig. 2. Epithelial cells from the oral mucosa examined under phase contrast		
Fig. 3. Epithelial cells from the oral mucosa stained with janus green		
Fig. 4. Same cells stained with neutral red		
Fig. 5. Peritoneal serosa treated with a silver nitrate solution		
Fig. 6. Simple squamous epithelium (mesothelium from peritoneum)		
3. Cytology III		6
Fig. 1. The Golgi apparatus		
Fig. 2. Mitochondria in liver cells		
Fig. 3. DNA in nuclei of liver cells		
Fig. 4. DNA and RNA in liver cells		
Fig. 5. RNA and DNA in liver cells		
Fig. 6. RNA in liver cells		
4. Cytology IV		8
Fig. 1. Protein synthesis in liver cells		
Fig. 2. Glycogen in liver cells		
Fig. 3. Lipids in liver cells		
Fig. 4. Lipochromes in liver cells		
Fig. 5. Alkaline phosphatase in cells from the tubule of the kidney		
Fig. 6. Acid phosphatase in cells from the tubule of the kidney		
5. Ultrastructure of the Cytoplasm (Endoplasmic Reticulum and Lysosomes)		10
6. Ultrastructure of the Cytoplasm (Endoplasmic reticulum, Golgi complex, peroxisomes and lipoproteins)		12
7. Ultrastructure of the plasma membrane, mitochondria, microtubules and microfilaments		14
8. Mitosis I		16
Fig. 1. Prophase in egg of Ascaris		
Fig. 2. Metaphase (lateral view)		
Fig. 3. Metaphase (viewed from a pole)		
Fig. 4. Initial stage of the anaphase		
Fig. 5. Late or advanced anaphase		
Fig. 6. Telophase		
9. Mitosis II		18
Fig. 1. Incorporation of thymidine H ³ in mononuclear leukocytes at premitotic phase		
Fig. 2. Thymidine in leukocytes in advanced prophase		

PLATE	Page
-------	------

Fig. 3. Human bone marrow (May-Grünwald method 800X)	
Fig. 4. Human bone marrow (May-Grünwald method 900X)	
Fig. 5. Human bone marrow. Anaphase	
Fig. 6. Human bone marrow. Late anaphase	
Fig. 7. Liver cells in mitosis. Prophase	
Fig. 8. Liver cell in initial anaphase	
Fig. 9. Giant chromosome from the salivary gland of <i>Drosophila</i>	
 10. Meiosis in the Testicle of the Locust	20
 11. Cariotype and Sexual Chromatin	22
Fig. 1. Metaphase in a smear of human leukocytes	
Fig. 2. Chromosomes from a leukocyte in metaphase	
Fig. 3. Human idiograms representing the pairs of chromosomes	
Fig. 4. Sexual chromatin in nerve cell from the cat	
Fig. 5. Sexual chromatin in epithelial cell from the oral mucosa from a woman	
Fig. 6. Leukocytes in a smear from human blood	

T I S S U E

Epithelial Tissue

 12. Epithelial Tissue	 24
Fig. 1. Simple columnar epithelial tissue	
Fig. 2. Simple columnar epithelium with striated border	
 13. Epithelial Tissue	 26
Fig. 1. Stratified squamous epithelium (sagittal section)	
Fig. 2. Stratified squamous epithelium (tangential section)	
 14. Epithelial Tissue	 28
Fig. 1. Pseudostratified columnar ciliated epithelium	
Fig. 2. Transitional epithelium (polymorphic)	
 15. Epithelial Tissue	 30
Fig. 1. Vibratile cilia of the trachea	
Fig. 2. Stereocilia of the epithelium of the epididymus	
Fig. 3. Striated border of the intestinal epithelium	
Fig. 4. Brush border of the convoluted tubule of the kidney	
Fig. 5. Tonofibrils of the stratified epithelium	
Fig. 6. Basement membrane in tracheal mucosa	
Fig. 7. Basement membrane in human epidermis	
Fig. 8. Basement membrane in human epidermis	
 16. Epithelial Tissue	 32
Fig. 1. Glycogen in the middle layers of stratified squamous epithelium	
Fig. 2. Nucleoproteins in the basal layers of stratified epithelium	
Fig. 3. Protein synthesis in the basal and middle layers of stratified epithelium	
Fig. 4. Mucin in the cells of the gastric mucosa	
Fig. 5. Goblet cells of the intestinal epithelium	
Fig. 6. Incorporation of radioactive sulphur S^{35} in the goblet cells	
 17. Ultrastructure of some Apical Differentiations in Epithelial Cells	 34
 18. Ultrastructure of some Apical Differentiations in Epithelial Cells	 36
 19. Differentiations in Epithelial Cells (Desmosomes and Tonofibrils)	 38
 20. Differentiations from the Basal Region of Epithelia (Basement Membrane)	 40
 21. Simple Branched Tubular Gland (diagram)	 42

PLATE	Page
22. Epithelial Tissue	44
Fig. 1. Compound acinous gland (diagram)	
Fig. 2. Compound alveolar gland (diagram)	
Connective Tissue	
23. Connective Tissue	46
Fig. 1. Fibroblasts of connective tissue	
Fig. 2. RNA in the cytoplasm of fibroblasts	
Fig. 3. Macrophage with hemosiderin granules	
Fig. 4. Ranvier's edema method	
Fig. 5. Mastocytes or mast cells	
Fig. 6. The plasmocyte. RNA in cytoplasm	
Fig. 7. Adipose cells (adipocytes)	
24. Ultrastructure of Connective Tissue Cells	48
25. Ultrastructure of the Plasmocyte or Plasma Cell	50
26. Connective Tissue	52
Fig. 1. Collagenous fibers from human dermis	
Fig. 2. Bundles of collagen from human dermis	
Fig. 3. Reticular fibers from human spleen	
Fig. 4. Elastic fibers from human dermis	
Fig. 5. The amorphous or interfibrillar substance	
27. Connective Tissue	54
Fig. 1. Loose (irregularly arranged) connective tissue	
Fig. 2. Dense connective tissue (irregularly arranged)	
Fig. 3. Cell types in connective tissue	
28. Mucous Tissue	56
Fig. 1. Mucous tissue. Wharton's jelly	
Fig. 2. Dense regular connective tissue (tendon)	
Fig. 3. Arterial and capillary beds in loose connective tissue	
Cartilage	
29. Cartilage	58
Fig. 1. Hyaline cartilage (fresh section)	
Fig. 2. Hyaline cartilage (under phase contrast)	
Fig. 3. Metachromasia of the hyaline cartilage	
Fig. 4. Incorporation of radioactive sulphate S ³⁵	
30. Cartilage	60
Fig. 1. Hyaline cartilage (postfixed and stained preparation)	
Fig. 2. Fibrous cartilage	
Fig. 3. Elastic cartilage	
31. Ultrastructure of Perichondrium and Cartilage	62
Bone	
32. Bone	64
Fig. 1. Compact bone, dried (transverse section of the diaphysis of the tibia)	
Fig. 2. Microscopic examination	
Fig. 3. Polarization microscopy	
33. Bone	66
Fig. 1. Cancellous bone; adult sternum; decalcified transverse section	
Fig. 2. Intramembranous bone formation (jaw-bone of a fetus of five months decalcified)	
34. Intracartilaginous Bone Formation: Developing Metacarpal Bone (Longitudinal Section)	68

PLATE	Page
35. Intracartilaginous Bone Formation	70
36. Formation of Bone in the Lower Mandible (Jaw Bone)	72
37. Bone (Histochemistry)	74
Fig. 1. Alkaline phosphatase in osteoblasts	
Fig. 2. Acid phosphatase in osteoclasts	
Fig. 3. Incorporation of radioactive phosphor P ³² in osteoid trabeculae	
Fig. 4. Incorporation of labelled sulphate S ³⁵ in cartilage in the process of growth	
38. Joint	76
Fig. 1. Section of a diarthrodial joint (hematoxylin-eosin 120X)	
Fig. 2. Section of a diarthrodial joint (hematoxylin-eosin 280X)	
Fig. 3. Section of a prolongation of the synovial capsule	
Muscle Tissue	
39. Muscle Tissue	78
Fig. 1. Smooth muscle fibers (from the wall of the distended bladder of the toad)	
Fig. 2. Striated muscle fibers (from the leg of a toad)	
40. Muscle Tissue	80
Fig. 1. Striated muscle (muscles of the tongue)	
Fig. 2. Smooth muscle (muscle layer of the intestine)	
Fig. 3. Cardiac muscle (myocardium)	
Fig. 4. Glycogen in striated muscle fibers	
41. Muscle Tissue	82
Fig. 1. Longitudinal section of striated fibers	
Fig. 2. Similar section at a higher magnification	
Fig. 3. Sensitive nerve endings or neuromuscular spindles	
Fig. 4. Motor plate on the surface of the muscle fiber	
Fig. 5. Blood vessels of the skeletal muscle tissue	
Fig. 6. Vascular bed demonstrated by enzymatic reaction	
42. Ultrastructure of cardiac muscle	84
Nervous Tissue	
43. Gray Matter	86
Fig. 1. Anterior horn of the spinal cord (Nissl's technique)	
Fig. 2. Anterior horn of the spinal cord (Cajal's method)	
Fig. 3. Anterior horn of the spinal cord (Golgi's method)	
Fig. 4. Anterior horn of the spinal cord (Weigert-Pal's method)	
Fig. 5. Synaptic knobs in the anterior horn (silver impregnation)	
44. Neuroglia	88
Fig. 1. Fibrous astrocytes of the brain	
Fig. 2. Oligodendrocytes of the brain	
Fig. 3. Microglia of the brain	
45. Ultrastructure of the Nervous Tissue (The Neuron)	90
46. Ultrastructure of the Nervous Tissue (Neuroglia and Neuropil)	92
47. Nerves	94
Fig. 1. Myelinated nerve fibers	
Fig. 2. Wallerian degeneration	
Fig. 3. Transverse section of a nerve	
48. Nerves and Nerve Fibers	96
Fig. 1. Portion of sciatic nerve (hematoxylin-eosin 50X)	
Fig. 2. Portion of sciatic nerve (hematoxylin-eosin 800X)	
Fig. 3. Transverse section of the sciatic nerve (hematoxylin-eosin 800X)	

PLATE	Page
-------	------

- Fig. 4. Portion of sciatic nerve (Protargol and aniline blue 800X)
 Fig. 5. Transverse section of the sciatic nerve (Protargol and aniline blue 800X)
 Fig. 6. Transverse section of a branch of the vagus nerve (Mallory-Azan 800X)

CARDIOVASCULAR SYSTEM

49. Blood and Lymph Vessels and Portal Vein	98
Fig. 1. Blood and lymph vessels (transverse section)	
Fig. 2. Large vein; portal vein (transverse section)	
50. Neurovascular Bundle and Aorta	100
Fig. 1. Neurovascular bundle (transverse section)	
Fig. 2. Artery of the elastic type: aorta (transverse section)	
51. Vascular Walls	102
Fig. 1. Layers that form the walls of the arteries	
Fig. 2. Reticulin fibers in the vessels	
Fig. 3. Elastic fibers of the aorta	
Fig. 4. Amorphous substance	
Fig. 5. Capillary (transverse section)	
Fig. 6. Capillary (tangential section)	
Fig. 7. Alkaline phosphatase reaction	
52. Ultrastructure of an Arteriole	104
53. Ultrastructure of a Blood Capillary (Pinocytosis and Micropinocytosis)	106
54. Heart: Left Atrium and Ventricle (Panoramic View); Longitudinal Section	108
55. Heart	110
Fig. 1. Pulmonary artery, pulmonary valve and right ventricle (panoramic view; longitudinal section)	
Fig. 2. Purkinje fibers (hematoxylin-eosin)	
Fig. 3. Purkinje fibers (Mallory-Azan)	

LYMPH SYSTEM

56. Lymph Node (panoramic view)	112
57. Lymph Node (sectional view)	114
58. The Spleen	116
Fig. 1. Panoramic view	
Fig. 2. Red and white pulp	
Fig. 3. Development of lymphocytes and related cells	
59. The Thymus	118
Fig. 1. Panoramic view	
Fig. 2. Thymic lobule (sectional view)	
60. Lymphatic Formations	120
Fig. 1. Palatine tonsil	
Fig. 2. Peyer's patch	
61. Lymph Nodes	122
Fig. 1. Lymphatic follicle	
Fig. 2. Lymphatic sinus	
Fig. 3. Protein incorporation by macrophages	
Fig. 4. Fig. 3 at a higher magnification	
Fig. 5. Incorporation of radioactive thymidine by lymphocytes	
Fig. 6. Anti-gammaglobulin reaction in plasmocytes	

PLATE		Page
BONE MARROW		
62. Bone Marrow		124
Fig. 1. Hemopoietic bone marrow		
Fig. 2. Bone marrow of a rabbit; India ink preparation		
63. Bone Marrow (Smear)		126
64. Ultrastructure of a Megakaryocyte and Blood Platelets		128
B L O O D		
65. Peripheral Blood Smear		130
66. Ultrastructure of an Eosinophil Leukocyte		132
67. Special Techniques		134
Fig. 1. Supravital stain		
Fig. 2. Pappenheim's and Celani's stains		
DIGESTIVE TRACT		
The Lip		
68. Sagittal section		136
Tooth		
69. Dried Tooth		138
Fig. 1. Panoramic view; longitudinal section		
Fig. 2. Layers of the crown		
Fig. 3. Layers of the root		
70. Developing Tooth		140
Fig. 1. Panoramic view		
Fig. 2. Sectional view		
The Tongue		
71. Anterior Region (Apex); Longitudinal Section; Panoramic View		142
72. The Tongue		144
Fig. 1. Circumvallate papilla		
Fig. 2. Foliate organ (longitudinal section)		
Esophagus		
73. Esophagus. Panoramic view (Transverse Section)		146
74. Upper Esophagus: Wall (Transverse Section)		148
75. Upper Esophagus: Mucosa (Transverse Section)		150
76. Esophagus		152
Fig. 1. Van Gieson's trichrome stain		
Fig. 2. Mallory-Azan's trichrome stain		
The Stomach		
77. The Stomach		154
Fig. 1. Fundus (transverse section)		
Fig. 2. Muciparous cells		
Fig. 3. Principal cells		

PLATE	Page
78. Gastric Mucosa. Mucosa of the Fundus (Transverse Section)	156
79. Fundus	158
Fig. 1. Superficial region of the mucosa of the fundus or body	
Fig. 2. Deep region of the mucosa of the fundus or body	
80. Ultrastructure of a Gastric Parietal Cell	160
81. Gastric Mucosa: Pyloric Region	162
82. The Cardias (Longitudinal Section)	164
83. Pyloro-Duodenal Junction	166
The Intestines	
84. The Small Intestine	168
Fig. 1. The jejunum-ileum (transverse section)	
Fig. 2. Glands of Lieberkühn with Paneth cells	
Fig. 3. Glands of Lieberkühn with argantaffine cells	
85. Small Intestine: The Duodenum (Longitudinal Section)	170
86. The Intestine	172
Fig. 1. The small intestine. Villi	
Fig. 2. The large intestine. Crypts of Lieberkühn. Deep region	
87. The Small Intestine	174
Fig. 1. Intestinal villi	
Fig. 2. Alkaline phosphatase in villi	
Fig. 3. Vascular bed of the intestinal villi	
88. Large Intestine	176
Fig. 1. Transverse section	
Fig. 2. Goblet cells	
89. Appendix (Transverse Section)	178
90. The Rectum (Transverse Section)	180
91. Anorectal Junction	182
Glands	
92. Salivary Gland: Parotid	184
93. Salivary Gland: Submaxillary	186
94. Salivary Gland: Sublingual	188
95. Histochemistry of Glands	190
Fig. 1. Mixed gland (submaxillary) (Mallory's trichromic method)	
Fig. 2. Region of the excretory ducts (Mallory's trichromic method)	
Fig. 3. Serous gland (pancreas) (toluidine blue stain)	
Fig. 4. Mixed gland (toluidine blue stain)	
Fig. 5. Mucous gland (PAS-hematoxylin stain)	
Fig. 6. Mucous gland (radioautography)	
The Liver	
96. Liver Lobule. Transverse Section under Low Magnification	192

PLATE		Page
97. Liver Lobule		194
Fig. 1 Liver lobule (partial view)		
Fig. 2. Biliar capillaries in the hepatic lobule		
98. Liver Lobule		196
Fig. 1. Glycogen in the hepatic lobule		
Fig. 2. Liver: reticuloendothelium, India ink preparation		
Fig. 3. Liver: reticuloendothelium, India ink preparation		
99. Liver Lobule		198
Fig. 1. Del Rio Hortega's stain: reticular fibers in an hepatic lobule		
Fig. 2. The perilobular and intralobular vascular system		
The Gall Bladder		
100. The Gall Bladder		200
The Pancreas		
101. The Pancreas		202
Fig. 1. The pancreas		
Fig. 2. Pancreatic acini (special preparation)		
Fig. 3. Islets of Langerhans (special preparation)		
102. Ultrastructure of Secretion (Cells of the Exocrine Pancreas)		204
103. Ultrastructure of Secretion (Cells of the Endocrine Pancreas)		206
RESPIRATORY TRACT		
104. Nasal Fossae		208
Fig. 1. Frontal section of the nasal fossae of pig		
Fig. 2. Olfactory area. 220X		
Fig. 3. Respiratory zone. 120X		
Fig. 4. Respiratory zone. 320X		
Fig. 5. Section of respiratory mucosa		
105. The Larynx (Frontal Section)		210
106. The Trachea		212
Fig. 1. Transverse section		
Fig. 2. Sectional view		
107. The Lung		214
108. The Lung (Detail of Different Fields)		216
Fig. 1. Secondary (lobar) bronchus		
Fig. 2. Terminal bronchiole		
Fig. 3. Respiratory bronchiole		
Fig. 4. Alveolar septa		
109. The Lung		218
Fig. 1. Reticulin in interalveolar stroma		
Fig. 2. Elastic fibers of the interalveolar septa		
Fig. 3. Perialveolar macrophages		
Fig. 4. Elastic layer in the visceral pleura		
Fig. 5. Interalveolar capillary bed		
110. Ultrastructure of the lung		220
URINARY TRACT		
111. The Kidney (Panoramic View of a Pyramid)		222

PLATE		Page
112 The Kidney	Fig. 1. Deep Cortical Area Fig. 2. Yuxtaglomerular complex	224
113. The Kidney (Deep Medular Area)	Fig. 1. Transverse section Fig. 2. Longitudinal section	226
114. The Kidney	Fig. 1. Section of the glomerulus and renal ducts (PAS-hematoxylin technique) Fig. 2. Section of the renal glomerulus Fig. 3. Section of the glomerulus and renal ducts (Mallory's trichromic method) Fig. 4. Reticulin fibers in the glomerulus and renal ducts	228
115. The Kidney	Fig. 1. Fluorescent protein in glomerular capillaries and the proximal convoluted tubule Fig. 2. Alkaline phosphatase in the kidney Fig. 3. Acid phosphatase in the kidney Fig. 4. Arterial circulation in the kidney	230
116. Ultrastructure of the Kidney Glomerulus		232
117. Ultrastructure of the Proximal Convolute Tubule		234
118. Ultrastructure of the Distal Convolute Tubule		236
119. The Ureter	Fig. 1. Transverse section Fig. 2. A sector of the wall	238
120. Urinary Bladder	Fig. 1. Panoramic view of the wall Fig. 2. Tunica mucosa	240

MALE REPRODUCTIVE SYSTEM

121. The Testis	Fig. 1. The testis (partial view) Fig. 2. Tubuli recti and rete testis	242
122. The Testis. Seminiferous Tubules (Transverse Section)		244
123. Testis and Epididymis	Fig. 1. Efferent tubules Fig. 2. The ductus epididymis (Duct of the epididymis)	246
124. The Testis	Fig. 1. Human fetal testis Fig. 2. Infantile testis Fig. 3. Presence of glycogen in a Sertoli cell Fig. 4. Acrosome in spermatids and spermatozoa Fig. 5. The Leydig cell from the human testis Fig. 6. Cholesterol in human Leydig cells Fig. 7. Smear of spermatozoa of guinea pig Fig. 8. Hyaluronidase in the acrosome of spermatozoa	248
125. Ductus Deferens and Seminal Vesicle	Fig. 1. Ductus deferens (transverse section) Fig. 2. Seminal vesicle	250
126. Prostate	Fig. 1. Section of the prostate Fig. 2. Acid phosphatase in the prostate	252

PLATE	Page
127. Prostate and Penis	254
Fig. 1. Colliculus seminalis	
Fig. 2. The Penis (Transverse Section)	
FEMALE REPRODUCTIVE SYSTEM	
128. The Ovary (Transverse Section; Panoramic View)	256
129. The Ovary	258
Fig. 1. Cortical region: primary and maturing follicles	
Fig. 2. Wall of a mature Graafian follicle	
130. Corpus Luteum	260
Fig. 1. Panoramic view	
Fig. 2. Peripheral margin at a higher magnification	
131. Human Ovary (General view)	262
132. The Ovary (Histochemistry)	264
Fig. 1. Lipids in the follicle	
Fig. 2. 3- β -ol-dehydrogenase reaction	
Fig. 3. Mucopolysaccharides in the follicular fluid	
Fig. 4. Incorporation of S ³⁵ sulphate in the follicular cells	
Fig. 5. Fluorescent proteins in the follicular fluid	
Fig. 6. 3- β -ol-dehydrogenase reaction in the corpus luteum	
133. The Ovarian Follicle	266
134. The Uterine Tube	268
Fig. 1: Panoramic view (transverse section)	
Fig. 2. Tunica mucosa	
135. Uterus. Proliferative Period; Estrogenic or Follicular Phase	270
136. Uterine Mucosa. Secretory Period. Progravidic or Progestational Phase	272
137. Mucosa of the Uterus. Menstrual Phase	274
138. Human Placenta	276
Fig. 1. Five months' pregnancy (panoramic view)	
Fig. 2. Chorionic villi. Placenta at five months	
Fig. 3. Chorionic villi. Placenta at term	
139. Cervix	278
Fig. 1. Longitudinal section	
Fig. 2. Mucin in the uterine cervix	
140. Vagina	280
Fig. 1. Longitudinal section	
Fig. 2. Glycogen in the human vaginal epithelium	
Fig. 3. Elastic fibers in middle layer of the lamina propria	
141. Vagina: Exfoliate Cytology	282
142. I. Mammary Gland	284
Fig. 1. Inactive gland	
Fig. 2. Mammary gland during the first half of pregnancy	
143. II. Mammary Gland	286
Fig. 1. Seventh month of gestation	
Fig. 2. During Lactation	