

Selected Papers
on
Neuro-Physiology

(From "Journal of General Physiology")

1961

CONTENTS

(from "Journal of General Physiology")

Relation of Function to Diameter in Afferent Fibers of Muscle Nerves.	
Hunt C. C. 33(1):117, 1954.....	1
A Theory of Taste Stimulation.	
Beidler L. M. 33(2):133, 1954.....	10
Bioelectric Effects of Ions Microinjected into the Giant Axon of Loligo.	
Grundfest H., Kao C. Y. and Altamirano M. 33(2):245, 1954.....	28
Microanatomy of the Abdominal Stretch Receptors of the Crayfish (<i>Astacus Fluviatilis</i> L.).	
Florey E. and Florey E. 39(1):69, 1955.....	61
Processes of Excitation in the Dendrites and in the Soma of Single Isolated Sensory Nerve Cells of the Lobster and Crayfish.	
Eyzaguirre C. and Kuffler S. W. 39(1):87, 1955.....	73
Further Study of Soma, Dendrite and Axon Excitation in Single Neurons.	
Eyzaguirre C. and Kuffler S. W. 39(1):121, 1955.....	111
Synaptic Inhibition in an Isolated Nerve Cell.	
Kuffler S. W. and Eyzaguirre C. 39(1):155, 1955.....	144
The Parallelism between the Action Potential, Action Current and Membrane Resistance at a Node of Ranvier.	
Tasaki I. and Freygang W. H., Jr. 39(2):211, 1955.....	174
Inhibition in the Eye of Limulus.	
Hartline H. K., Wagner H. G. and Ratliff F. 39(5):651, 1956.....	187
Vagal and Sympathetic Effects on the Pacemaker Fibers in the Sinus Venosus of the Heart.	
Hutter O. F. and Trautwein W. 39(5):715, 1956.....	210

The Relations between Prepotential, Resting Potential and Latent Period in Frog Muscle Fibers.	
Jenerick H. P. 39(5):773, 1956.....	229
The Non-Correlation of Bioelectric Potentials with Ionic Gradients.	
Shaw F. H., Simon S. E. and Johnstone B. M. 40(1):1, 1956.....	244
The Responses of the Pupil of Gekko Gekko to External Light Stimulus.	
Denton E. J. 40(2):201, 1956.....	261
The Nature of the Gecko Visual Pigment.	
Crescitelli F. 40(2):217, 1956.....	281
The Excitable Properties of Three Types of Motor Axons.	
Adelman W. J., Jr. 40(2):251, 1956.....	296
Intracellular Recording from the Giant Synapse of the Squid.	
Bullock T. H. and Hagiwara S. 40(4):565, 1957.....	306
Maintained Activity in the Cat's Retina in Light and Darkness.	
Kuffler S. W., FitzHugh R. and Barlow H. B. 40(5):683, 1957.....	321
Steps in the Production of Motoneuron Spikes.	
Fuortes M. G. F., Frank K. and Becker M. C. 40(5):735, 1957.....	341
The Effects of Hydrostatic Pressure upon the Normal and Narcotized Nerve Fiber.	
Spyropoulos C. S. 40(6):849, 1957.....	359
Demonstration of Two Stable Potential States in the Squid Giant Axon under Tetraethylammonium Chloride.	
Tasaki I. and Hagiwara S. 40(6):859, 1957.....	368
The Statistical Detection of Threshold Signals in the Retina.	
FitzHugh R. 40(6):925, 1957.....	396
The Effect of Temperature, Potassium and Sodium on the Conductance Change Accompanying the Action Potential in the Squid Giant Axon.	
Amatniek E., Freygang W., Grundfest H., Kiebel G. and Shanes A. 41(2):333, 1957.....	419
Blue-Light and the Regeneration of Human Rhodopsin in Situ.	
Rushton W. A. H. 41(2):419, 1957.....	429

The Rhodopsin System of the Squid.	
Hubbard R. and St. George R. C. C.	41(3):501, 1958..... 439
Effects of External Ions on Membrane Potentials of a Lobster Giant Axon.	
Dalton J. C.	41(3):529, 1958..... 467
An Analysis of Extracellular Potentials from Single Neurons in the Lateral Geniculate Nucleus of the Cat.	
Freygang W. H., Jr.	41(3):543, 1958..... 481
A Statistical Analyzer for Optic Nerve Messages.	
FitzHugh R.	41(4):675, 1958..... 503
The Refractory State of the Generator and Propagated Potentials in a Pacinian Corpuscle.	
Loewenstein W. R. and Altamirano-Orrego R.	41(4):505, 1958..... 521
Generator Processes of Repetitive Activity in a Pacinian Corpuscle.	
Loewenstein W. R.	41(4):525, 1958..... 541
Facilitation by Previous Activity in a Pacinian Corpuscle.	
Loewenstein W. R.	41(4):547, 1958..... 562
The Effect of Illumination on the Electrical Conductance of Rhodopsin Solutions.	
Hara T.	41(5):857, 1958..... 572
Spatial Summation of Inhibitory Influences in the Eye of Limulus and the Mutual Interaction of Receptor Units.	
Hartline H. K. and Ratliff F.	41(5):1049, 1958..... 593
Synaptic Potential in the Motor Giant Axon of the Crayfish.	
Hagiwara S.	41(6):1119, 1958..... 611
The Spectral Sensitivities of the Dorsal Ocelli of Cockroaches and Honeybees (An Electrophysiological Study).	
Goldsmith T. H. and Ruck P. R.	41(6):1171, 1958..... 621
Effects of Potassium, Sodium and Azide on the Ionic Movements That Accompany Activity in Frog Nerves.	
Asano T. and Hurlbut W. P.	41(6):1187, 1958..... 636

The Sites for Mechano-Electric Conversion in a Pacinian Corpuscle.	
Loewenstein W. R. and Rathkamp R. 41(6):1243, 1958.....	653
Electrical Activity in the Chemoreceptors of the Blowfly,	
1. "Responses to Chemical and Mechanical Stimulation".	
Wolbarsht M. L. and Dethier V. G. 42(2):393, 1958.....	674
Electrical Activity in the Chemoreceptors of the Blowfly,	
2. "Responses to Electrical Stimulation".	
Wolbarsht M. L. 42(2):413, 1958.....	694
The Influence of High Hydrostatic Pressure on Cocaine and Veratrine Action in a Vertebrate Nerve.	
Gershfeld N. L. and Shanes A. M. 42(3):647, 1959.....	710
Recurrent Facilitation of Spinal Reflexes.	
Wilson V. J. 42(4):703, 1959.....	717
Extracellular Potentials from Single Spinal Motoneurons.	
Freygang W. H., Jr. and Frank K. 42(4):749, 1959.....	728
Electrokinetic Membrane Processes in Relation to Properties of Excitable Tissues, 1. "Experiments on Oscillatory Transport Phenomena in Artificial Membranes".	
Teorell T. 42(4):831, 1959.....	740
Electrokinetic Membrane Processes in Relation to Properties of Excitable Tissues, 2. "Some Theoretical Considerations".	
Teorell T. 42(4):847, 1959.....	755
The Responses of Limulus Optic Nerve Fibers to, Patterns of Illumination on the Receptor Mosaic.	
Ratliff F. and Hartline H. K. 42(6):1241, 1959.....	772

RELATION OF FUNCTION TO DIAMETER IN AFFERENT FIBERS OF MUSCLE NERVES*

By CARLTON C. HUNT

(From the Laboratories of The Rockefeller Institute for Medical Research)

(Received for publication, April 26, 1954)

Nerves to muscle contain, in addition to motor fibers, large numbers of myelinated afferent fibers of various diameters (2, 17). Recently the distribution of afferent fibers according to diameter has been studied in de-efferented nerves to a number of hind limb muscles of cat (11, 15). The diameter spectra so obtained show that, as a general pattern, the afferent fiber diameters have three maxima of distribution. Segregation of fibers according to diameter permits classification into the following categories: group I (12 to 20 μ), group II (4 to 12 μ), and group III (1 to 4 μ). The present study attempts to assign receptors of known structure, and function, to fibers of the various diameter groups.

Limb muscles contain two major receptor structures, muscle spindles and tendon organs. The muscle spindle is a complex sense organ, fusiform in shape, which lies parallel to the contractile muscle fibers. Within the encapsulated spindle structure are a number of slender (intrafusal) muscle fibers which receive the terminations of small motor fibers. In the central portion of the spindle the intrafusal muscle elements are entwined by terminal ramifications of afferent fibers (1, 16, 17). Each spindle contains the ending of one large diameter afferent axon, the primary or annulospiral ending. In addition, many spindles receive one or two smaller myelinated afferent fibers which terminate, adjacent to the primary ending, in secondary or flower-spray endings. Stretch deformation of the afferent terminals of the spindles leads to depolarization and initiation of impulses (8). External stretch of muscle evokes discharge in spindle afferent fibers, the frequency of which is a function of rate as well as magnitude of stretch (12). Contraction of the parallel extrafusal muscle fibers decreases the amount of stretch deformation on spindle afferent terminals and causes a cessation or slowing of discharge. Recently another factor has been found to modify discharge in spindle afferent fibers. The group of small diameter (3 to 8 μ) efferent fibers to hind limb muscles of cat has been shown to provide the motor innervation to intrafusal muscle fibers (9, 10). Small motor fibers can be isolated in ventral roots and on stimulation initiate a

* A preliminary report of this work was presented at the XIX International Physiological Congress, Montreal, 1953.

discharge, or increase the frequency of an established discharge, in afferent fibers from spindles; an effect which must result from contraction of the intrafusal muscle fibers. Nerve fibers may be identified as afferent from muscle spindles by discharge pattern during contraction and by their response to excitation of small motor fibers.

The tendon organ is a simpler receptor in which the stretch-sensitive ending of a large diameter afferent nerve fiber lies effectively between the contractile muscle elements and the tendon. Frequency of its discharge provides a fairly direct measure of tension in the tendon whether this be developed by the muscle in contraction or by application of external stretch (12). Increase in discharge rate during contraction is the principal identifying feature of the tendon organ afferent fibers.

Afferent fibers may terminate within the muscle in structures other than spindles and tendon organs. The morphology and discharge characteristics of afferent endings other than those responding to stretch are little known. In addition to myelinated fibers, here under study, there exist unmyelinated afferent fibers from muscle, in smaller proportion than in skin nerves (14). While many afferent fibers from muscle are concerned with stretch reception, others must convey impulses concerned with muscle pain and perhaps other, as yet unknown, types of afferent message.

Afferent fibers from tendon organs and muscle spindles were found to occupy the group I diameter band (12 to 20 μ) although a few smaller spindle afferent fibers were also detected (6). Recently, Merton (13) has also described several spindle afferent fibers from tibialis anterior which were of group II diameter. However, no systematic study has yet been made which would permit assignment of function to the various fiber groups. The present study will show that a reasonably complete accounting of function-diameter relation in groups I and II now is possible. The function of group III fibers remains unknown.

Method

Afferent fibers to soleus and to medial gastrocnemius of the cat have been examined. Individual fibers to these muscles were isolated in filaments of dorsal root, their identity being established by the recording of an impulse following stimulation of the appropriate muscle nerve. Conduction times from muscle nerve to dorsal root were recorded and, at the conclusion of the experiment, the conduction distance was measured. Conduction velocities were calculated from measurements of conduction time and distance. The direct relation between fiber diameter and conduction velocity enabled the latter to be converted to calculated diameter using a factor of 6 (7). Several details of procedure deserve mention.

Laminectomy and limb dissection were performed on adult cats which were either decapitate or anesthetized with dial-urethane. Exposed tissues were covered with

paraffin oil equilibrated with 5 per cent CO_2 and 95 per cent O_2 and kept at $37\text{--}39^\circ\text{C}$. Fibers in spinal roots were isolated by gentle longitudinal teasing of filaments, usually performed on a glass plate. The filaments were raised onto platinum electrodes in oil for recording. Upon completion of the examination of isolated fibers a stimulus was usually applied to the reassembled dorsal root and an antidromic volley recorded from the then cut muscle nerve. The nerve in continuity from dorsal root to muscle nerve was excised and the conduction distance measured with the nerve stretched taut. From stimulus-response intervals 0.1 msec. has been deducted to allow for delay in initiation of impulses at the stimulating cathode.

Damage incurred by longitudinal splitting of dorsal root filaments may cause block or slowing of impulse conduction at some point after, or even before, the filament leaves the rest of the spinal root. To minimize error in latency measurement from this cause the first recording electrode was placed near the point of exit of the filament from volume, latency being measured from shock artefact to onset of spike negativity. Conduction in the final portion of the nerve pathway was often abnormal due to flow of demarcation currents from cut ends as well as from points of injury. Fortunately the region of abnormality represents but a small fraction of the total conduction path and hence was cause for little error in latency measurement.

The aim was to detect fibers of all diameters which were afferent from the muscle under study. Since spike potential amplitude varies with fiber diameter, impulses in larger fibers are recorded most readily. In order to detect the smaller fibers it was necessary to record from thin strands of dorsal root to minimize the external shunt. In each experiment a systematic search was made for afferent fibers in a number of adjacent dorsal root filaments. The rootlets were teased into thin strands and the strands were examined one after another so as to detect, as far as possible, every afferent fiber from the muscle under study. This method of serial examination of root filaments was employed in order to avoid selection of fibers preferentially according to diameter. A different method was used by Hunt and Kuffler (6) to detect muscle afferent fibers. They examined dorsal root filaments for impulses evoked by stretch or contraction of the muscle. The root filaments were subdivided until the response of a single fiber remained. Such a procedure clearly favored isolation of larger fibers since those giving the largest potentials were detected most readily. The method used in the present study was designed to permit isolation of all fibers which yield impulses of detectable size following a stimulus to the muscle nerve. The relation of spike potential amplitude and fiber diameter may be expected to limit the sample to fibers above a certain size but within some diameter range the sample should be representative.

RESULTS

The Sample of Isolated Fibers

By systematic examination of dorsal root filaments, 322 afferent fibers from soleus and 306 from gastrocnemius have been studied. The selection of

isolated fibers is certain to be influenced by fiber size because of the relation of diameter to spike potential amplitude. In order to determine the diameter range over which a representative sample has been obtained, a diameter spectrum has been made relating numbers of isolated fibers to their estimated diameters (calculated according to the velocity/diameter ratio of Hursh). Fig. 1 shows the reconstructed spectrum of afferent fibers from soleus (solid

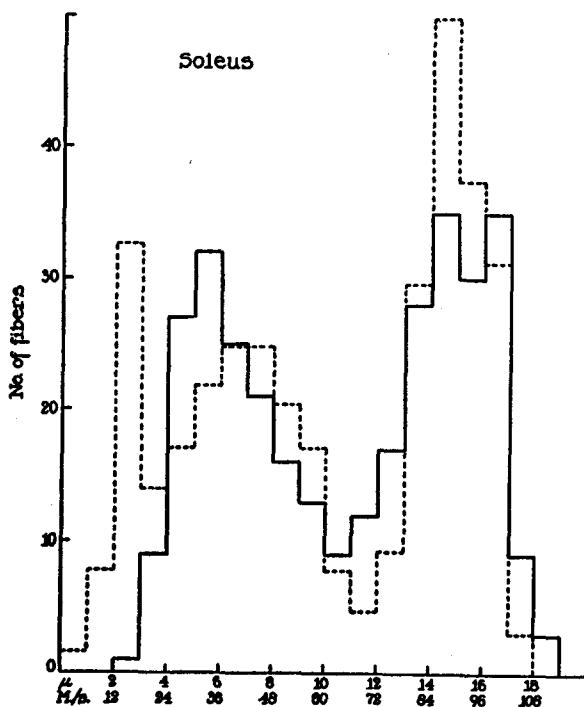


FIG. 1. Comparison of the diameter distribution of isolated fibers and the histological afferent fiber distribution. Soleus nerve. Solid line, numbers of isolated fibers according to conduction velocity and calculated diameter. Dashed line, diameter spectrum by histological measurement (from cat II, Lloyd and Chang (11)). Both distributions are scaled so that the numbers in the range 4 to 20 μ are the same.

line) together with the histological diameter distribution of afferent fibers in a nerve to soleus (dotted line, taken from Lloyd and Chang (11)). The distributions of the fiber diameters in the two spectra are similar for the range above 4 μ , a fact that suggests that fibers in this diameter range can be detected by the isolated fiber technique in proportion to their incidence in the muscle nerve. Further, the similarity of the isolated fiber and histological spectra indicates that the measurement of conduction velocities and the factor used for conversion of velocity to diameter are valid.

Fig. 2 compares, in the manner described above, the reconstructed isolated fiber diameter distribution and the histological diameter spectrum (from cat II, Lloyd and Chang (11)) of afferent fibers to medial gastrocnemius. As for soleus, comparison of the two spectra indicates a representative sampling of group I and II fibers. Fibers in group III have seldom been detected, the principal difficulty being size of recordable potential.

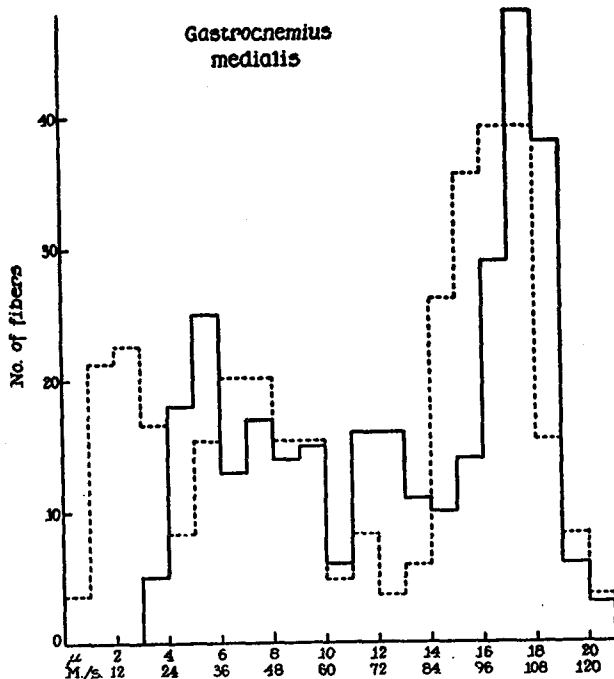


FIG. 2. Comparison of the diameter distribution of isolated fibers and the histological afferent fiber distribution. Nerve to medial gastrocnemius. Solid line, numbers of isolated fibers according to conduction velocity and calculated diameter. Dashed line, diameter spectrum by histological measurement (from cat II, Lloyd and Chang (11)). Both spectra are scaled as in Fig. 1

One factor should be considered in comparing the histological and isolated fiber diameter distributions. The former is derived from one animal, the latter from many. Thus, the isolated fibers from soleus were studied in thirteen animals, those from medial gastrocnemius in ten. The extent of variation from animal to animal in the diameter spectra of muscle nerves has not been studied systematically. However, skin nerves from several animals have been compared. Thus the saphenous nerve is known to vary considerably from cat to cat, not in the relative position of the various fiber groups but in absolute position on the diameter scale (3). The extent to which similar variation occurs in muscle nerves is not known but comparison of maximal conduction

velocities of afferent volleys in the nerves studied (Table I) indicates that variation in maximal diameter is approximately 10 per cent. If the absolute position of the spectrum on the diameter scale differed considerably among animals, a representative sample of fibers would produce a reconstructed spectrum with contours less distinct than a spectrum derived from one nerve.

Fiber Diameter and Receptor Function

It has been shown that the method of isolation of single afferent fibers used in the present study permits a representative sampling of group I and II

TABLE I
Maximum Conduction Velocities of Afferent Volleys in Nerves Employed for Isolated Fiber Analysis

	Conduction distance	Maximal velocity
	cm.	m./sec.
Soleus	16.0	94
Average maximal velocity 101.8 m./sec.	18.0	97
	16.3	102
	16.5	97
	16.0	114
	17.0	97.5
	14.5	111
Gastrocnemius medialis	14.2	112
Average maximal velocity 118.4 m./sec.	14.0	127
	14.8	124
	14.6	111
	14.2	123
	16.7	128
	16.0	113
	16.5	119
	15.3	122

fibers and that the calculation of fiber diameter is without serious error. In addition to measurement of conduction velocity, each fiber isolated was characterized as to its discharge pattern with the purpose of identifying the type of receptor in which it terminated.

Of the fibers isolated with calculated diameter greater than $4\ \mu$, only two from soleus and one from medial gastrocnemius could not be identified as to receptor function. Since these three fibers form a negligible proportion of the group I and II fibers examined and since damage to an occasional fiber distal to the stimulating electrodes on the intact muscle nerve could not be excluded, they have not been entered in the data presented below. All the remaining fibers of group I and II diameter were found to terminate in stretch receptors.

Group I was found to be composed of fibers of two types: (a) afferent fibers displaying discharge patterns characteristic of muscle spindle receptors and (b) afferent fibers showing the discharge pattern of tendon organ receptors. Group II as arbitrarily defined in the histological spectrum was composed almost entirely of muscle spindle afferent fibers. The distribution of spindle (A) and tendon organ (B) fibers, plotted according to velocities and calculated diameters, is shown for soleus in Fig. 3 and for medial gastrocnemius in Fig. 4.

If the division between groups I and II is placed arbitrarily at $12\ \mu$ all tendon organ (B) fibers fall into group I except four fibers to soleus and two

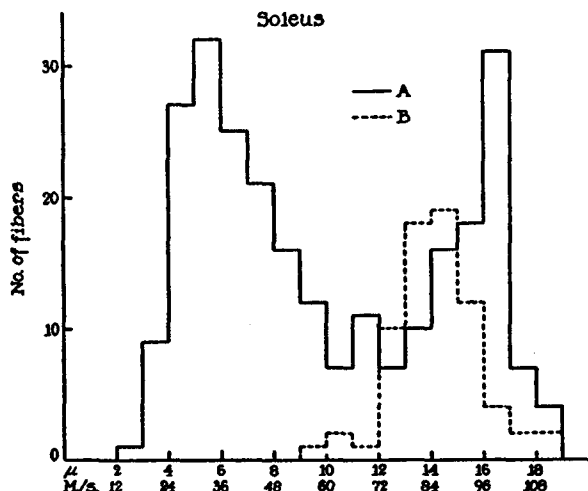


FIG. 3. Diameter distribution of afferent fibers from muscle spindles (A) and tendon organs (B). Soleus nerve. Note the unimodal distribution of tendon organ fibers and the bimodal distribution of afferent fibers from muscle spindles.

to medial gastrocnemius. The distribution of tendon organ fibers is certainly unimodal and all fibers of this type have been included in the category group IB with the reservation that the trailing edge of their distribution may extend slightly into the upper diameter range of group II.

Muscle spindle afferent fibers have a bimodal diameter distribution. The large spindle afferent fibers (12 to $20\ \mu$), hereafter called group IA, together with tendon organ fibers (IB), account for the total of group I fibers. Group II appears as a virtually homogeneous population of spindle afferent fibers. The two distinct groups of spindle afferent fibers can be seen in the diameter distribution of fibers from soleus (Fig. 3) and from medial gastrocnemius (Fig. 4). Certain differences exist between the afferent supply to the two muscles; these may be of general interest since soleus and medial gastrocnemius are

red and pale muscles respectively. For instance, isolated fiber analysis shows that fibers from medial gastrocnemius have larger maximal diameter than those from soleus, a fact in accord with histological measurement (11). A comparison of the reconstructed spectra of the two muscle nerves also shows that for soleus the distribution maximum of group IA fibers falls at a slightly higher diameter than the peak of group IB fibers, while for medial gastrocnemius the maxima of groups IA and IB coincide.

It was shown above that afferent fibers of groups I and II were isolated in proportion to their numbers in the muscle nerve. Since each fiber was detected by recording the impulse therein which followed a stimulus to the

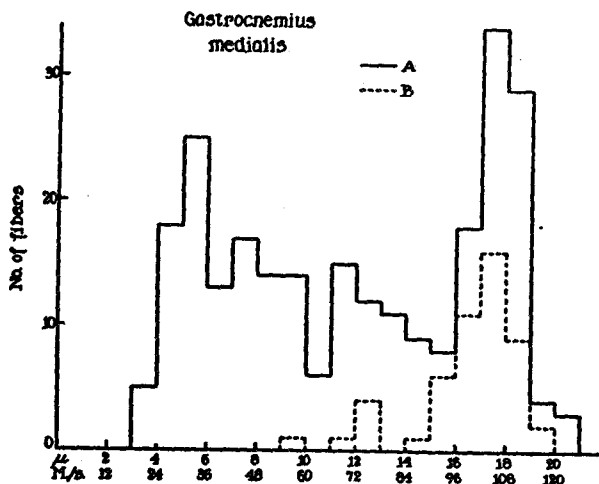


FIG. 4. Diameter distribution of afferent fibers from muscle spindles (A) and tendon organs (B). Nerve to medial gastrocnemius. Note the unimodal distribution of B fibers and the bimodal distribution of A fibers.

muscle nerve, no factor other than diameter should enter into the selection of fibers. In other words, fibers of the same diameter but with different discharge patterns should be detected with equal ease. Therefore, the incidence of the various fiber types (A and B) in groups I and II may be considered representative. From the total number of group I and II fibers in a muscle nerve and from the proportion of different fiber types determined by isolated fiber analysis, one may estimate the number of fibers in the various categories (IA, IB, and II) to a muscle. This has been done for soleus and medial gastrocnemius (Table II). Several comparisons may be made: The numbers of fibers in group I and in group II appear in very nearly the same ratio in both isolated fiber and histological counts. The nerve to medial gastrocnemius differs from that to soleus in having a larger number of fibers, the majority

of which appear in group IA. Thus both nerves have a similar number of fibers in group IB and in group II.

Characteristics of the Receptor Discharge in Fibers of the Various Groups

Two types of afferent fiber from muscle spindles may now be recognized, those contained in group IA and those in group II of the diameter distribution. Fibers of group IA probably terminate in primary endings while group II fibers probably terminate in secondary endings within the spindles (see Discussion). A comparison of the receptor discharge in fibers of these two types was undertaken to determine any differences which might exist in receptor characteristics. The discharge patterns have been compared in the following circumstances: externally applied stretch, active contraction, and small motor fiber stimulation.

TABLE II

Incidence of Various Afferent Fiber Groups from Gastrocnemius medialis and Soleus

	I	IA	IB	II
Gastrocnemius medialis				
Histological (Lloyd and Chang).....	156	—	—	98
Calculated fiber types in representative nerve*.....	150	108	42	104
Soleus				
Histological (Lloyd and Chang).....	106	—	—	95
Calculated fiber types in representative nerve*.....	103	58	45	98

* Calculated so that the total of group I and II fibers equals that determined in the histological study of the representative muscle nerve.

Responses to External Stretch.—Threshold to steady stretch was determined in a number of group IA and II spindle afferent fibers by measurement of the minimal amount of stretch required to evoke maintained discharge. Individual threshold values have been entered on a plot relating stretch threshold to conduction velocity (Fig. 5). The average threshold of twenty group IA fibers (conduction velocity above 72 m./sec.) was 3.3 gm., while the average threshold for thirty-six group II fibers (conduction velocity 24 to 72 m./sec.) was 19.0 gm. However, it is clear that the two fiber groups cannot be separated completely on this basis for some of the group II receptors may have lower thresholds than certain of the IA fiber receptors. In comparison with tendon organs the receptors of muscle spindles, both of the group IA and group II afferent fiber type, exhibit a low stretch threshold. The receptors of B fibers usually require tensions of 100 to 200 gm., or more, for sustained firing.

Responses during Contraction.—During the period of contractile shortening of extrafusal muscle fibers, the stretch-evoked discharge in spindle afferent

fibers is slowed in frequency or completely silenced. Considerable variation exists in the modification of discharge during contraction among different fibers of group IA. To detect such small differences as may exist in this respect between fibers of groups IA and II would require more extensive an analysis than has been considered feasible. In short, it seems doubtful whether there is any difference of functional significance in the behavior during contraction between receptors of the two groups of spindle afferent fibers.

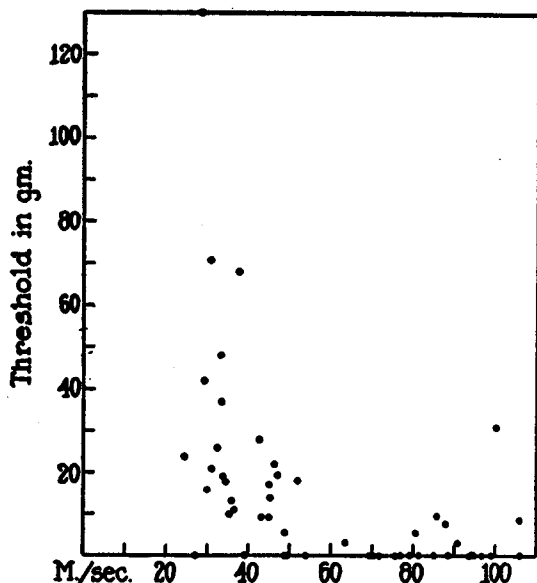


FIG. 5. Threshold to steady stretch of spindle afferent fibers from soleus. Individual fibers are entered on the plot relating the minimal amount of tension in grams which causes the receptor of the fiber to discharge continuously and the conduction velocity of the fiber. Note that the fibers conducting at group I velocities (above 72 m./sec.) have the lower average threshold to steady stretch.

When a muscle is caused to contract under rigidly isometric conditions certain afferent fibers from spindles may exhibit an increase in discharge rate during the period of tension development and yet if some shortening is permitted they show a typical silent period. Matthews (12) considered fibers which showed this discharge characteristic (his A2 units) to come from primary endings within the spindles, the increased discharge resulting from contraction of intrafusal muscle elements. Hunt and Kuffler (6) suggested that the A2 type of discharge was probably not the result of intrafusal fiber contraction, but was caused by some unusual distribution of tension within the muscle which increased the amount of stretch deformation on some spindle endings during

rigidly isometric contraction. A number of spindle afferent fibers of groups IA and II have been examined for the above pattern. Certain fibers of both groups have shown the A2 type of discharge. If group IA and II fibers terminate in primary and secondary endings respectively (see below), this increased

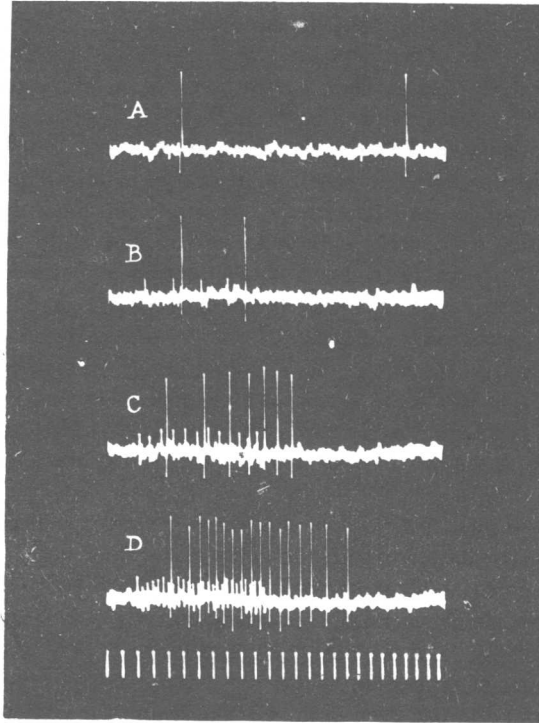


FIG. 6. Response of a group II afferent fiber to stimulation of an isolated small nerve fiber to its spindle. This afferent fiber from soleus had measured conduction velocity of 51 m./sec. (calculated diameter 8.5μ) and showed a discharge pattern during contraction typical of a spindle receptor. A, base line discharge at low initial tension. B, C, and D; stimulation of small motor fiber. B, 4 stimuli at 55/sec. C, 12 stimuli at 130/sec. D, 27 stimuli at 300/sec. Time 100 cycles/sec.

discharge during isometric contraction cannot be attributed only to the primary endings.

Following a stimulus to an intact muscle nerve, certain afferent fibers from muscle exhibit a brief, high frequency burst of impulses which occurs very early in contraction (6). This "early discharge" has been seen less frequently in group II fibers than in fibers of group IA and has also tended to consist of fewer impulses at lower frequencies when it has occurred in group II fibers.