Principles And Techniques Of Electron Microscopy

BIOLOGICAL APPLICATIONS

Edited by M.A.Hayat

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Principles and Techniques of Electron Microscopy

BIOLOGICAL APPLICATIONS

Volume 9

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PREFACE

This is the ninth volume of a multi-volume series on the principles and techniques employed for studying biological specimens with the aid of an electron microscope. Since its inception in 1970, the series has successfully reflected the growth of electron microscopy in instrumentation as well as in methodology. There was a pressing need to keep readers abreast of the remarkable expansion of the field in recent years and the ever growing importance of its contributions to the understanding of many problems in biological and medical sciences. This treatise serves as an international authoritative source in the field, and is designed to cover important new developments systematically. The treatise departs from the tradition that books on methodology present only the contemporary consensus of knowledge. It is written by scholars, and when they have anticipated the potential usefulness of a new method, they have so stated. The authors have not hesitated to include ideas in progress. The treatise should serve as a guide and survey, which can save a newcomer the tedious search for information scattered in biological journals.

This volume has developed over the years through the joint effort of ten distinguished author-scientists. As a result, a most comprehensive compilation of methods developed and used by a large number of competent scientists has been achieved. The book contains new viewpoints with particular regard to current problems. Areas of disagreement and potential research problems have been pointed out. It is hoped that the readers will become aware that correct interpretation of the information retrieved from electron micrographs is dependent

vi PREFACE

upon an understanding of the principles underlying the methodology and instrumentation.

The basic approach in this volume is similar to that in the previous eight volumes, in that the methods presented have been tested for their reliability, and are the best of those currently available. The instructions for the preparation and use of various solutions, media, stains, and apparatus are straightforward and complete, and should enable the worker to prepare his or her specimens without outside help. Before undertaking the processing, one should read the entire procedure and prepare necessary solutions and other media. Each chapter is provided with an exhaustive list of references with complete titles. Full author and subject indexes are included at the end of the book.

It is encouraging to know that the previous volumes have been received favorably. It is my impression that this volume will also fulfill its purpose: to provide an understanding of the usefulness, limitations, and potential applications of special methods employed for studying the structure, composition, size, number, and location of cellular components, and to provide details of current improvements in the instrumentation.

M. A. HAYAT Berkeley Heights, N.J.

Contents to

Volume 1, M. A. Hayat

FIXATION
EMBEDDING
SECTIONING
STAINING
SUPPORT FILMS

Volume 2

FREEZE-SUBSTITUTION AND FREEZE-DRYING, Lionel I. Rebhun
THE FREEZE-ETCHING TECHNIQUE, James K. Koehler
NEGATIVE STAINING, Rudy H. Haschemeyer and Robert J. Meyers
SHADOW CASTING AND REPLICATION, W. J. Henderson and K. Griffiths
HIGH RESOLUTION AND SHADOWING, R. Abermann, M. M. Salpeter and
L. Bachmann
AUTORADIOGRAPHY, M. M. Salpeter and L. Bachmann

Volume 3

THE ELECTRON MICROSCOPE, Saul Wischnitzer
ELECTRON MICROSCOPY OF SELECTIVELY STAINED MOLECULES,
T. Koller, M. Beer, M. Müller and K. Mühlethaler
HIGH RESOLUTION DARK-FIELD ELECTRON MICROSCOPY, Jacques
Dubochet
IN-FOCUS PHASE CONTRAST ELECTRON MICROSCOPY, H. M. Johnson
ELECTRON MICROSCOPIC EVALUATION OF SUBCELLULAR FRACTIONS OBTAINED BY ULTRACENTRIFUGATION, Russell L. Deter
STEREOLOGICAL TECHNIQUES FOR ELECTRON MICROSCOPIC MORPHOMETRY, Ewald R. Weibel and Robert P. Bolender
CRITICAL POINT-DRYING METHOD, M. A. Hayat and B. R. Zirkin

Volume 4

OPTICAL SHADOWING, Glen B. Haydon

RELATIVE MASS DETERMINATION IN DARK FIELD ELECTRON

MICROSCOPY, G. J. Brakenhoff

CORRELATIVE LIGHT AND ELECTRON MICROSCOPY OF SINGLE

CULTURED CELLS, Zane H. Price

DENATURATION MAPPING OF DNA, Ross B. Inman and Maria Schnöss

EXAMINATION OF POLYSOME PROFILES FROM CARDIAC MUSCLES.

Kenneth C. Hearn

PARTICLE COUNTING OF VIRUSES, Mahlon F. Miller II

ULTRAMICROINCINERATION OF THIN-SECTIONED TISSUE, Wavne R.

Hohman

PREPARATORY METHODS FOR ELECTRON PROBE ANALYSIS, James R.

Coleman and A. Raymond Terepka

Volume 5

QUANTITATIVE MAPPING WITH THE ELECTRON MICROSCOPE, Peter Sterling

PHOTOGRAPHIC ASPECTS OF ELECTRON MICROSCOPY, G. C. Farnell and R. B. Flint

ENVIRONMENTAL DEVICES IN ELECTRON MICROSCOPY, David L.

OPTICAL DIFFRACTOMETRY, Bigrn V. Johansen

THE ANALYTICAL ELECTRON MICROSCOPY, EMMA-4, Barry A. Weavers

Volume 6

HIGH VOLTAGE ELECTRON MICROSCOPY, Colin Humphreys

THE PRINCIPLES OF HIGH RESOLUTION ELECTRON MICROSCOPY.

J. M. Cowley

CONTRAST AND IMAGE FORMATION OF BIOLOGICAL SPECIMENS.

R. E. Burge

THE ANALYSIS OF BIOLOGICAL STRUCTURE WITH X-RAY DIFFRAC-

TION TECHNIQUES, Alexander McPherson Jr.

TILTING EXPERIMENTS IN THE ELECTRON MICROSCOPE, Rainer H.

ELECTRON AUTORADIOGRAPHY OF FREE SPECIMENS, Nadir M. Maraldi CRYOULTRAMICROTOMY, René Simard

ELECTRON INTERFERENCE MICROSCOPE, T. Hibi and K. Yada

Volume 7

SPECIMEN DAMAGE IN THE ELECTRON MICROSCOPE, M. S. Isaacson FREEZE-DRYING FOR ELECTRON MICROSCOPY, M. V. Nermut IMAGE RECONSTRUCTION OF ELECTRON MICROGRAPHS BY USING EQUIDENSITE INTEGRATION ANALYSIS, Klaus-Rüdiger Peters G-BANDING OF CHROMOSOMES, F. Ruzicka AUTORADIOGRAPHIC LOCALIZATION OF DNA IN NONMETABOLIC CONDITIONS, M. Gueskens OPTICAL ANALYSIS AND RECONSTRUCTION OF IMAGES, A. J. Gibbs and A. J. Rowe

MIRROR ELECTRON MICROSCOPY, R. S. Gvosdover and B. Ya. Zel'dovich ELECTRON MICROSCOPY OF BANDED MAMMALIAN CHROMOSOMES, Gary D. Burkholder

EQUIDENSITOMETRY: SOME NEUROBIOLOGICAL APPLICATIONS L. T. Ellison and D. G. Jones

Volume 8

SPECIMEN SUPPORTS, W. Baumeister and M. Hahn
PREPARATION AND ANALYSIS OF SERIAL SECTIONS IN ELECTRON
MICROSCOPY, Robert L. Knobler, Jerome G. Stempak and Mary Laurencin
CALIBRATION OF MAGNIFICATION IN TRANSMISSION ELECTRON
MICROSCOPE, Robert F. Dunn

CONTRAST ENHANCEMENT BY USING TWO ELECTRON MICROGRAPHS, D. L. Misell

INTERFERENCE PHENOMENON ON OSMIUM TETROXIDE-FIXED SPECIMENS FOR SYSTEMATIC ELECTRON MICROSCOPY, Karl Hermann Andres and Monika von Düring

COMPUTER PROCESSING OF ELECTRON MICROGRAPHS, P. W. Hawkes

CONTENTS

1 ELECTRON MICROSCOPY OF ATOMS John P. Langmore	
Introduction	1
History	. 1
Single-Atom Microscopes	. 2
Theory of Single-Atom Imaging	5
Image of Single Atom	5
Contrast and Resolution	9
Limitations of the Theory	12
Atom Visibility	13
Image Artifacts	13
Noise and Minimum Exposure Needed to "See" Single Atoms	15
Other Practical Considerations	23
Experimental Determination of Resolution and Contrast	25
Observation of Model Compounds	29

Preface

viii CONTENTS

Practical Techniques of Single-Atom Microscopy	34
Low-Noise Support Films	<i>35</i>
Clean Support Films and Specimens	<i>38</i>
Image Processing	42
Specimen Damage	47
Chemical Damage	47
Radiation Damage	48
Experimental Evidence of Radiation Effects	50
Biological Applications	,52
Future Developments	54
References	57
2 ELECTRON MICROSCOPY OF DNA John O. Thomas	64
Introduction	64
Methods for Mounting DNA	65
Cytochrome c	. 65
Benzalkonium Chloride	69
Intercalating Agents	72
Anthrabis	73
Activated Carbon Grids	73
Applications	77
Length Measurements	77
Heteroduplex Mapping	77
Denaturation Mapping	<i>78</i>
DNA-Protein Complexes	80
References	81
3 TECHNIQUES FOR VISUALIZATION OF GENETIC MATERIAL	
Aimée H. Bakken and Barbara A. Hamkalo	84
Introduction	84
The Technique	85
Preparation of Support Films	85

	CONTENTS	ix
Preparation of Hydrophilic Carbon Surfaces		87 [°]
Preparation of Specimens	-	87
Examples of Methods of Cell Lysis and Dispersal		90
Dispersal of Folded Genomes		92
Observations		94
Prokaryotic Preparations		94
Eukaryotic Preparations		96
Other Applications and Future Prospects		104
References		104
4 LOCALIZATION OF ACETYLCHOLINE RI Mathew P. Daniels and Zvi Vogel	ECEPTORS	
Introduction		107
General Procedures and Controls for the α BT Immuno Method	-	110
Specific Procedures and Results		112
Preparation of Reagents		112
Mouse Diaphragm Endplates		113
Muscle Monolayer Cultures		118
Human Muscle Biopsies		120
Concluding Remarks		122
References	j	123
5 ELECTRON MICROSCOPY OF ACT	PIBI	
Arthur Forer		
Introduction		126
Muscle Proteins	1	127
Electron Microscopical Appearances of Muscle Actin	. 1	129
Negatively Stained Muscle Actin		129
Muscle Actin in Thin Sections	1	121

x CONTENTS

Muscle Actin plus Myosin Subfragments	132
Techniques for Studying Actin-Containing Filaments	139
Electron Microscopical Appearances of Non-Muscle Actin	145
HMM (or S1) plus Non-muscle Actin	145
Cells That Contain Actin	152
Biochemical Isolation of Actin from Non-muscle Cells	<i>153</i>
Technical Aspects of Electron Microscopy of Actin in Non-muscle Cells	155
Appendix I	159
Solutions	<i>159</i> .
Procedure	159
Appendix II	160
Appendix III	161
A. Protocol for Preparing S1 from Myosin	161
B. Protocol for Preparing S1 from Glycerinated Muscles (based on that	
given by Cooke, 1972)	162
Appendix IV	164
A. Preparing the Acetone Powder	164
B. Extracting Actin from the Acetone Powder	165
References	166
6 ELECTRON MICROSCOPY OF GLYCOPROTEINS BY HIGH RESOLUTION METAL REPLICATION Henry S. Slayter	Y
Introduction	175
Macromolecular Replication Methods	176
Rationale	176
Development of Present Methodology	180
The Effect of Variable Parameters on Nucleation and Growth of	
Metal Crystallites	183
Control of Replica Thickness	186
Present Methodology	187
Interpretation of Results	193
Application of Metal Replica Method to Glycoprotein Structure	206
Electron Microscopy of CEA	206
Electron Microscopy of TA3-Ha Tumor Cell Surface Glycoproteins	219

CONTENTS	S x
Electron Microscopy of Blood Group Substances Electron Microscopy of Fibrinogen and Fibrin	224 230
Concluding Remarks	238
References	. 241
7 EXAMINATION OF THYROGLOBULIN MOLECULES THE ELECTRON MICROSCOPE Gertrud Berg	IN
Introduction The Formation and Function of Thyroglobulin	246 246
Preparatory Techniques Isolation of Thyroglobulin Preparation of Specimens for Electron Microscopy Staining	249 249 251 252
Interpretation Studies of Hormone-Containing Thyroglobulin Studies of Non-Hormone Containing Thyroglobulin Molecules	256 256 257
Concluding Remarks	260
References	260
8 ELECTRON OPTICAL MEASUREMENT OF SURFACE CHARGES Imre Veres	
Introduction	262
Surface Charges	263
Appearance of Surface Charges on Biological Membranes	264
Electron Optical Measuring Methods	265
Semiquantitative Topological Determination of Coulomb Forces	271

xii CONTENTS

Biological Examples and Significance of Surface Charge	272
SEM Visualization of Surface Charges of Biomembranes	278
Future Developments of Labeling Technique and Its Correlation With Selective Staining of Glycocalyx	284
References	285
AUTHOR INDEX	289
SUBJECT INDEX	299

1. ELECTRON MICROSCOPY OF ATOMS

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INTRODUCTION

The electron microscope has the inherent capability of atomic resolution, by virtue of the very short wavelength of medium energy electrons, and the implied capability of somehow "seeing" single atoms. However, since a micrograph is a record of the projection of an object, even a thin specimen such as a single protein molecule would be imaged as a confusing array of overlapping atoms. Fortunately, electrons interact much more strongly with heavy atoms than with light ones, so that single heavy atoms could be localized from a projection by virtue of their high contrast. It is this possibility that has led some biologists to dream of visualizing heavy atoms placed at strategic locations within biological molecules, as bright dots in a sea of faint overlapping images of light atoms. With the recent achievements of electron microscopy in imaging single heavy atoms, the prospects for realizing this dream are becoming increasingly bright.

History

The earliest calculations of single-atom contrast were made by Hillier (1941) and Schiff (1942), followed by much more realistic estimates by Boersch (1947), who showed that a microscope of 3 Å resolution should be capable of visualizing atoms of low atomic number, $Z \gtrsim 3$ by phase contrast bright field and $Z \gtrsim 13$ by dark field microscopy. Soon after, Scherzer (1949) described a practical and elegant method to achieve almost perfect phase contrast by properly balancing

2 PRINCIPLES AND TECHNIQUES OF ELECTRON MICROSCOPY

defocus against the inherent spherical aberration of electron lenses. These early theoretical predictions were not immediately followed by experimental verification, simply because electron microscopes were not yet able to achieve atomic resolution ($\lesssim 5$ Å). Even when atomic resolution became possible, atoms were not immediately visualized. M. Beer, one of the earliest and most dedicated proponents of the use of specific heavy atom staining of biological molecules, concluded that even two gold atoms were barely detectable by phase-contrast microscopy (Highton and Beer, 1968).

Meanwhile, Müller (1957) had shown conclusively that his field ion microscope (FIM) could clearly image single atoms placed on the end of a very sharp tungsten tip. Although this technique has been of tremendous value to the study of atomic adsorption, desorption, and diffusion, there are few indications that the FIM will ever be useful to biology. The present difficulties in FIM imaging of biological molecules are field denaturation, field desorption, and the fact that imaging seems to involve an uncertain combination of many physical and chemical, bulk and surface properties of the specimen (Machlin et al., 1974).

The promise of atomic electron microscopy was finally realized when Crewe et al. (1970) reported the first clear images of uranium and thorium atoms, obtained using a scanning transmission electron microscope (STEM) developed by Crewe and Wall (Crewe et al., 1968; Crewe, 1970; Crewe and Wall, 1970). Operated in the dark field mode proposed by its inventor, von Ardenne (1938), the STEM was able to produce high contrast, low noise images by use of very thin, clean carbon support films and by use of the inelastically scattered electrons to reduce the carbon film noise. Soon after, single atom imaging in the conventional transmission electron microscope (CTEM) was also reported (Formanek et al., 1971; Henkelman and Ottensmeyer, 1971; Hashimoto et al., 1971).

This chapter reviews the present state-of-the-art of single atom microscopy and outlines those aspects the author believes will be of importance in the future. It is hoped that it will lead to a better understanding of the theoretical and experimental basis of atomic imaging and will direct interest toward the most important problems in the application of atomic microscopy to biological research.

Single-Atom Microscopes

A brief review of the geometries of several types of microscopes useful for imaging single atoms will make the calculations and experiments presented later more understandable. Schematic illustrations of these microscopes (Figs. 1.1 and 1.2) show the essential similarities and differences among the instruments.

The mainstay of biological microscopy is the bright field conventional transmission electron microscope (CTEM) (Fig. 1.1a). The specimen is illuminated by a nearly parallel beam of coherent electrons, with a small convergence semiangle β_c .

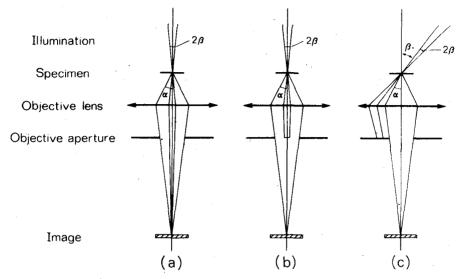


Fig. 1.1 Schematic diagrams of the conventional transmission electron microscopes: (a) bright field, (b) beam stop dark field, and (c) tilted illumination dark field. Angle α is the objective aperture angle and β_C is the convergence angle of illumination.

A magnetic lens is used to image the transmitted electrons that pass through the objective aperture of semiangle α , consisting of the unscattered electrons, most of the inelastically scattered electrons, and ~20% of the elastically scattered electrons. Phase contrast is produced by interference of the unscattered with the elastically scattered electrons. Scattering contrast is formed simultaneously, by stopping most of the elastically scattered electrons at the objective aperture; but is generally much weaker than phase contrast and is therefore ignored. The inelastically scattered electrons are not useful for forming single-atom images because (a) few electrons are inelastically scattered by heavy atoms, (b) inelastic events are not highly localized (Isaacson et al., 1974b), and (c) chromatic aberration prevents the inelastically scattered electrons from being correctly focused (e.g. Crick and Misell, 1971).

To reduce the background caused by the noninformative, unscattered and inelastically scattered electrons, a physical beam stop can be placed at the objective aperture plane, as shown in Fig. 1.1b. This configuration is called a beam stop dark field CTEM. The contrast in this geometry is very high, since the background due to the unscattered electrons is not present.

Dark field can also be achieved by tilting the axis of illumination, by the angle β_T , so that the unscattered beam is stopped by the objective aperture (Fig. 1.1c). This seems to be the most practical dark field geometry for the CTEM, since the unscattered beam can be placed far from the edge of the aper-