



Measurement and Manipulation of Intracellular Ions

Edited by

**Jacob Kraicer
S. Jeffrey Dixon**

Methods in Neurosciences



Methods in Neurosciences

Volume 27

Measurement and Manipulation of Intracellular Ions

Edited by

Jacob Kraicer

Department of Physiology

Health Sciences Centre

The University of Western Ontario

London, Ontario, Canada

and

Human Frontier Science Program

Strasbourg, France

S. Jeffrey Dixon

Division of Oral Biology and Department of Physiology

Health Sciences Centre

The University of Western Ontario


London, Ontario, Canada



ACADEMIC PRESS

San Diego New York Boston London Sydney Tokyo Toronto

Front cover photograph: Fura-2 stained neuron. Neuron from the nucleus accumbens in cell culture stained with fura-2. The image was obtained using Axon Imaging Workbench by digitizing an integrated frame (500 msec) from a CCD camera (Cohu). The cell was illuminated at 380 nm. Courtesy of J. Plant and B. A. MacVicar, University of Calgary.

This book is printed on acid-free paper. 

Copyright © 1995 by ACADEMIC PRESS, INC.

All Rights Reserved.

No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopy, recording, or any information storage and retrieval system, without permission in writing from the publisher.

Academic Press, Inc.

A Division of Harcourt Brace & Company
525 B Street, Suite 1900, San Diego, California 92101-4495

United Kingdom Edition published by
Academic Press Limited
24-28 Oval Road, London NW1 7DX

International Standard Serial Number: 1043-9471

International Standard Book Number: 0-12-185297-0

PRINTED IN THE UNITED STATES OF AMERICA

95 96 97 98 99 00 EB 9 8 7 6 5 4 3 2 1

Methods in Neurosciences

Volume 27

Measurement and Manipulation of Intracellular Ions

Methods in Neurosciences

Editor-in-Chief

P. Michael Conn

*To my mentors, John Logothetopoulos,
Claude Fortier, and Don Hatcher*

JACOB KRAICER

To my parents, Michael and Helen

S. JEFFREY DIXON

Contributors to Volume 27

Article numbers are in parentheses following the names of contributors. Affiliations listed are current.

JULIO ALTAMIRANO (19), Departamento de Neurobiología, Instituto Mexicano de Psiquiatría, México 14370 D. F., México, and Department of Physiology and Biophysics, University of Texas Medical Branch, Galveston, Texas 77555

FRANCISCO J. ALVAREZ-LEEFMANS (19), Departamento de Neurobiología, Instituto Mexicano de Psiquiatría, México 14370 D. F., México, and Departamento de Farmacología y Toxicología, Centro de Investigación y de Estudios Avanzados del IPN, 07000 D.F., México

HERMAN S. BACHELARD (8), Magnetic Resonance Center, Department of Physics, University of Nottingham, Nottingham NG7 2RD, United Kingdom

SUSAN E. BATES (7), Department of Pharmacology, United Medical and Dental Schools, St. Thomas' Hospital, London SE1 7EH, United Kingdom

MARK O. BEVENSEE (12), Department of Cellular and Molecular Physiology, Yale University School of Medicine, New Haven, Connecticut 06510

JOACHIM BIWERSI (17), Cardiovascular Research Institute, University of California, San Francisco, San Francisco, California 94143

WALTER F. BORON (12), Department of Cellular and Molecular Physiology, Yale University School of Medicine, New Haven, Connecticut 06510

JONATHAN A. COLES (18), Neurobiologie Intégrative, INSERM U.394, F-33077 Bordeaux, Cedex, France

WILLIAM E. CROWE (19), Department of Physiology, Medical College of Pennsylvania and Hahnemann University, Philadelphia, Pennsylvania 19129

CHARLES M. DEBER (4), Research Institute and Department of Biochemistry, The Hospital for Sick Children, University of Toronto, Toronto, Ontario, Canada M5G 1X8

JOACHIM W. DEITMER (14), Abteilung für Allgemeine Zoologie, FB Biologie, Universität Kaiserslautern, D-67653 Kaiserslautern, Germany

J. DEMPSTER (6), Department of Physiology and Pharmacology, University of Strathclyde, Glasgow, Scotland, United Kingdom

S. JEFFREY DIXON (9), Division of Oral Biology and Department of Physiology, Faculty of Dentistry, The University of Western Ontario, London, Ontario, Canada N6A 5C1

J. KEVIN FOSKETT (13), Division of Cell Biology, The Hospital for Sick Children, Toronto, Ontario, Canada M5G 1X8

KENNETH A. GIULIANO (1), Department of Biological Sciences, Center for Light Microscope Imaging and Biotechnology, Carnegie Mellon University, and Department of Neurological Surgery, University of Pittsburgh School of Medicine, Presbyterian University Hospital, Pittsburgh, Pennsylvania 15213

ALISON M. GURNEY (7), Department of Pharmacology, United Medical and Dental Schools, St. Thomas' Hospital, London SE1 7EH, United Kingdom

HERBERT R. HALVORSON (16), Department of Pathology, Division of Biochemical Research, Henry Ford Health Sciences Center, Detroit, Michigan 48202

J. A. HELPERN (16), Division of Medical Physics, Nathan S. Kline Institute for Psychiatric Research, Orangeburg, New York 10962

JOHN HOYLAND (6), Laboratory of Neural and Secretory Signalling, Department of Neurobiology, BBSRC Babraham Institute, Babraham, Cambridge CB2 4AT, United Kingdom

STEPHEN J. KARLIK (3), Department of Radiology, University Hospital, London, Ontario, Canada N6A 5A5

RICHARD P. KRAIG (10), Departments of Neurology, and Pharmacological and Physiological Sciences, The University of Chicago, Chicago, Illinois 60637

CHRISTOPHER D. LASCOLA (10), Department of Neurology, The University of Chicago, Chicago, Illinois 60637

P. M. LLEDO (6), Institut Alfred Fessard, CNRS, F-91198 Gif-sur-Yvette, France

ROBERT E. LONDON (15), Laboratory of Molecular Biophysics, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, North Carolina 27709

W. T. MASON (6), Laboratory of Neural and Secretory Signalling, Department of Neurobiology, BBSRC Babraham Institute, Babraham, Cambridge CB2 4AT, United Kingdom

MARY E. MORRIS (2), Departments of Anesthesia, Medicine and Pharmacology, University of Ottawa, Ottawa, Ontario, Canada K1H 8M5

THOMAS MUNSCH (14), Institut für Physiologie, Medizinische Fakultät, Universität Magdeburg, D-39120 Magdeburg, Germany

ELIZABETH MURPHY (15), Laboratory of Molecular Biophysics, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, North Carolina 27709

OGNEN A. C. PETROFF (11), Department of Neurology, Yale University School of Medicine, New Haven, Connecticut 06510

DOUGLAS R. PFEIFFER (4), Department of Medical Biochemistry, Ohio State University, Columbus, Ohio 43210

JAMES W. PRICHARD (11), Department of Neurology, Yale University School of Medicine, New Haven, Connecticut 06510

MARLI A. ROBERTSON (13), Division of Gastroenterology and Nutrition, The Hospital for Sick Children, Toronto, Ontario, Canada M5G 1X8

GERRY A. SMITH (8), Department of Biochemistry, University of Cambridge, Cambridge CB2 1QW, United Kingdom

D. LANSING TAYLOR (1), Department of Biological Sciences, Center for Light Microscope Imaging and Biotechnology, Carnegie Mellon University, Pittsburgh, Pennsylvania 15213

A. S. VERKMAN (17), Departments of Medicine and Physiology, University of California, San Francisco, San Francisco, California 94143

DAVID A. WILLIAMS (5), Muscle and Cell Physiology Laboratory, Department of Physiology, The University of Melbourne, Parkville, Victoria 3052, Australia

JOHN X. WILSON (9), Department of Physiology, The University of Western Ontario, London, Ontario, Canada N6A 5C1

G. ANDREW WOOLLEY (4), Department of Chemistry, University of Toronto, Toronto, Ontario, Canada M5S 1A1

R. ZOREC (6), Institute of Pathophysiology, School of Medicine, University of Ljubljana, Ljubljana, Slovenia

Preface

Neuroscientists, as do other biologists, recognize the crucial importance of intracellular ions in cellular physiology and pathology. Under resting conditions, cytosolic ion concentrations are closely regulated by intracellular buffers and membrane transport systems. Changes in ion concentrations contribute to the regulation of numerous cell functions, including metabolism, secretion, motility, membrane permeability, cell proliferation, and apoptosis. Phenomena such as long-term potentiation and depression and excitotoxic cell death are thought to be mediated through changes in intracellular ion concentrations. To understand these processes, it is important that experimental biologists monitor changes in intracellular ion levels in response to various physiological and pathological stimuli. Similarly, it is crucial to experimentally manipulate intracellular ion concentrations and observe the resulting effects on cell function.

The purpose of this volume is to provide the reader with expert guidance on practical methods for measuring and manipulating cytosolic ionic activities. The first four chapters provide an overview of the use of optical probes, ion-selective microelectrodes and magnetic resonance spectroscopy for measuring ion levels, and the use of ionophores for manipulating intracellular ion concentrations. These are followed by chapters presenting relevant techniques, grouped according to the ion of interest (Ca^{2+} , H^+ , Na^+ , Mg^{2+} , and Cl^-). We have not attempted to include chapters on all available techniques or even all biologically relevant ions. Emphasis has been placed on widely studied ions, such as Ca^{2+} , and techniques that can be easily modified to measure related ionic species. The concluding chapter presents methods for the measurement of cell volume, the control of which is intimately linked to regulation of intracellular ion levels. In many cases, the techniques presented can be applied only to *in vitro* preparations; however, where possible, techniques for *in vivo* measurement of ion levels have also been included. Although many of the biological preparations described in this volume are of neural origin, most techniques are readily applicable to other tissues. Therefore, we hope that this volume will be of use to all cell biologists with interests in the role of cytosolic ions in cellular function, regulation, and pathology.

We thank the authors for their excellent contributions. Their enthusiasm and cooperation have made our role as editors a pleasant task. We especially thank Drs. Mary E. Morris and Stephen J. Karlik for their invaluable assis-

tance in editing the chapters on ion-selective microelectrodes and NMR spectroscopy, respectively. Last, we thank Dr. P. Michael Conn, Editor-in-Chief of *Methods in Neurosciences*, and Shirley Light of Academic Press for their assistance and encouragement.

JACOB KRAICER
S. JEFFREY DIXON

Methods in Neurosciences

- Volume 1 Gene Probes
Edited by P. Michael Conn
- Volume 2 Cell Culture
Edited by P. Michael Conn
- Volume 3 Quantitative and Qualitative Microscopy
Edited by P. Michael Conn
- Volume 4 Electrophysiology and Microinjection
Edited by P. Michael Conn
- Volume 5 Neuropeptide Technology: Gene Expression and Neuropeptide Receptors
Edited by P. Michael Conn
- Volume 6 Neuropeptide Technology: Synthesis, Assay, Purification, and Processing
Edited by P. Michael Conn
- Volume 7 Lesions and Transplantation
Edited by P. Michael Conn
- Volume 8 Neurotoxins
Edited by P. Michael Conn
- Volume 9 Gene Expression in Neural Tissues
Edited by P. Michael Conn
- Volume 10 Computers and Computations in the Neurosciences
Edited by P. Michael Conn
- Volume 11 Receptors: Model Systems and Specific Receptors
Edited by P. Michael Conn
- Volume 12 Receptors: Molecular Biology, Receptor Subclasses, Localization, and Ligand Design
Edited by P. Michael Conn
- Volume 13 Neuropeptide Analogs, Conjugates, and Fragments
Edited by P. Michael Conn
- Volume 14 Paradigms for the Study of Behavior
Edited by P. Michael Conn
- Volume 15 Photoreceptor Cells
Edited by Paul A. Hargrave
- Volume 16 Neurobiology of Cytokines (Part A)
Edited by Errol B. De Souza
- Volume 17 Neurobiology of Cytokines (Part B)
Edited by Errol B. De Souza
- Volume 18 Lipid Metabolism in Signaling Systems
Edited by John N. Fain

- Volume 19 Ion Channels of Excitable Membranes
Edited by Toshio Narahashi
- Volume 20 Pulsatility in Neuroendocrine Systems
Edited by Jon E. Levine
- Volume 21 Providing Pharmacological Access to the Brain: Alternate Approaches
Edited by Thomas R. Flanagan, Dwaine F. Emerich, and Shelley R. Winn
- Volume 22 Neurobiology of Steroids
Edited by E. Ronald de Kloet and Win Sutanto
- Volume 23 Peptidases and Neuropeptide Processing
Edited by A. Ian Smith
- Volume 24 Neuroimmunology
Edited by M. Ian Phillips and Dwight Evans
- Volume 25 Receptor Molecular Biology
Edited by Stuart C. Sealfon
- Volume 26 PCR in Neuroscience
Edited by Gobinda Sarkar
- Volume 27 Measurement and Manipulation of Intracellular Ions
Edited by Jacob Kraicer and S. Jeffrey Dixon
- Volume 28 Quantitative Neuroendocrinology (in preparation)
Edited by Michael L. Johnson and Johannes D. Veldhuis

Table of Contents

Contributors to Volume 27	ix
Preface	xiii
Volumes in Series	xv
1. Light-Optical-Based Reagents for the Measurement and Manipulation of Ions, Metabolites, and Macromolecules in Living Cells	1
<i>Kenneth A. Giuliano and D. Lansing Taylor</i>	
2. Use of Ion-Selective Microelectrodes for Recording Intracellular Ion Levels	17
<i>Mary E. Morris</i>	
3. Use of NMR Spectroscopy to Measure Intracellular Ion Concentrations	39
<i>Stephen J. Karlik</i>	
4. Use of Ionophores for Manipulating Intracellular Ion Concentrations	52
<i>G. Andrew Woolley, Douglas R. Pfeiffer, and Charles M. Deber</i>	
5. Fluorescence Imaging of Cytosolic Calcium: An Introduction to Basic Experimental Principles	69
<i>David A. Williams</i>	
6. Fluorescence Measurements of Cytosolic Calcium: Combined Photometry with Electrophysiology	81
<i>W. T. Mason, J. Dempster, R. Zorec, John Hoyland, and P. M. Lledo</i>	
7. Use of Chelators and Photoactivatable Caged Compounds to Manipulate Cytosolic Calcium	123
<i>Alison M. Gurney and Susan E. Bates</i>	
8. Measurement of Cytosolic Calcium by ^{19}F NMR	153
<i>Gerry A. Smith and Herman S. Bachelard</i>	
9. Fluorescence Measurement of Cytosolic pH in Cultured Rodent Astrocytes	196
<i>S. Jeffrey Dixon and John X. Wilson</i>	
10. Brain Cell Acid–Base Changes Measured with Ion-Selective Microelectrodes	214
<i>Christopher D. Lascola and Richard P. Kraig</i>	

11. Measurement of Cytosolic pH by Nuclear Magnetic Resonance Spectroscopy <i>Ogden A. C. Petroff and James W. Prichard</i>	233
12. Manipulation and Regulation of Cytosolic pH <i>Mark O. Bevensee and Walter F. Boron</i>	252
13. Fluorescence Measurements of Cytosolic Sodium Concentration <i>Marli A. Robertson and J. Kevin Foskett</i>	274
14. Measurement of Cytosolic Sodium Using Ion-Selective Microelectrodes <i>Joachim W. Deitmer and Thomas Munsch</i>	289
15. A Practical Guide to the Use of Fluorescent Indicators for the Measurement of Cytosolic Free Magnesium <i>Elizabeth Murphy and Robert E. London</i>	304
16. Measurement of Cytosolic Magnesium by ^{31}P NMR Spectroscopy <i>J. A. Helpen and Herbert R. Halvorson</i>	319
17. Chloride-Sensitive Fluorescent Indicators <i>A. S. Verkman and Joachim Biwersi</i>	328
18. Measurement of Cytosolic Chloride Activity by Ion-Selective Microelectrodes <i>Jonathan A. Coles</i>	340
19. Use of Ion-Selective Microelectrodes and Fluorescent Probes to Measure Cell Volume <i>Francisco J. Alvarez-Leefmans, Julio Altamirano, and William E. Crowe</i>	361
Index	393

[1] Light-Optical-Based Reagents for the Measurement and Manipulation of Ions, Metabolites, and Macromolecules in Living Cells

Kenneth A. Giuliano and D. Lansing Taylor

The chemical processes that comprise living organisms are the result of the precise orchestration of ions, metabolites, macromolecules, macromolecular assemblies, and organelles in time and space. Regulation of free ion concentration is crucial to processes that include maintenance of membrane potential, control of biochemical energetics, and mediation of signal transduction. Hence, we begin this survey by describing the progress made in the design and use of fluorescent indicators of the major intracellular free ion concentrations. Next, the development of fluorescent analogs of macromolecules, fluorogenic substrates, and protein-based optical biosensors is discussed. Then, we describe how free ion indicators are coupled with other fluorescence-based reagents in the same living cell to provide an understanding of the complex temporal and spatial relationships between molecular processes. Finally, we discuss the progress and the prospects of what we believe to be an exciting application of optical-based reagents—the manipulation of specific molecular events using photomodulated ions, metabolites, and macromolecules. The use of photomodulated reagents to change the chemical and molecular properties of cells coupled with the use of reagents that detect specific chemical and molecular events in living cells will propel us toward the dissection of the myriad of chemical reactions that define cellular physiology. Figure 1 portrays in graphical form how cells can be used as living cuvettes for the measurement and manipulation of ions, metabolites, macromolecules, macromolecular assemblies, and organelles.

Fluorescent Indicators of Intracellular Ion Concentration: Powerful Tools for Describing Cellular Physiology

Rationale for Using Fluorescent Indicators in Living Cells

Cells contain endogenous fluorescent compounds such as the nicotinamide adenine dinucleotides, the flavins, and the fluorophoric amino acids trypto-

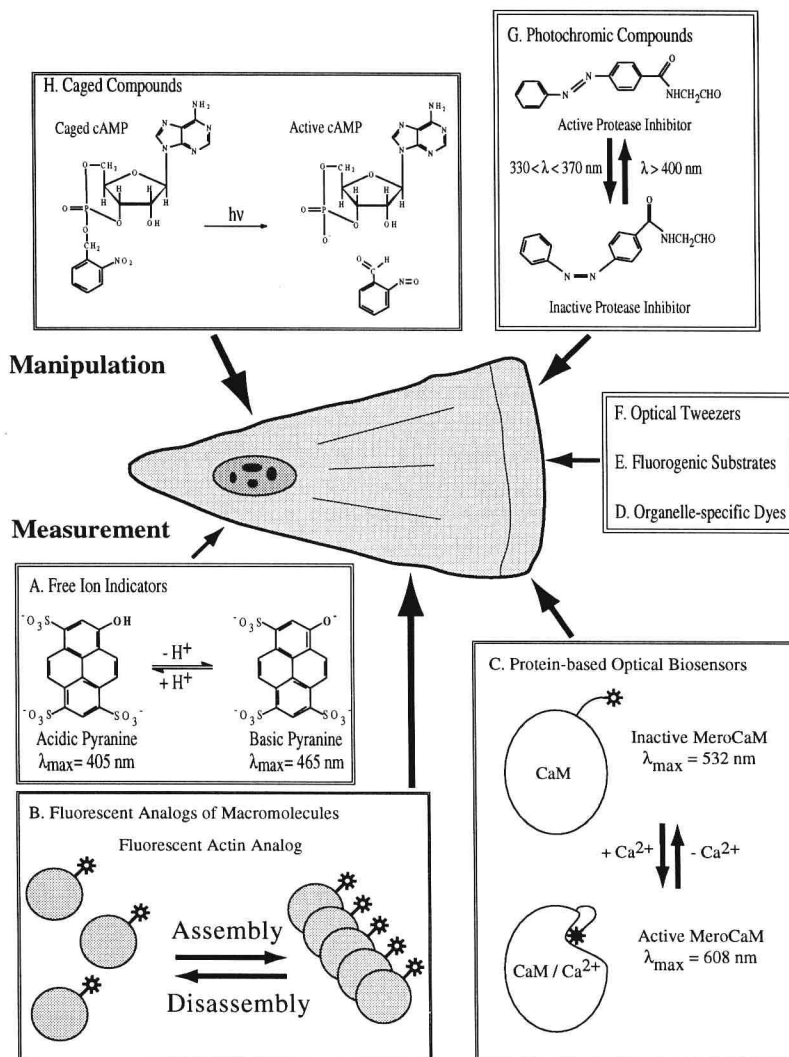


FIG. 1 Cells as living microcuvettes: Measurement and manipulation of chemical and molecular activities in single living cells. The potential to measure and manipulate several specific molecular processes in the same cell is being realized. Optical-based methods for the measurement (A–E) and manipulation (F–H) of cellular components are depicted surrounding a living cell. Examples of several classes of optical reagents are shown. These include (A) pyranine as a fluorescent indicator of pH_i [K. A. Giuliano and R. J. Gillies, *Anal. Biochem.* **167**, 362 (1987)]; (B) fluorescent analogs of actin to measure the dynamics of the actin cytoskeleton [Y. L. Wang and D. L.