DEVELOPMENTS IN FOOD SCIENCE 8

MYCOTOXINS

Production, Isolation, Separation and Purification

Edited by

Vladimír Betina

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ELSEVIER
Amsterdam — Oxford — New York — Tokyo 1984

ELSEVIER SCIENCE PUBLISHERS B.V.
Molenwerf 1
P.O. Box 211, 1000 AE Amsterdam, The Netherlands

Distributors for the United States and Canada:

ELSEVIER SCIENCE PUBLISHING COMPANY INC. 52, Vanderbilt Avenue New York, N Y. 10017

ISBN 0-444-42289-7 (Vol. 8) ISBN 0-444-41688-9 (Series)

© Elsevier Science Publishers B.V., 1984

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Printed in The Netherlands

PREFACE

Mycotoxins are important fungal metabolites that have been recognized within the last two decades as a potential threat to human and animal health, and the effects of mycotoxins result in economic losses to various extents (e.g. aflatoxins, trichothecenes). Mycotoxins present a challenge for scientists working in a wide range of disciplines such as microbiology, biochemistry, structural chemistry, toxicology, pharmacology and genetics.

The evaluation of the role and extent of mycotoxin contamination in feed-stuffs and foods requires the isolation of the toxin(s) from a suspected commodity or from fungal cultures isolated from the suspected material. This book has been written with the aim of helping investigators in the field by bringing together the most important data, scattered in the literature, on the production, isolation, purification and characterization of the best known mycotoxins. Expert scientists in the field of mycotoxin isolation and characterization have provided contributions based on extensive personal experience.

The book consists of two parts. In the General Part, data on the occurrence, production, biological activities and biochemical modes of action of mycotoxins are discussed, followed by chapters describing screening for toxigenic fungi and applications of chromatographic techniques in the isolation, purification and characterization of mycotoxins.

In the Special Part, key chemical families and the most important individual mycotoxins have been chosen, the largest families being the trichothecenes, the epipolythiodioxopiperazines and the cytochalasans. As would be expected, ergot alkaloids and toxins from Basidiomycetes have not been included. Data on producing organisms and the physical, chemical and biological properties of each mycotoxin are given, followed by methods of production, isolation, purification, chromatographic characterization, detection and assays.

I express my thanks to all of the contributors for their excellent cooperation in preparing their manuscripts.

Bratislava December 1983

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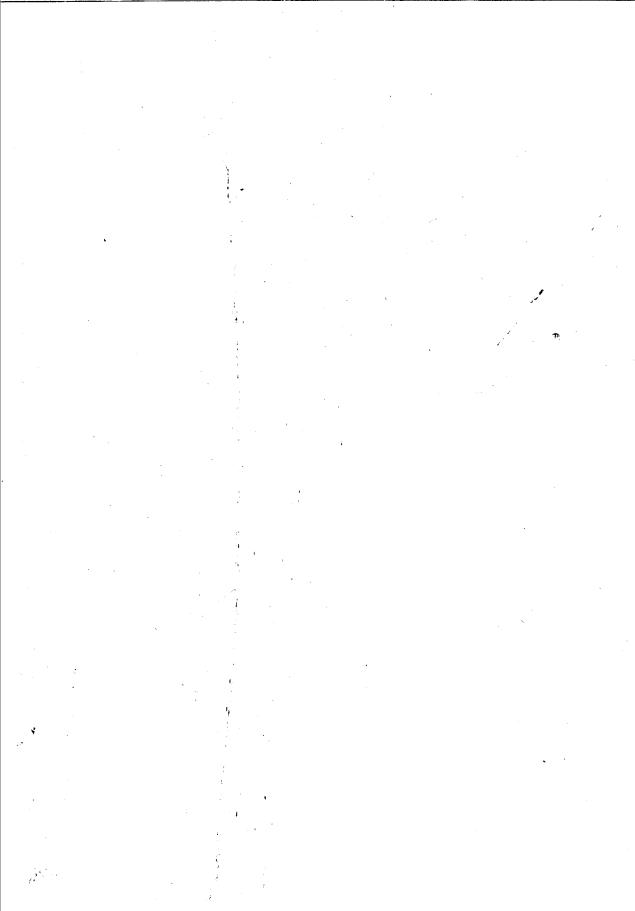
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General Part



CHAPTER 1

TOXIGENIC FUNGI AND THEIR OCCURRENCE

V. BETINA

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1.1. INTRODUCTION

The development of man has been associated with various uses of fungi. The age-old role of these microorganisms in the production of bread, alcoholic beverages or Oriental food fermentations is well known. Today, fungi are applied in the production of cheese, livestock feeds, enzymes, organic acids, vitamins, antibiotics and steroids, and also have a substantial role in plant, animal and human diseases and in crop and food spoilage. However, mycotoxins and mycotoxicoses received little attention until the early 1960s, when the aflatoxins were discovered. Mycotoxicoses are defined as "poisonings of the host which follow the entry into the body of toxic substances of fungal origin".

The oldest known mycotoxicoses are associated with various species of the parasitic genus *Claviceps* and their metabolic products. The toxic effects of these compounds have been the cause of mass poisonings in both man and animals over several centuries. The animal poisonings usually resulted from feeding on grass infected with the ergot fungus, whereas the toxic effects in man stemmed from eating bread of other products made from rye flour which was contaminated with sclerotia of *Claviceps purpurea*².

Outbreaks of other mycotoxicoses have been reported in many countries, such as diseases of pigs and horses associated with the ingestion of barley infected with Fusarium graminearum (barley scab)³ in the U.S.A.; stachybotryotixicosis of horses in the U.S.S.R., first reported in the 1930s, which occurred as a result of the fungus Stachybotrys atra growing on straw¹; facial eczema in sheep in New Zealand that had grazed on pasture infected with the fungus Pithomyces chartarum⁴, from which sporidesmin was isolated⁵; liver tumours induced by yel-

lowed rice toxins in $Japan^6$; and alimentary toxic aleukia (ATA) reported in eastern Siberia in 1913 and caused by fusaria growing on grain that had overwintered on the ground⁷.

1.2. TOXIGENIC FUNGI

Toxigenic species can be found in all major taxonomic groups of fungi. However, most of the mycotoxins hitherto known have been recognized as metabolic products of genera such as Aspergillus, Penicillium and Fusarium. In addition to the already mentioned Claviceps, other Ascomycetes are known to produce various mycotoxins. Examples of toxigenic fungi belonging to several taxonomic groups and their mycotoxins are given in Tables 1.1, 1.2 and 1.3 The mycotoxins listed are far to be complete. Toxins from Basidiomycetes are not included in this book.

As mycotoxin-producing fungi are abundant in nature and favourable conditions for their production are easily achieved, it was concluded that few edible substances are considered safe from mycotoxin contamination. Fungal invasion of foods can occur at any stage of the food production chain, i.e., from the time of planting through harvesting, transportation and storage, during the prepara-

TABLE 1.1

MYCOTOXINS PRODUCED BY SPECIES OF THE GENUS ASPERGILLUS

Mycotoxin	Produc ng organism	Mycotoxin	Producing organism					
Aflatorins Aflatrem Asclad ol Aspergillic acid Citreov ridins Cyclopi zonic acid 5 6-D methoxy- st r gmatocystin Emodin Eryth oglaucin Flavu oxin Fumagillin Fumitoxins	A. flavus, etc. A. flavus A. clavatus A. flavus A. terreus A. flavus A. multicolor A. wenti A. glaucus A. flavus A. flavus A. fumigatus A. fumigatus	Fumitremorgins Helvolic acid Kojic acid Malformins Ochratoxins Patulin Physcion Sterigmatocystin Terretonin Viomellein Xanthocillin X	A. fumigatus A. fumigatus A. fumgatus, etc. A. niger A. ochraceus A. clavatus, etc. A. glaucus A. versicolor A. terreus A. ochraceus A. chevalieri A. ochraceus					

TABLE 1.2

MÝCOTOXINS PRODUCED BY SPECIES OF THE GENUS PENICILLIUM

Mycotoxin	Producing organism	Mycotoxin	Producing organism					
Brevianamide A Cyclochlorotine Cyclopenin Citreoviridin Citrinin Erythroskyrine Griseofulvin Islanditoxin Janth trems Luteos yrin Mycophenolic acid B-Nitropropanoic acid Batulin Baxilline	P. viridicatum P. islandicum F. viridicatum F. citreo-viride P citrinum, etc. P islandicum P gr.seofulvum P is andicum P janthinellum P. slandicum P. evi- compactum, etc. P. atrovenetum P. u ticae, etc. P. pax lli	Penicilic acid Penitrems PR toxin Roquefortine Rubratoxins Rubrosul hin Rugulosin Secalonic acids Verruculogen Viomellein Viridicatin Viridicatumtoxin Xanthocillin X Xanthomegnin Xanthoviridicatins	P. puberulum, etc P. crustosum P. roqueforti P. roqueforti P. rubrum P. viridicatum P. rugulosum P. oxalicum P. verruculogenum P. vir dicatum P. viridicatum					

MYCOTOXINS PRODUCED BY VARIOUS GENERA

TABLE 1.3

Mycotoxin	Producing organism	Mycotoxin	Produ .ng organ sm					
Alternariol Butenolide Chaetog obosins Cytochalasins Diplodiol Mollicellins	Alternaria spp. Fusarium trivinctum Chaetomium globosum Phoma exigua, etc. Diplodia macrospora Chaetomium mole cellum	Paspalicine Paspaline Pat lin Tenuazonic acid Trichothecenes Zearalenone	Clavice s paspali Clavi eps paspali Bysso anys f lva Altern ria spp Fusar um. Myr t.ecium, etc Gibberella zeae					

tion of foods in restaurants and homes, and until the food is con umed. It should be added that mycotoxin-contaminated feed has caused serious economic losses, of which the Turkey X disease is a well known example.

1.3. NATURAL OCCURRENCE OF TOXIGENIC FUNGI AND MYCOTOXINS

Because mycotoxins belong to environmental chemicals which can poison both man and animals (and in some instances plants), their occurrence and also he

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Occurrence of their producing fungal strains have been studied by many workers. **Toxigenic** fungi can grow and produce their toxins on growing crops, on leaves and stems, on grains and seeds, on meat and milk products, etc. In order to demonstrate the importance of the natural occurrence of toxigenic fungi and their toxins, this section presents a selection of data from the literature. **More** systematic studies of this topic have been published elsewhere $^{9-11}$.

The production of aflatoxins by Aspergillus flavus strains on agricultural crops such as groundnut, cottonseed, wheat, rice, barley, coconut, corn, dried peas, oat, sweet potato, millet and cassava is well documented 12. Several mycotoxins, including citrinin, zearalenone and trichothecenes, have been found in cereals. Some examples are given below. Citrinin was isolated from a strain of Penicillium viridicatum from a batch of barley which, on ingestion by pigs and rats, caused chronic kidney degenerat on 13.

Many trichothecene mycotoxins, when produced by *Fusaria* either in vitro or naturally, cause emesis in a variety of laboratory animals. Morooka et al. ¹⁴ isolated from fusaria-infected barley a trichothecene toxin. Yoshizawa and Morooka ¹⁵ described its structure and assigned a new trivial name, 4-deoxynivalenol. Vesonder et al. ¹⁶ isolated vomitoxin from *Fusarium*-contaminated field corn. Deoxynivalenol and vomitox n are identical ¹⁷.

Vomitoxin at levels ranging from 0.5 to 10 µg g⁻¹ was found in 24 of 52 preharvest corn samples collected in mid-October, 1977, from farms in north-west Ohio¹⁸. Ellison and Kotsoni ⁹ presented evidence that T-2 toxin was another emetic factor in mouldy co n. Of those fungi which produce T-2 toxin, Fusarium tricinatum has been recogn zed as particularly important with regard to mouldy corn toxicosis in the mid western U.S.A.²⁰. A sample of poefusarin (a mycotoxin suspected of being one of the toxins involved in alimentary toxic aleukia) contained T-2 toxin in an amount sufficient to explain the toxicity of the sample. In addition, neosolanio, T-2 tetraol and zearalenone were present in the sample²¹.

The highest known oesophageal cancer rate in Africa occurs in the southwestern districts of the Republic of Transkei. Corn is the main dietary staple in the area. Two Fusarium toxins, deoxynivalenol and zearalenone, were detected at biologically significant levels in hand-selected, visibly Fusarium-infected corn kernels. The level of natural contamination of corn kernels with both mycotoxins was considerably higher in the high-incidence area of oesophageal cancer than in the north-eastern region of the country, where the cancer rate is relatively low²².

Citrinin, implicated in porcine nephropathy 13 , has been found as a natural contaminant of corn, rice, wheat, rye, barley and oats 23 .

A wide variety of foodstuffs, including rice, soybeans, wheat, bread, flour, white and black peppercorn, beans and groundnuts support the growth of the storage mould *Penicillium islandicum*. Out of 385 samples of soybeans collected from the $\widehat{\mathbb{L}}$ S. market, 94 were contaminated with this fungus. A large number of toxic and non-toxic metabolites are known to be produced by P. islandicum. Among the potent toxins are luteoskyrin, cyclochlorotine, islanditor in and simatoxin P4.

Aspergillus flavus readily grows on groundnuts and gr ins when they are stored traditionally in the open, in hot, humid conditions. Approximate relationships have been reported between aflatoxin contamination of market food samples and the incidence of primary liver cancer in Uganda, Swaziland and Thailand. A significant correlation between the incidence of liver cancer and the amount of aflatoxin actually consumed in three different areas in Kenya has been reported 25.

Toxic fungi have been shown to occur in a variety of nuts (pistachio nuts, chestnuts). Isolates of known toxigenic fungi from p stachio nuts in Iran included Aspergillus flavus, A. ochraceus, A. versicolor and others. All isolates of A. flavus and A. parasiticus produced aflatoxin B₁ and the isolates of A. versicolor and A. nidulans produced sterigmatocystin²⁶.

Emodin, a toxic fungal metabolite which is regarded as a precursor of many fungal anthraquinones, was characterized by Wells et al 27 as a metabolite of Aspergillus wentii isolated from weevil-damaged chestnuts.

Because of their high protein content and relatively low cost, dried beans are consumed in substantial amounts. Mislivec et al ²⁸ evaluated the potential for mycotoxin production by mou ds in dried beans. The flora was dominated by species of the Aspergillus glaucus group, the toxigenic species A. ochraceus, Penicillium cyclopium and P. ciridicatum, and species of Alternaria, Cladosporium and Fusarium. Of 209 species of Aspergillus and Penicillium screened for mycotoxin production on sterile rice substrate, 114 produced one or more of the following mycotoxins: A. flavus, aflatoxins; A. ochraceus, ochratoxins; A. nidulans, A. unguis and A. versicolor, sterigmatocystin; P cyclopium, penicillic acid; P. citrinum and P. viridicatum, citrinin; and P urticae, patulin and griseofulvin.

The widespread occurren e of ochratoxin-producing fung, their ability to grow on a variety of economically important feeds and foodstuffs and the natural occurrence of ochratoxin const tute a threat to both animal and public health 29

Chemically related tremorgenic mycotoxin have been discovered in recent years in fungi isolated from ryegrass pastures in New Zealand, Australia and North America. These toxins include verruculogen, fumitremorgins, paxilline and janthitrems $^{30-32}$.

An isochroman mycotoxin was isolated from *Penicillium steckii* from mouldy millet hay suspected of causing deaths in cattle ³³. The practice of baling hay

in large round or square bales has become popular. However, as these bales are generally stored in the open, they become contaminated with fungi. A strain of Aspergillus terreus was isolated from a bale (A. terreus is known to produce the toxins citrinin and patulin) and from this strain a new toxic metabolite, terretonin, was isolated ³⁴. From extracts of cultures of Aspergillus fumigatus isolated from silage, new mycotoxins called fumitoxins were isolated in France ³⁵.

The formation of cytochalasin B by a strain of Hormiscium sp. in tomatoes 36 and in potatoes by strains of Phoma exigua var. exigua 37 has been reported. Among mould isolates from decaying tomato fruit, toxigenic strains were isolated and their toxicity was attributed to tenuazonic acid (Alternaria alternata), I-2 toxin (Fusarium sulphureum), citrinin and patulin (both from Penicillium expansum) 38.

Bacon et al.³⁹ demonstrated the ability of *Aspergillus ochraceus* to colonize poultry feed and produce penicillic acid and ochratoxin A at favourable temperatures and moisture levels.

Trichothecenes are produced by various species of fungi from the genera Fusarium, Trichothecium, Myrothecium, Trichoderma, Cephalosporium, Verticimonosporium and Stachybotrys. As mentioned above, these fungi attack many agricultural plant products. In our laboratory, from an apple with a bitter taste a strain of Trichothecium roseum was isolated which produced trichothecin and related secondary metabolites 40.

Byssochlamys fulva and its imperfect state Paecilomyces varioti have been isolated from processed fruits, cassava bread, sorghum brandy and Manitoba butter 41 . Various metabolites of B. fulva (P. varioti) have been reported, including the mycotoxins byssochlamic acid, byssotoxin A and patulin. Patulin is also produced by several fungi of Aspergillus and Penicillium. P. expansum, owing to its ability to grow at 0° C, is a major cause of loss of apples and pears in storage. Twenty-seven isolates of P. expansum obtained from Europe, Australia and North America from seven different fruit hosts all produced patulin in culture 42 . Patulin has been found in apple fruits 43 and in apple juice 44 . Wilson and Nuovo 45 found patulin in all of 60 cultures tested (all isolated from apples). Scott and Somers 46 observed stability of patulin and penicillic acid in fruit juices and flour.

While sun-drying on the tree, fig fruits are particularly vulnerable to infection and colonization by *Aspergillus flavus*. Aflatoxin accumulation equaled levels frequently reported for groundnuts and cereal grains⁴⁷.

Aspergillus oryzae is widely used for the preparation of koji, a starter in the production of soy sauce, and miso, a fermented soybean paste used in Japanese cooking (900,000 metric tons annually). Miso is prepared by first inoculating rice with A. oryzae, a combination called koji-rice, which produces an

enzyme source for subsequent admixture with soybeans. The process is analogous to that in which malt is used in brewing. The subsequent soybean-koji-rice fermentation is conducted at 27° C for 50 h. As this fungus was widely used in food processing, the organism was thoroughly investigated for production of toxins. El-Hag and Morse ⁴⁸ reviewed papers by other workers who had all concluded that A. oryzae was not an aflatoxin producer. The former authors, however, found that a variant of A. oryzae approved for use in food processing is variable and has a propensity to produce aflatoxin on other substrates.

The simultaneous production of penicillic acid and patulin by an atypical strain of $Penicillium\ roqueforti$ isolated from cheddar cheese was reported by Olivigni and Bullerman 49 . $P.\ roqueforti$, the essential fungus used in the manufacture of roquefort and other varieties of blue cheeses, has been shown to produce other toxic compounds. These include PR toxin 50 and roquefortine, which has been detected in blue cheese samples from different countries 51 . Orth 52 reported finding PR toxin-producing $P.\ roqueforti$ isolated in nut ice cream, a dessert cream and on a red wine bottle cork. The significance of these toxins in terms of human health is unclear, as blue-veined cheeses have been consumed for centuries without apparent ill-effects. However, the potential carcinogenic nature of penicillic acid and patulin 53 possibly makes them a cause for concern.

Cyclopiazonic acid production by P. comemberti and the natural occurrence of this mycotoxin in cheese has also been reported 54 , together with other findings of penitrem A^{55} and sterigmatocystin 56 .

Several investigators have isolated toxigenic fungi from meat products. Halls and Ayres or reported that isolates of Aspergillus versicolor from country-cured ham were capable of producing sterigmatocystin. Three of these isolates were tested and found to produce sterigmatocystin on country-cured ham. Toxigenic aspergilli and penicillia were isolated and identified from country-cured hams and fermented sausages by other workers. Aflatoxin-, ochratoxin-, strigmato-cystin- and citrinin-producing strains were isolated from country-cured hams. Wu et al. solated cultures of Aspergillus and Penicillium from aged, cured meats and tested their toxicity to chicken embryos. Several isolates of A. ruber, A. repens, A. sydowi, A. restrictus, A. amstelodami, A. chevalieri and A. fumigatus exhibited toxicity. Similarly, some isolates of P. notatum, P. brevi-compactum and Penicillium spp. were found to be highly toxic. In these investigations, there was no direct or indirect evidence that aged, cured meats contain toxic metabolites.

However, two strains of *A. ochraceus* isolated from country-cured hams produced ochratoxin A and B on rice, defatted groundnut meal and country-cured ham. After incubation of ham for 21 days, one third of the toxin was found in the mycelial mat on the ham surface, whereas two thirds had penetrated into the meat ⁵⁹.

From contaminated candy, we isolated a toxigenic fungus identified as Aspergillus repens 60. Apart from antibacterial and antifungal activity, crude mycelial extracts of this fungus were toxic to Artemia salina larvae and HeLa cells. Of the six metabolites, isolated and identified as echinulin, physcion, erythroglaucin, flavoglaucin, asperentin (cladosporin) and asperentin 8-methyl ether, the broadest activity spectrum was displayed by asperentin. The latter, at a concentration of 86 μg ml $^{-1}$, caused 50% mortality in *A. salina* larvae. The highest toxicity towards HeLa cells was found in physcion, which caused 50% growth inhibition at a concentration of 0.1 ug ml^{-1} .

We could conclude that the natural occurrence of toxigenic fungi is very broad. However, one has to distinguish between the natural occurrence of their toxins and the production of these toxins under laboratory conditions. The best conclusion seems to be the following statement concerning the natural occurrence of aflatoxins (appliable also to other mycotoxins) in a recent book 61 .

"Aflatoxins arise naturally when a toxin-producing strain of Aspergillus flavus 'or A. parasiticus grows on a substrate in a geographical area where environmental conditions are suitable for the development of the mould. Many strains of A. flavus do not produce aflatoxins, so the mere presence of mouldiness is not, of itself, indicative of toxin production. The natural occurrence of aflatoxins can only be described as such when the initial inoculation by the mould spores and subsequent mould development are a natural sequence of events. In view $\dot{ extstyle 6} extstyle f$ the fact that the extstyle A. extstyle flavus group of moulds is ubiquitous and capable of development over a wide range of temperature on substrates of high carbohydrate content, agricultural commodities and their products are most vulnerable to contamination. Because of this vulnerability a world-wide effort, involving scientists of many disciplines, was initiated to establish the conditions under which A. flavus produces aflatoxins. Research was also directed to establishing what foods are potentially at risk to A. flavus infection during their production. Much research work has entailed the deliberate infection of commodities with isolates of A. flavus, and it is important to make a distinction between this method of aflatoxin production and the natural occurrence of the toxin referred to above".

REFERENCES

¹ J. Forgacs and W.T. Carll, Adv. Vet. Sci., 7 (1962) 273-382.

¹ J. Forgacs and W.I. Carii, Adv. vet. Sci., / (1902) 2/3-302.
2 H.G. Floss, Tetrahedron, 32 (1976) 873-912.
3 C.M. Curtin, III. Vet., 10 (1968) 23-27.
4 J.C. Percival and R.H. Thornton, Nature (London), 182 (1958) 1095-1096.
5 R.L.M. Synge and D.P. White, Chem. Ind. (London), (1959) 1546-1547.
6 M. Saito, M. Enomoto and T. Tatsuno, in A. Ciegler, S. Kadis and S.J. Ajl (Editors), Microbial Toxins, Vol. VI, Fungal Toxins, Academic Press, New York, London 1971 pp. 200-380 London, 1971, pp. 299-380.

7 A.Z. Joffe, in A. Ciegler, S. Kadis and S.J. Ajl (Editors), Microbial Toxins, Vol. VII, Algal and Fungal Toxins, Academic Press, New York, London, 1971, pp. 139-189.

8 A.A. Stark and A.L. Demain, ASM News, 46 (1980) 80-83.

9 K. Uraguchi and M. Yamazaki (Editors), Toxicology, Biochemistry and Pathology of Mycotoxins, Halsted Press, New York, 1978.

10 I.F.H. Purchase (Editor), Mycotoxins, Elsevier, Amsterdam, 1974.

11 J.V. Rodricks, C.W. Heseltine and M.A. Mehlman (Editors), Mycotoxins in Human and Animal Health, Pathotox, Park Forest South, 1977.

12 L.A. Goldblatt (Editor), Aflatoxin, Academic Press, New York, 1969.

13 P. Krogh, E. Hasselager and P. Friis, Acta Pathol. Microbiol. Scand., Sect. B, 78 (1970) 401-413.

14 N. Morooka, N. Uratsui, T. Yoshizawa and H. Yamamoto, Jap. J. Food Hyg., 13 (1972) 368-375.

15 T. Yoshizawa and N. Morooka, Agr. Biol. Chem., 37 (1973) 2933-2934.

- 16 R.F. Vesonder, A. Ciegler and A.H. Jensen, Appl. Microbiol., 25 (1973) 1008-1010.
- · 17 R.F. Vesonder, A. Ciegler, H. Jensen, W.K. Rohwedder and D. Weisleder, Appl. Environ. Microbiol., 31 (1976) 280-285. 18 R.F. Vesonder, A. Ciegler, R.F. Rogers, K.A. Burbridge, R.J. Bothast and

A.H. Jensen, Appl. Environ. Microbiol., 36 (1978) 885-888.

19 R.A. Ellison and F.N. Kotsonis, Appl. Microbiol., 26 (1973) 540-543.

20 E.B. Smalley, J. Amer. Vet. Med. Ass., 163 (1973) 1278-1281. 21 C.J. Mirocha and S. Pathre, Appl. Microbiol., 26 (1973) 719-724.

- 22 W.F.O. Marasas, S.J. van Rensburg and C.J. Mirocha, Agr. Food Chem., 27 (1979) 1108-1112.
- 23 L.K. Jackson and A. Ciegler, Appl. Environ. Microbiol., 36 (1978) 408-411.
- 24 A.C. Ghosh, A. Manmade, J.M. Townsend, A. Bousquet, J.P. Howes and A.L. Demain, Appl. Environ. Microbiol., 35 (1978) 1074-1078.

A.J. Zuckerman, Nature (London), 256 (1975) 540.

- 26 H. Mojtahedi, C.J. Rabie, A. Lüben, M. Steyn and D. Danesh, Mycopathologia, **67** (1979) 123-127.
- 27 J.M. Wells, R.J. Cole and J.W. Kirksey, Appl. Microbiol., 30 (1975) 26-28
- 28 P.B. Mislivec, C.T. Dieter and V.R. Bruce, Appl. Microbiol., 29 (1975) 522-
- 29 W.E. Huff, R.D. Wyatt and P.B. Hamilton, Appl. Microbiol., 30 (1975) 48-51. 30 R.T. Gallagher and G.C.M. Latch, Appl. Environ. Microbiol., 33 (1977) 730-731.
- 31 P.A. Cockrum, C.C.J. Culvenor, J.A. Edgar and A.L. Payne, J. Nat. Products, 42 (1979) 534-536.
- 32 R.T. Gallagher, G.C.M. Latch and R.G. Keogh, Appl. Environ. Microbiol., 39 (1980) 272-273.
- 33 R.H. Cox, O. Hernandez, J.W. Dorner, R.J. Cole and D.I. Fennell, Agr. Food Chem., 27 (1979) 999-1001.
- 34 J.P. Springer, J.W. Dorner, R.J. Cole and R.H. Cox, J. Org. Chem., 44 (1979) 4852-4854.
- 35 J.P. Debeaupuis and P. Lafont, Appl. Environ. Microbiol., 36 (1978) 8-10.

36 A. Pribela, J. Tomko and L. Dolejs, Phytochemistry, 14 (1975) 285.

- 37 P.M. Scott, J. Harwig, Y.K. Chen and B.P.C. Kennedy, J. Gen. Microbiol., 87 (1975) 177-180.
- 38 J. Harwig, P.M. Scott, D.R. Stoltz and B.J. Blanchfield, Appl. Environ. Microbiol., 38 (1979) 267-274.
- 39 C.W. Bacon, J.G. Sweeney, J.D. Robbins and D. Burdick, Appl. Microbiol., 26 (1973) 155-160.

40 V. Betina and M. Vanková, Biológia (Bratislava), 32 (1977) 943-949.

- 41 R.K. Kramer, N.D. Davis and V.L. Diener, Appl. Microbiol., 29 (1975) 455-457. 42 N.E. Sommer, J.R. Buchanan and R.J. Fortlage, Appl. Microbiol., 28 (1974)
- **43** P.W. Brian, G.W. Elson and D. Lowe, Nature (London), 178 (1956) 263-264.
- 44 P.M. Scott, W.F. Miles, P. Toft and J.G. Dubé, J. Agr. Food Chem., 20 (1972) 450-451.