Introduction to Analytical Gas Chromatography

History, Principles, and Practice

John A. Perry

Introduction to ANALYTICAL GAS CHROMATOGRAPHY

History, Principles, and Practice

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On the invitation of Dr. Jay Curtice, now Chairman of the Department of Chemistry of Roosevelt University, in the spring of 1964 I began teaching a 16-week graduate-level evening credit course in gas chromatography. Since then it has been presented at least once each year.

Responding to my questionnaires, students in the course have described themselves as full-time working chemists. Ranging in age from 21 to over 70 years, they have an average age of perhaps 28. Usually, they have earned at least a bachelor's degree, have not had experience in gas chromatography, and have registered only for this course.

This book was written for people such as these, to provide them with orientation and insight in language they find clear and comprehensible. For perspective, a topic is often presented against its history. Throughout, reasons and principles are stressed.

The book is selective in choice of topic and depth of treatment. For each topic, once it has been chosen, the relevant technical background is furnished. The topic is then developed in a way that is understandable to readers usually thoroughly ill at ease with mathematics and electrical circuitry. In discussions, for instance, names always accompany symbols. Derivations can be read.

Because the treatment is cumulative, the book should be read from front to back. However, the chapters on qualitative and quantitative analysis (Chaps. 13 and 14) can be understood without first reading the chapter on derivatization.

I acknowledge the friendly and effective assistance of Miss Anne O'Donnell and of Dr. Peter Klein in the treatment of gas chromatography-mass spectrometry. Mr. E. M. Bens reviewed the chapter on supports, helped clarify and strengthen the section on the preferred diatomaceous earth, and furnished additional valuable (and beautiful) photographs. Dr. Mathieu H. J. van Rijswick reviewed the chapter on quantitative analysis and commented generously and constructively on my treatment of his high-precision algorithm. I thank both of these experts for their help.

vi Preface

I thank Miss Bernadine Palka, who has read the complete text several times, suggested many improvements, caught innumerable errors. The book is the better for her.

I am deeply grateful for the manifold cooperation of the Regis Chemical Company of Morton Grove, Illinois. And my gratitude to my wife, Inger, is boundless.

John A. Perry

NOTATION

For each symbol, the location of its definition or first appearance follows its explanation.

a 1. Constant expressing fraction of corrected retention volume that may be used as the maximum acceptable sample volume (pages 9, 28):

$$V_{\text{max}} = a \frac{V_{\text{M}}^{\circ} + KV_{\text{L}}}{\sqrt{n}} = a \frac{V_{\text{R}}^{\circ}}{\sqrt{n}}$$

- 2. Constant in equation for vapor pressure p° (Chap. 11 only). P. 224, Eq. 11.3.
- A The eddy diffusion term of the Jones-van Deemter equation. P. 6.
- ADC Analog-to-digital converter. P. 362.
- A_{\min} The minimum detectable peak area as a function of digital noise. P. 377, Eq. 14.10.
- b Constant in equation for vapor pressure p°. P. 224, Eq. 11.3.
- B The axial molecular diffusion term of the van Deemter equation. P. 7.
- c Constant in equation for vapor pressure p° (Chap. 11 only). P. 224, Eq. 11.3.
- $c_{_{T_{\rm c}}}$ Constant in the $c_{_{T_{\rm c}}}$ term of the Jones-van Deemter equation. P. 86, Eq. 6.30.
- Constant in the C₂ term of the Jones-van Deemter equation. P. 92, Eq. 6.63.
- c1...c12 Constants.
- The overall term that expresses resistance to mass transfer in the Jonesvan Deemter equation. P. 8.
- This symbol, cited on p. 232 in a quotation, has the meaning of our C_{G} .
- C_{lig} This symbol, cited on p. 232, in a quotation has the meaning of our C_{LP} .
- The weight-per-volume concentration in the mobile phase of solute at the peak apex (page 346, Eq. 14.1):

$$C_{\text{max}} = \frac{\sqrt{n}}{V_{\text{p}}^{n}} w_{\text{I}} \sqrt{2\pi}$$

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The solute concentration at time t following injection at concentration C_0 into an insufficiently hot injection port that produces exponential rather than plug sample introduction. P. 199. Eq. (10.16).

- C_D The maximum solute weight-per-volume concentration in the mobile phase that does not exceed the linear dynamic range of the detector. P. 346.
- C_G The term of the Jones-van Deemter equation that expresses resistance to mass transfer in the gas phase. P. 91. Eq. (6.54).
- C_L The weight-per-volume concentration of solute in the liquid stationary phase. P. 20. P. 21, Eq. (2.8).
- C_{LP} The term of the Jones-van Deemter equation that expresses resistance to mass transfer in the liquid stationary phase. P. 88, Eq. (6.40).
- C_M The weight-per-volume concentration of solute in the gas mobile phase. P. 21, Eq. (2.8).
- C_O Initial sample concentration on sample injection. P. 199, Eq. (10.16).
- C₂ The term of the Jones-van Deemter equation that expresses resistance to mass transfer due to velocity distribution in the mobile phase. P. 92, Eq. (6.64).
- C $_3$ The term of the Jones-van Deemter equation that expresses interaction of the nominally independent $\rm C_G$ and $\rm C_2$ terms. Not a very important term.
- d Internal diameter of an open tubular column. P. 214.
- d_ Particle diameter. P. 6
- The particle diameter as used in the reduced plate-height equation. In the case of gas-liquid chromatography, d* refers to the pore size of the inert support rather than to the inert support particle diameter. P. 111, Eq. (7.13).
- d_{C} The internal diameter of a column. P. 9.
- d_G The effective thickness of the gas phase; the shortest distance through the carrier gas from one surface of the stationary phase to another. P. 89, Eq. (6.45).
- d_{T} The effective thickness of the liquid stationary phase. P. 8.
- db Decibels. A decibel is equal to 10 times the base-10 logarithm of a power or rejection ratio. For example, 60 db implies a ratio of one million.
- D Diffusivity; dimensions: (distance) 2/time. Defined p. 199, Eq. (10.18).

 The diffusivity is identically the diffusivity coefficient; see p.78, Eq. (6.1).
- Detectability: the smallest quantity that will cause a detector response equal to twice the noise level. P. 143.
- D_m The diffusivity of the mobile phase, as used in the equation for reduced plate height. P. 111, Eq. (7.15).
- D_{G} The diffusivity of the carrier gas. P. 84, Eq. (6.21).
- D. The diffusivity of the liquid stationary phase. P. 8.
- e The base of the natural logarithms. First used, p. 171.
- e Bucking voltage. This is the voltage that is developed by a potentiometric recorder within itself to equal the signal being measured, so that the difference (e $_{\rm b}$ e $_{\rm i}$) approaches zero as the recorder approaches balance. P. 352.
- e, The voltage of a source of potential. P. 151.

e i-span An electric potential just sufficient to cause a full-scale pen deflection of a potentiometric recorder. P. 354.

- e The voltage delivered by a voltage divider.
- e A reference voltage within a potentiometric recorder in terms of which an incoming voltage is measured. P. 352.
- E Electron affinity, pertaining to electron capture. P. 171.
- E_d Dispersion interaction energy. P. 57.
- E, Dipole-induced dipole interaction energy. P. 57.
- E* The concentration of an electron-capturing species in an electron capture detector. P. 171.
- F Flow rate of column effluent; p. 145. Gas flow rate entering mixing chamber; p. 199, Eq. (10.17). And gas flow rate, p. 240.
- F_a Flow rate of added gas in flame ionization detector, p. 145 (see also V_e , effective detector volume, p. 145).
- $_{\rm C}^{\rm F}$ The temperature-corrected carrier gas flow rate, measured at the column exit pressure and corrected to column temperature. P. 18.
- h The height equaivalent to a theoretical plate: h = L/n. Longer symbol: HETP. P. 5, Fig. 1.5.
- h The reduced plate height. P. 111, Eq. (7.13).
- H The height equivalent to an effective theoretical plate: H = L/N. Longer symbol: HEETP. P. 213.
- Differential heat of vaporization of solute from infinitely dilute solution. P. 225. Eq. (11.7).
- ΔH_{v} Heat of vaporization of the pure organic liquid. P. 224, Eq. (11.4).
- Hz Hertz: one cycle per second.
- i Current of electricity.
- I Kovats Retention Index. P. 60. Eq. (5.1).
- I_b The standing current of an electron capture detector. P. 165.
- The current observed in an electron capture detector while an electroncapturing species is in the detector. P. 171.
- $I_{\mathrm{PT}(i)}$ Kovats Retention Index of substance i from programmed temperature data. P. 246, Eq. (11.30).
- ${\bf I_v}$ Ionization energy for species X. P. 57.
- ΔI Kovats Retention Index difference. P. 60.
- j Pressure gradient correction factor. P. 19.
- k Capacity factor: $k = K/\beta = p/q = t'_R/t_M$. Also called partition ratio or, less frequently, the mass distribution ratio. P. 22. Eq. (2.10).
- k_B Boltzman constant. P. 56.
- $k_{_{\mbox{\scriptsize TC}}}$ Coefficient of thermal conductivity. P. 149, Table 9.1.
- Δk_{mc} Thermal conductivity coefficient difference. P. 149, Table 9.1.
- K Partition coefficient: $K = C_L/C_M = (w_L/V_L)/(w_M/V_M) = (w_L/w_M)(V_M/V_L) = k\beta$. Also called distribution coefficient. Pages 21, 22.
- Ka Adsorption partition coefficient. P. 59.

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- K_H Henry's Law constant. P. 225, Eq. (11.7).
- K* Coefficient of electron capture. P. 171.
- A distance, the travel through which causes a given type of peak spreading during molecular "random walking." P. 83. Eq. (6.14).
- L Column length. See Fig. 1.5, p. 5.
- m 1. Constant.
 - 2. In the filtration method of packing preparation, the ratio m of solution volume retained on the support to the weight of the support: $V_S = mW_S$. P. 182; see also p. 183, Eq. (10.3).
- M Subscript; refers to mobile phase. P. 20.
- n l. Number of theoretical plates: $n = 16 (t_p/w_h)^2$. P. 4.
 - Number of carbons in an homologous series (Chap. 11 only). P. 224, Eq. (11.1).
 - 3. In the filtration method of packing preparation, the ratio n of solution volume $\mathbf{V}_{\mathbf{S}}$ retained on the support to the total volume $\mathbf{V}_{\mathbf{T}}$ of the solution (Chap. 10 only). P. 182, Eq. (10.4).
- Number of opportunities for causing broadening. Chap. 6 only. See, for instance, p. 83, Eq. (6.15).
- n₁, n₂ Numbers of interdiffusing molecules in derivation of Einstein's Law of Diffusion. (Chap. 6). P. 79. Eq. (6.2).
- N 1. Number of theoretical effective plates: $N = 16 \left(\frac{t'_R}{w_b}\right)^2 = n/[k/(1+k)]^2$. P. 41, Eqs. (4.9), (4.12).
 - Net number of molecules crossing area O in derivation of Einstein's Law of Diffusion (Chap. 6 only). P. 79. Eq. (6.2).
- $N_{_{
 m C}}$ Effective carbon number: the number of carbons a n-paraffin would have to have to yield the same response from a flame ionization detector as a given molecular species. For n-heptane, $N_{_{
 m C}}=7.00$. P. 161.
- The designation and magnitude of an area, in the derivation of Einstein's Law of Diffusion. P. 78. Eq. (6.1).
- p 1. The weight fraction of solute in the stationary phase. P. 22. Eq. (2.11).
 - 2. Partial pressure (Chap. 11 only). Pages 226, 227.
- p° Standard vapor pressure for a given molecular species. P. 224. Eq. (11.3).
- Δp_{no} Pressure drop necessary for a separation. P. 214. Eq. (10.28).
- P₁, P₂ Equivalent grams of stationary phase on supports 1 and 2, in equivalent loading. P. 129. Eq. (8.1).
- P Carrier gas pressure at the inlet of a column. P. 19.
- P Carrier gas pressure at the outlet of a column. P. 19.
- PP Performance parameter: $PP = \Delta p_{ne} t$. P. 214. Eq. (10.28).
- q The weight fraction of solute in the mobile phase. P. 22. Eq. (2.11).
- r l. Electrical resistance. P. 151.
 - 2. Internal radius of column (Chap. 10 only), P. 190. Eq. (10.14).
 - Heating rate in programmed temperature gas chromatography (Chap. 11 only).
 P. 240.
 - 4. Effective distance of molecular separation (Chap. 5 only). P. 57.

χV

R Gas constant. P. 224. Eq. (11.4).

 R_c Resolution. P. 41. Eq. (4.13).

$$R_{S} = \frac{\sqrt{N}}{4} \frac{\alpha - 1}{\alpha}$$

R_T Intrinsic resolution. P. 245. Eq. (11.29).

 R_{TT} Isothermal intrinsic resolution. P. 244. Eq. (11.27).

R_{T.} Linear dynamic range of a detector. P. 347.

[R] The time-dependent resolution of Struppe. P. 215. Eq. (10.33).

S The sensitivity of a detector in grams per second. P. 143.

SF The Glueckauf separation factor. P. 42; see Fig. 4.2.

- t 1. The time for a molecule of diffusivity D to diffuse distance Δ : D = $\Delta^2/2t$. Eq. (6.4), p. 79.
 - The time interval after sample injection (Chap. 10 only). P. 199, Eq. (10.16).

t The time at which a peak is found to end. P. 377, Eq. (14.11).

t_c The time at which a peak is found to start. P. 377, Eq. (14.11).

t ne The minimal time necessary for a separation. P. 214, Eq. (10.28).

 t_G The time spent by a solute molecule in the gas phase while passing through a column. P. 84. Eq. (6.23).

t_L The time spent by a solute molecule in the liquid stationary phase while passing through a column. P. 86. Eq. (6.31).

The gas holdup time: The elapsed time from sample injection to peak apex of an unretained solute. P. 17.

 t_{M}° The time from sample injection to peak apex of an unretained solute, corrected for pressure gradient: $t_{M}^{\circ} = j t_{M}^{\bullet}$.

 t_N The net retention time: $t_N = t_R^{\circ} - t_M^{\circ}$.

 $t_{\rm R}$ The time from sample injection to peak apex; the unadjusted, uncorrected, observed retention time. P. 4.

 t_R' The adjusted retention time: $t_R' = t_R - t_M$. P. 24.

 t_R° The corrected retention time: $t_R^{\circ} = j t_R^{\circ}$.

T Absolute temperature. P. 171.

In Eq. (14.6), p. 348, the isothermal temperature for elution, following sample injection at a lower temperature T_0 .

T_b Boiling point. See Trouton's rule. P. 228, Eq. (11.18).

- 1. In programmed temperature gas chromatography, the initial temperature.

 This is the column temperature at the beginning of a temperature program and may be the column temperature on sample injection. P. 240. Eq. (11.23)
 - In Eq. (14.6), p. 348, the injection temperature of a cold column or precolumn, as used in enhancement of trace detection.
- Tr In programmed temperature gas chromatography, the retention temperature. This is the column temperature at which a solute is eluted during a temperature program. P. 240. Eq. (11.23).
- T' In programmed temperature gas chromatography, the significant temperature, at which a programmed temperature process is equivalent to an isothermal.
 P. 241.

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u 1. Carrier gas velocity, measured. P. 87. Eq. (6.34).

- 2. Dipole moment (Chap. 5 only). P. 57.
- a Average carrier gas velocity. P. 4.
- $\overline{\mathbf{u}}_{\text{o}}$ Optimum carrier gas velocity. P. 5, Fig. 1.5.
- u Optimum carrier gas velocity. P. 98. Eq. (7.3).
- \mathbf{u}_{L} The velocity of a peak apex along a column. P. 86. Eq. (6.33).
- u_S The velocity of a peak apex with respect to the carrier gas. P. 90. Eq. (6.50).
- V In the equation for reduced plate height, the reduced velocity: $v = d_n^* \text{ u/D}_m$. P. 111. Eq. (7.14).
- V 1. Retention volume. P. 240.
 - 2. Mixing chamber volume (Chap. 10 only). P. 199. Eq. (10.17).
 - Effective volume of a flame ionization detector without added gas (Chap. 9 only). P. 145.
- V_a The isothermal retention volume for elution temperature T_a . P. 348. Eq. (14.6).
- $V_{_{\rm P}}$ The effective internal volume of a detector. P. 145.
- $_{
 m eff}^{
 m V}$ The effective internal volume of a detector (same meaning as $_{
 m e}^{
 m V}$). P. 203. Eq. (10.21).
- V The specific retention volume. P. 224. Eq. (11.2).
- That volume of a sample that will not cause any more than 10% peak broadening with respect to any smaller volume of it. $V_{max} = a \ V_R^0 / \sqrt{n}$, but the "constant" a is determined by experiment, namely, by injecting successively smaller samples until resolution no longer improves with further decrease in sample size. P. 9.
- The isothermal retention volume for elution temperature T_r (see Eq. (14.6)). The elution temperature is usually an arbitrarily chosen temperature for isothermal elution from a column used as a cold trap; the elution temperature is thus to be distinguished from the retention temperature, which is associated with a temperature program. P. 243.
- $\rm V_F$ The filtrate volume (V $_T$ V $_S$) of solution from filtration impregnation. P. 184. Eq. (10.12).
- Volume of stationary phase in the column. P. 9. See also p. 24, Eq. (2.21).
- $\rm V_{M}$ The retention volume for air, uncorrected for gas compressibility. It is used in determining $\rm V_{R}^{\bullet}.$ P. 19. Eq. (2.2).
- V_M° The column dead volume: $V_M^{\circ} = j V_M^{\bullet}$. P. 20. Eq. (2.4).
- VN The net retention volume, adjusted for column gas volume and corrected for gas compressibility: $V_N = V_N^\circ V_N^\circ$. It is the net retention volume that is directly and linearly related to the stationary phase volume V_L by the partition coefficient K: $V_N = K \ V_L$. P. 20. Eq. (2.6).
- V_R The unadjusted, uncorrected, retention volume. $V_R = F_o t_R$. P. 19. Eq. (2.1)
- V'_R The adjusted retention volume, uncorrected for gas compressibility: $V_R^i = V_R^i V_M^i$. P. 20. Eq. (2.5).
- V_P The corrected retention volume. P. 19. Eq. (2.3).
- V_S In filtration impregnation, the volume of solution that is retained on the support. P. 182. On p. 184, Eq. (10.12).
- ${
 m V}_{
 m T}$ In filtration impregnation, the total volume of solution used. P. 184, Eq. (10.12).

Notation xvii

 ${\rm V}_{_{\rm T\!P}}$ Retention volume measured isothermally at retention temperature. P. 244.

- V₀ The isothermal retention volume corresponding to a column temperature T₀ on sample injection. P. 243. See also Eq. (14.6), p. 348.
- w Weight.
- w_b Peak width at base, where the base is defined as the distance between the intercepts of the peak tangents with the interpolated baseline. P. 4.
- W_f The width of a digital matched filter. P. 375.
- w The peak width detected as part of the van Rijswick algorithm. P. 377.
- W_L The weight of stationary phase in a column. P. 24, Eq. (2.21).
- $W_{_{\mathrm{D}}}$ In filtration impregnation, the weight of the packing. P. 182. Eq. (10.1).
- $W_{\rm c}$ In filtration impregnation, the weight of the support. P. 182. Eq. (10.1).
- $W_{\mathrm{T,L}}$ In filtration impregnation, the total weight of stationary phase to be used. P. 183. Eq. (10.11).
- WD The maximum weight of a given solute that may be injected without exceeding the linear dynamic range of the detector. P. 346, Eq. (14.2). P. 347, Eq. (14.3).
- $w_{\rm L}$ Weight of solute dissolved in the stationary phase: $k = w_{\rm L}/w_{\rm M}$. P. 22, Eq. (2.9).
- W_{M} Weight of solute in the mobile phase: $k = W_{T}/W_{M}$. P. 22, Eq. 2.9.
- $W_{\rm S}$ The minimum detectable weight of a trace component. P. 347. Eq. (14.4).
- x In filtration impregnation, the weight fraction of stationary phase in a packing. P. 183, Eq. (10.2).
- Z Carbon number of a n-paraffin, in connection with the Kovats Retention Index. P. 60. Eq. (5.1).
- Z* The temperature-independent factor in the equation describing the temperature dependence of the electron capture coefficient. P. 171.
- α Relative retention. $\alpha = t_{R2}^{1}/t_{R1}^{1} = K_{2}/K_{1} = k_{2}/k_{1}$. P. 24. Eq. (2.19).
- α Polarizability. P. 57.
- β Phase ratio, also called column characteristic. β = $V_{\mbox{\scriptsize M}}/V_{\mbox{\scriptsize L}}.$ P. 22. Eq. (2.12).
- γ Tortuosity factor. P. 84. Eq. (6.23).
- δ Solubility parameter. P. 70.
- λ Packing irregularity factor. P. 83. Eq. (6.16).
- $\rho_{T.}$ Density of stationary phase (in bulk). P. 24. Eq. (2.21).
- σ Standard deviation, the square root of the variance. P. 80.
- $\sigma_{\rm T}^2$ 1. The total variance, which is the sum of the variances arising from independent causes. P. 80. Eqs. (6.5) and (6.6).
 - The plate height contribution from the effective internal volume of a detector (Eq. (10.21) only). P. 203.
- Response time, p. 199, Eq. (10.16). This is identically the time constant used in Eq. (10.20), p. 199; and is also the effective time interval in Eq. (10.21), p. 203. The concept is the same in each case.
- τ_{D} The time constant for concentration decrease at the entrance to a mixing chamber. P. 199, Eq. (10.18).
- τ_G The typical time required for a solute molecule to diffuse through the mobile phase to a gas-liquid interface. P. 88. Eq. (6.42).

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	The	typical	. time	requ	ired	for	a	solute	molec	ule	to	diffuse	through	the
	stat	tionary	phase	to a	gas-	-liqu	iid	inter	face.	P.	86.	Eq. (6.30).	

- τ_{M} The time constant for concentration decrease at the exit of a mixing chamber. P. 199. Eq. (10.17).
- τ_2 The time during which unit peak spreading due to velocity distribution takes place. P. 91. Eq. (6.56).
- 1. The distance diffused in time t according to Einstein's Law of Diffusion: $D = \Delta^2/2t$. P. 79. Eq. 6.4. See also p. 78.
 - Sampling interval (Chap. 14 only). P. 377, Eq. (14.7); p. 377, Eq. (14.8); and p. 377, Eq. (14.10).
- Δp Pressure required to produce a desired carrier gas flow. P. 180.

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GAS CHROMATOGRAPHY: A SIMPLE DESCRIPTION

The word "chromatography," formed from the Greek "chroma" meaning "color," and "graphein" meaning "to write," was coined by Tswett [1, 2] around 1900 to describe his process of separating mixtures of plant pigments. He washed the pigments down a column of adsorbent powder, thus separating them into colored (chroma) bands (graphein) on the powder. Such a separation of the components of a mixture for qualitative or quantitative analysis, or for isolation and recovery of the components, is the desired end of any type of chromatography.

In gas chromatography (Fig. 1.1), the mixture (solute) to be separated is vaporized and swept over a relatively large absorbent or adsorbent surface inside a long narrow tube, or column. The sweeping is done by a steady stream of inert "carrier" gas, which serves only to move the solute vapors along the column. The different components are moved along the column at different rates and, under proper circumstances, become separated. The analyst arranges matters so that the separation is complete within a minimum time and can then be detected and recorded.

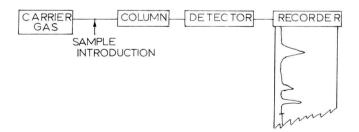


FIGURE 1.1 The gas chromatograph usually consists of a regulated supply of carrier gas, a means for sample introduction, a column for separating the components of the sample, a detector for detecting and signaling the components as they emerge in sequence from the column, and a recorder for measuring and recording the signal from the detector.