MICRODIFFUSION ANALYSIS AND VOLUMETRIC ERROR

EDWARD J. CONWAY

P47/16 0653/5

MICRODIFFUSION ·ANALYSIS AND VOLUMETRIC ERROR

BY

EDWARD J. CONWAY

M.D., D.Sc., Sc.D., F.R.I.C., F.R.C.P.I., F.R.S.A., F.R.S.

PROFESSOR OF BIOCHEMISTRY, UNIVERSITY COLLEGE, DUBLIN;
MEMBER OF THE ROYAL IRISH ACADEMY;
CHAIRMAN OF THE DUBLIN INSTITUTE OF ADVANCED STUDIES;
MEMBER OF THE ACADÉMIE SEPTENTRIONALE, PARIS;
MEMBER OF PONTIFICAL ACADEMY OF SCIENCES.

LONDON
CROSBY LOCKWOOD & SON LTD.
26 OLD BROMPTON ROAD S W 7

HEFACE TO FIFTH EDITION

1948), adapted for microdiffusion. Gutamic seid may also betermined simultaneously with glutamine.

PREFACE TO FIFTH EDITION

The use of microdiffusion analysis for various micro determinations being now widespread, it is considered sufficient in this preface to draw attention only to some new applications and developments of interest.

With regard to new methods and applications, and taking such in the order in which they appear in the present volume, one may note the following:

(a) The blood ammonium method has been much used of late in the study of hepatic disease. Among the many publications one may instance the paper of McDermott and Adams (1954), of Riddell and McDermott (1954), of Singh, Punjab, Barclay and Cooke (1954), of Sherlock, Summerskill, White and Phear (1954), of Traeger, Gabuzda, Ballou and Davidson (1954) and of White et al (1954).

As a result blood ammonium determinations are now being conducted in other laboratories as a routine procedure, using the microdiffusion technique, whereas before its introduction such measurements were regarded as an analytical refinement necessitating long experience and elaborate precautions, and unsuitable for the easy or rapid accumulation of exact data. In this connection, a recent publication of Hulme and Cooper (1956) has the title 'Blood ammonium estimation as a routine laboratory procedure'. Reference may be also made to a paper by A. E. Reif on 'The Ammonia Content of Blood and Plasma' (1960).

Besides a description of the blood ammonium method there is included in the present volume a brief review of results obtained in the study of hepatic disease (Chapter XIII, pp. 124–128).

- (b) In connection with total nitrogen determinations the procedure of Weil-Malherbe and Green (1955) for total nitrogen determination in brain slices is incorporated (Chapter XIV, p. 138), the interest here being the use of the microdiffusion procedure in conjunction with a mercury catalyst in the micro-Kjeldahl digest.
- (c) A method for the determination of glutamine is described in Chapter XXI, p. 187. This method is essentially that of Krebs

(1948), adapted for microdiffusion. Glutamic acid may also be determined simultaneously with glutamine.

(d) The determination of monoamine oxidase and of histaminase by Cotzias and Dole (1951) using a microdiffusion procedure is described in Chapter XXII, p. 190, with the distribution of histaminase in animal tissues.

With these enzyme determinations may also be grouped that of acetylcholinesterase by Serlin and Cotzias (1955) as described in Chapter XXIX, p. 239.

(e) A group of microdiffusion determinations by Feldstein and Klendshoj (1954) including cyanide (even in normal blood) sulphide, phenols, methanol and isopropanol is described in Chapters XXX and XXXI, pp. 241–247; also their system for the determination of volatile poisons of toxicological interest in Chapter L, p. 335.

In this connection there may be mentioned the determination of azide by Brady and O'Callaghan (1955) (Chapter XXX, p. 242) and of urethane by Boyland and Rhoden (1949), the latter depending on the liberation of ethanol by alkali as described in Chapter XXXIII, p. 256.

(f) The application to the determination of formaldehyde, using the chromotropic acid reaction has led to the determination of the formaldehydogenic steroids by groups of workers. Bassil and Hain (1950) first reported the use of the microdiffusion procedure for absorbing the formaldehyde produced by periodic acid oxidation, the procedure being improved by Hollander, DiMauro and Pearson (1951), also by Wilson and Hildegarde (1953) as described in Chapters XXXV and XXXVI, pp. 262–268.

Brooks and Norymberski (1953) and Edwards and Kellie (1954) employed a similar technique after oxidation by sodium bismuthate instead of periodic acid in the outer chamber. Their procedures are given in Chapter XXXVI, p. 266. In this connection also, one may note the determination of the amino acid, glycine, by Schwartz, Robertson and Holmes (1955), depending on the conversion of glycine to formaldehyde by the action of ninhydrin (Chapter XXXVII, p. 269).

(g) The application of the microdiffusion technique by Burbridge, Hine and Schick (1950) to the determination of acetalde-

hyde, using semicarbazide as the absorbent, is described in Chapter XXXVIII (p. 273). In this connection reference may also be made to the more recent work of Berka on the estimation of carbon monoxide in blood (1955). He uses the microdiffusion procedure and in one variant shows how a semi-quantitative estimation of carbon monoxide in 0.2 ml. of a blood sample can be completed in five minutes.

The procedure, using semicarbazide absorption, has recently been employed by Ryan (1958) in this laboratory for an improved method for the determination of lactic acid in blood, and applicable also to tissues (Chapter XXXIX, p. 277). In this the lactic acid is oxidised to acetaldehyde by ceric sulphate in acid solution in the outer chamber of the microdiffusion 'unit'. With this improved method, which gives practically quantitative results over a wide range, glucose need not be removed prior to the oxidation, and a trichloroacetic acid filtrate can be used directly.

- (h) Developments in halogen determinations may be noted with respect to the use of Fast Green as absorbent by Gordon (1952), Chapter XLIII, p. 301, and the determination of organically bound halogens by Gordon (1952) and by Pirt and Chain (1952), as described in Chapters XLIII (p. 301) and XLVI (p. 320).
- (i) With regard to carbon monoxide determinations, a rapid clinical method by Lehmann (1944) is described in Chapter XLIX, p. 333, his procedure for plasma bicarbonate being outlined in Chapter XXIV, p. 201.

New developments in apparatus

A modification of the standard microdiffusion unit, by Öbrink (1955) is described in Chapter II, p. 13. It should prove useful in special conditions, e.g. relatively high temperatures, and in halogen determinations, at the lowest concentration levels. Another modification has been described by Berka (1959) which could be used also for relatively high temperatures.

A modification used by Edwards and Kellie (1954) which they found useful in the determination of formadehydogenic steroids is described in Chapter XXXIV, p. 259, and a 'diffusion-distillation unit' introduced by Kirk (1950), in Chapter II, p. 14.

A rack and holder used for the handling of large numbers of

'units' at the same time and introduced by Schwartz, Robertson and Holmes (1955) is shown in Fig. 57, p. 270.

For occasional shaking of 'units' when this is required, a new 'vibrating table' is described in Chapter III, p. 24. A 'vibrating table' for shaking has been in occasional use in this laboratory since 1953 and is open to further improvement. From the results obtained therefrom, it appears that a vibrating table in one form or another is the most efficient shaker for the 'units'.

My thanks are due to the various workers mentioned above for their kind permission to include material and illustrations from their published work; and also the editors and publishers of the Annals of Surgery; Analytical Chemistry; the Analyst; the Biochemical Journal; the Journal of Biological Chemistry; the Journal of Clinical Endocrinology; the Journal of Laboratory and Clinical Medicine; and the Lancet.

While this edition was in press my attention was drawn by Professor Ishizaka of Nagoya City University (who translated this book into Japanese) to a microdiffusion unit in use in Japan. The upper edge has a rim which projects out some few millimetres, and the unit is covered by an octagonal shaped lid. A flexible and simply made metal band can be used (if desired) to fix the lid more firmly in position. Professor Ishizaka has also designed a special lid with suitable inlet tubes, which he has used successfully to determine accurately the oxygen content of blood. It is hoped to describe this work in a subsequent edition.

EDWARD J. CONWAY

University College Dublin FIRST PUBLISHED 1939
2ND REVISED EDITION 1947
3RD REVISED EDITION 1950
4TH REVISED EDITION 1962

© Crosby Lockwood & Son Ltd., 1962

THE.

'Yello trade the property of the control of the con

the state of the s

745 dollar or amination of the cine

Tell arotanimedes ses ann —

By Add Add Ary on the ses and a con
ME 2005, nonlearner of the con
ME 2005, nonlearner of the con-

rea mecanimation, TV.

. Let incidence roun on the

PRINTED IN GREAT BRITAIN

PRINTED IN GREAT BRITAIN
BY ROBERT MACLEHOSE AND CO. LTD.
THE UNIVERSITY PRESS, GLASGOW

SECULORS OF LOND SECTION ACT

(c'ur III of heeks)

lette or extended 127 129.

Lette or extended constant chemical constant place.

Lette sail constant place.

need of the control o

onto the control of t

The Horold of Lord Constant of the State of

andardisction of grassvers, see 412-410.

And the matter of farmenting

Apr. transport in Modern Springer, S

orniuszani ar mir zontest 6f, 12 Standard Joseph est ir 36: 361 Standardise est ir 36: 361 , bucultel 11 , papeter 41

-y volumer e heart, 416.
Standard vol tions, 117.
Standard builtie, 46.
The nale, together unities of, 41.
South as piper, and harmon 3+ 14.

Temperature of the second seco

The real philade of the control of t

Francis w. 121

Francis manus executes a grant in

that the other executes a grant in

that the other and than the

Estimated a second a second a

Estimated a second a second a

1 second area of the other and a second a

Internation to the according to the property of the property of the according to the accord

The many of the second of the

, see geroe p. . est o par no bloche.
Anno.
—, ilementions or of 10 elemention.
Antonimel base or of 10 elemention.
Antonimel base of the see

- Normality status.

The grant and the second secon

method, 117

CONTENTS
CHAPTER
I. Introductory
PARTI
APPARATUS AND PRINCIPLES USED IN MICRODIFFUSION ANALYSIS
II. A STANDARD MICRODIFFUSION APPARATUS OR 'UNIT'
III. FACTORS INFLUENCING THE ABSORPTION RATE FROM OUTER TO INNER CHAMBER WITH SPECIAL REFERENCE
TO AMMONIA
IV. GENERAL PRINCIPLES GOVERNING THE ABSORPTION TIME IN MICRODIFFUSION ANALYSIS
V. PIPETTES (SUITABLE FOR USE WITH THE STANDARD UNITS) AND THEIR DELIVERY ERRORS
VI. MICRO-BURETTES (SUITABLE FOR USE WITH THE STAN- DARD UNITS) AND ERRORS INVOLVED IN THEIR USE 47
VII THE MICRODIFFUSION METHOD WITH END-POINT VOL- UMES AROUND 20 CUBIC MILLIMETRES
VIII. COLORIMETRY IN THE MICRODIFFUSION METHODS . 75
Property in the control of the contr
PART II
DESCRIPTION OF METHODS WITH THE STANDARD UNITS
A. VOLATILE BASES
Aa. Ammonia Group
IX. Ammonia. General Method using Standard Acid as Absorbent
X. Ammonia. General Method (using the Boric-HCl Procedure)
XI. Special Factors Influencing the Rate of Ammonia
Absorption
XII. OTHER METHODS FOR DETERMINING THE ABSORBED AMMONIA IN THE MICRODIFFUSION PROCEDURE . 105
XIII AMMONIA BIOLOGICAL DETERMINATIONS
ATTI. MIMONIA. DIOLOGICAL INCLEMINATIONS

XIV. Total Nitrogen (over 100 μ g N).	PAGE 134
XIV. TOTAL NITROGEN (OVER 100 μ g N). XV. TOTAL NITROGEN (UNDER 100 μ g N) Further	134
Procedure	139
XVI. Total Nitrogen (1 to 0.1 μ g N)	154
XVII. UREA (BLOOD AND URINE)	162
XVIII. UREA IN TISSUES	175
XIX. Adenosinetriphosphoric Acid, Adenylic Acid,	
ADENOSINE, ETC.	178
XX. NITRATE, NITRITE AND AMIDE NITROGEN	184
XXI. AMIDES (continued). GLUTAMINE	187
XXII. Monoamine Oxidase and Histaminase in Tissues	190
Ab. Amine Group	
XXIII. DETERMINATION OF VOLATILE AMINES	195
B. Volatile Acids	
Ba. The Carbon Dioxide Group	
XXIV. CARBONATES AND BICARBONATE	201
XXV. BLOOD GLUCOSE AND FERMENTABLE SUGAR IN NORMAL URINE	215
XXVI. DETERMINATION OF CARBONIC ANHYDRASE	221
XXVII. OXIDATION RATES OF ORGANIC SUBSTANCES WITH A STANDARD OXIDANT WITH APPLICATION TO DETER- MINATION OF MINUTE AMOUNTS OF CALCIUM AS	
OXALATE	225
Bb. 1 Volatile Fatty Acids	
XXVIII. ACETIC ACID AND OTHER LOWER FATTY ACIDS .	234
XXIX. Assay of Acetylcholinesterase	239
Bc. Volatile Weak Inorganic Acids and Phenols	
XXX. CYANIDE, AZIDE, SULPHIDE, PHENOLS	241
C. Volatile Alcohols	
XXXI. METHANOL AND ISOPROPANOL GROUP	246
XXXII. ETHANOL	248
XXXIII. ETHANOL FROM URETHANE	256

D. VOLATILE ALDEHYDES

CHAPTER	Da. Formaldehyde Group	PAGE
XXXIV.	FORMALDEHYDE	258
XXXV.	FORMALDEHYDOGENIC STEROIDS (PERIODIC ACID	\
	AS OXIDANT)	262
XXXVI.	FORMALDEHYDOGENIC STEROIDS (SODIUM BIS-	222
	MUTHATE AS OXIDANT)	266
XXXVII.	GLYCINE (FORMALDEHYDE PRODUCED BY NIN- HYDRIN OXIDATION)	269
	Db. Acetaldehyde Group	
XXXVIII.	ACETALDEHYDE (SEMICARBAZIDE ABSORPTION) .	273
XXXIX.	ACETALDEHYDE FROM LACTIC ACID AND THREONINE	070
	WITH BISULPHITE ABSORPTION	276
	E. VOLATILE KETONES	
XL.	ACETONE (INCLUDING A RAPID CLINICAL METHOD USING THE NESSLER SOLUTION)	281
F. VARIO	OUS VOLATILE OXIDISING AND REDUCING SUBSTANCE	ES
	Fa. The Halogens	
XLI.	The Halogens (Introductory)	286
XLII.	CHLORIDE (BY OXIDATION TO CHLORINE AND ABSORPTION INTO IODIDE)	289
XLIII.	CHLORIDE (BY OXIDATION TO CHLORINE AND AB-	
	SORPTION INTO FAST GREEN)	301
XLIV.	Bromide	306
XLV.	IODIDES AND HALOGEN MIXTURES	316
XLVI.	SERIAL DETERMINATION OF ORGANICALLY BOUND HALOGEN .	320
VIVII	VOLATILE HALOGENATED HYDROCARBONS (CHLORO-	020
ALVII.	FORM, TRICHLORETHYLENE AND CARBON TETRA-	
	CHLORIDE)	326
	Fb. Carbon Monoxide	
XLVIII.	Carbon Monoxide	328

CONTENTS

XLIX. A Rapid Clinical Method for Carbon Monoxide Determination	PAGE
	000
G. Volatile Substances of Toxicological Interest	
L. FELDSTEIN AND KLENDSHOJ'S SYSTEM FOR THE DETERMINATION OF VOLATILE POISONS BY MICRO-	
DIFFUSION.	33 5
H. Other Miscellaneous Applications of the Microdiffusion Principle	
LI. Total Molecular Concentration in Fluid Samples of about 3-4 Milligrams .	340
LII. SEPARATION OF CRYSTALS AND 'GUMS' BY MICRO-DIFFUSION	343
QUALITATIVE MICRODIFFUSION ANALYSIS	
LIII. Some Considerations on Qualitative Micro- diffusion Analysis	345
$PART\ III$	
THE ERROR OF VOLUMETRIC TITRATION	
LIV. Introductory	349
LV. THE VARIABLE GLASS ERROR	356
LVI. THE VARIABLE GLASS ERROR (continued)	372
LVII. THE TOTAL VARIABLE GLASS ERROR AND ITS CON-	
	383
	395
	402
LX. THE CONSTANT GLASS ERROR	410
LXI. THE CONSTANT CHEMICAL ERROR	419
LXII. VOLUMETRIC ERROR IN KJELDAHL NITROGEN	
	429
	440
	451
INDEX OF SUBJECTS	461

LIST OF ILLUSTRATIONS

	PAGE
1. Plan of standard Conway 'unit'	8
la. Standard 'unit'	8
2. Standard 'unit' with 'units' No. 2 and 2A	10
2a. Plan of 'unit' No. 2	10
3A-F. Other forms of microdiffusion apparatus	12
3aG. Modified 'unit' after Öbrink	13
3aH. Kirk's distillation-diffusion 'unit'	14
3aI. Black's microdiffusion vessel	15
3aJ. Berka's modification; cross section	15
3aK. Berka's modification for high temperatures	15
4. Ammonia absorption curves in standard 'unit'	17
5. Further ammonia absorption curves in standard 'unit'	18
6. Rocking device for 'units' if required for special experiments (with the general use of the 'unit' it is not required).	22
7. Dale's oscillating table for Conway 'units'	23
8. Vibrating table	24
8a. Drawings of 1 ml. Ostwald and other pipettes from samples in	
use	34
9. Original form of Ostwald pipette	38
10. 2 ml. Bang burette, with soda-lime tube attachment	40
11. The Trevan syringe pipette	41
12. The Krogh-Keys syringe pipette	42
13. Conway microburette	49
14. Diagram of Conway burette (slightly modified after Wilson 1949)	50
15. Modified Conway burette (Harman and Webster, 1948)	52
16. Further microburettes of interest in connection with micro- diffusion methods	53
17. Rehberg burette	55
18. Curves of variable error in titrating 1 ml. hydrochloric acid solution with carbon dioxide free alkali	61
19. Ramsay's modification of the Conway burette for titration with titanous sulphate	64
20. McFarlane's modification of the Conway burette for titration	1 17
with titanous salts	65
21. Titration arrangement and small wax 'unit' for minute ammonia determinations (order of $0.1 \mu g \ NH_3-N)$.	68

	PAGP
22. The completed absorption tube and titration arrangement as used by Linderstrøm-Lang and Holter	72
23. Arrangement for fine pipetting (order of 10 c.mm.) by Linder-strøm-Lang and Holter	73
24. Construction principle of the Duboscq colorimeter (Leitz)	76
25. Compensation arrangement for a coloured solvent	77
26. Diagram illustrating an essential principle of the Pulfrich photometer	84
27. Diagram of optical arrangement in the Pulfrich photometer .	84
28. The optical arrangement and photoelectric circuit of the Spekker Absorptiometer	86
29. Times for full $(99\cdot5\%)$ absorption in standard 'unit' (No. 1) and No. 2	93
30. Effect of carbon dioxide on ammonia formation in human blood after shedding	118
31. Characteristic curves of the blood ammonia for man (crosses) and rabbit (dots) after shedding into an open flask. Room temperature. Each point is the median of consecutive samples of eleven observations ranged in order of time after shedding	119
31a, b, c. Serial curves of blood ammonia for man, rabbit, and fowl	119
32. Diagram of ammonia metabolism	125
32a. Blood-ammonium curves in tolerance test (Sherlock et al) .	126
32b. Further blood-ammonium curves in tolerance test	127
32c. Digestion rack and Kjeldahl tubes as used by Borsook and Dubnoff	139
33. Dimensions of 'unit' as used by Borsook and Dubnoff .	140
33a. Titration apparatus as used by Borsook and Dubnoff .	141
33b. Electrode assembly details of titration apparatus used by Borsook and Dubnoff	142
34. Apparatus used by Needham and Boell for combined digestion and microdiffusion	143
35. Apparatus used by Tompkins and Kirk for combined digestion and microdiffusion	144
36. Sectional view of microdiffusion apparatus as used by Hawes and Skavinski	145
37. Apparatus used in the technique of Linderstrøm-Lang and associates	155
38. Further apparatus connected with the technique of Linderstrøm-Lang and associates	158

LIST OF ILLUSTRATIONS	xvii
artina juniture i na transferie garaga se a sur	PAGE
39. Franke's lancet and collecting tube for blood urea determination	162
40. pH effect on rate of urea hydrolysis by urea-phosphate solution	168
41. Salt (phosphate) effect on rate of urea hydrolysis by urease .	168
42. Distribution of normal human blood urea concentration .	170
43. Distribution of urine urea concentrations in the normal human subject. Water drinking to produce diuresis excluded	173
44. Distribution of urine volumes for the normal human subject. Water drinking to produce diuresis excluded	174
45. Ammonia absorption curves from amides after Hallam.	185
46. Further absorption curves after Hallam	185
 Rates of absorption of amines. I, ammonia. II, iso-butyl-amine. III, methylamine. IV, iso-amylamine. V, ethylamine. VI, benzylamine. At 37° C. in standard 'unit'. 	197
48. Absorption rates of CO ₂ in the microdiffusion 'units'.	203
49. Titration in a stream of carbon dioxide free air.	208
50. Separating funnel and bottle with moistened glass beads for saturating plasma with alveolar air	209
51. Lehmann's microdiffusion procedure for plasma bicarbonate, showing the introduction of plasma into the outer chamber	211
52. Lehmann's microdiffusion procedure for plasma bicarbonate, showing the procedure for titrating with 0·1 N HCl .	211
52a. Increased losses of CO_2 with increasing concentration of carbonic anhydrase as described in text	223
53. Oxidation rates of organic substances with a standard permanganate oxidant	230
54. Murnaghan's procedure for determining Ca content of small volumes of serum	231
55. Absorption rates of ${\rm CO}_2$ after oxalate oxidation	232
56a, b, c. Modified Conway 'unit' after Edwards and Kellie .	259
57. Rack and holder used for handling Conway 'units' after Schwartz et al	270
58. Glycine standard curve after Schwartz $et\ al$	272
59. Nomograph for clinical determination of acetone (Werch)	284
60. Effect of permanganate concentration on chloride oxidation rate in 'unit'.	297
61. Effect of varying the acid concentration on chloride oxidation	298
62. Effect of dichromate concentration on bromide oxidation $$.	315
62a. Effect of varying the acid concentration on bromide oxidation	315
63. Micro-Carius furnace after Pirt and Chain	321
$64.\ $ Chlorine determination in organic substances (Pirt and Chain) .	324
65 Bramina determinations in organic substances (Pirt and Chain)	394

xvii	ii LIST OF ILLUSTRATIONS	PAGE
66.	Iodine determinations in organic substances (Pirt and Chain) .	325
67.	Extinction values in Burgen's blood chloroform method, 1948	325
68.	Carboxyhaemoglobin determination according to Lehmann .	333
69.	Separation of 'gums' and crystals, older method	343
70.	Cells for qualitative microdiffusion analysis	346
71.	The curve of normal distribution	351
72.	Curves showing the summation of equal and uncorrelated rectangular distributions around a mean	354
73.	Drainage curves of a standard 100 ml. pipette from Stott .	361
74.	Curves A-D give drainage rates of a 50 ml. burette after deliveries of 50 ml. in 18, 40, 60 and 120 seconds. Curves from data of Lindner and Haslwanter	377
75.	Device for facilitating meniscus reading; medified from Kolthoff's Die Massanalyse	379
76.	Theoretical curves and experimental data illustrating the variable error in titrating 1 ml. acid with 1 ml. of carbon dioxide free alkali from 2 ml. Bang burette	406
77.	Curves of volumetric error in the classical Kjeldahl nitrogen	

78. Effect of urine volume on urea excretion in the human subject

79. Distribution of urea normality ratios for the normal human

determination.

subject

433

444

448

CHAPTER I

INTRODUCTORY

In the development of modern biological research large numbers of observations become increasingly necessary in determining the questions at issue. It becomes essential to have analytical methods of a micro kind in which, without sacrifice of accuracy with respect to macro procedure, the labour and time expended with the single analysis are reduced to a minimum.

In the search for such a method, primarily for ammonia and urea, the microdiffusion principle of analysis was elaborated here and a special diffusion apparatus and burette designed to give it effect. It was subsequently found applicable to a variety of micro determinations by the author and other workers.

The method in general would appear to be the simplest possible consistent with the maximum attainable accuracy in the handling of micro volumes. Distillation and aeration are eliminated, the passage of ammonia or other substance taking place by diffusion from one chamber, in which it exerts a certain tension, into an absorbing fluid in another chamber in which its tension is reduced to zero. With a knowledge of the diffusion conditions controlling this passage and used in the design of the standard microdiffusion apparatus or unit, full absorption times with many substances run from about half an hour to two hours.

With serial determinations the time expended on each determination need be only a few minutes; one of the special advantages of the method being the ease with which large numbers of accurate data can be assembled.

With the standard unit, the accuracy of ammonia and consequently, of urea and other determinations, is limited only by the accuracy of delivering and titrating fluid volumes of the order of 1 ml. With the development of the titration principles described in the text, this accuracy can be brought to any desirable level in practice, so that with comparative ease the percentage error need