

## Liquid Chromatography

Fundamentals and Instrumentation

Salvatore Fanali Paul R. Haddad Colin F. Poole Peter Schoenmakers David Lloyd

# LIQUID CHROMATOGRAPHY: FUNDAMENTALS AND INSTRUMENTATION



DAVID LLOYD





Elsevier

225, Wyman Street, Waltham, MA 02451, USA The Boulevard, Langford Lane, Kidlington, Oxford OX5 1GB, UK Radarweg 29, PO Box 211, 1000 AE Amsterdam, The Netherlands

Copyright © 2013 Elsevier Inc. All rights reserved.

No part of this publication may be reproduced, stored in a retrieval system or transmitted in any form or by any means electronic, mechanical, photocopying, recording or otherwise without the prior written permission of the publisher Permissions may be sought directly from Elsevier's Science & Technology Rights Department in Oxford, UK: phone (+44) (0) 1865 843830; fax (+44) (0) 1865 853333; email: permissions@elsevier.com. Alternatively you can submit your request online by visiting the Elsevier web site at http://elsevier.com/locate/ permissions, and selecting Obtaining permission to use Elsevier material

#### Notice

No responsibility is assumed by the publisher for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions or ideas contained in the material herein. Because of rapid advances in the medical sciences, in particular, independent verification of diagnoses and drug dosages should be made

Library of Congress Cataloging-in-Publication Data Application Submitted

British Library Cataloguing in Publication Data A catalogue record for this book is available from the British Library

ISBN: 978-0-12-415807-8

For information on all Elsevier publications visit our web site at store.elsevier.com

Printed and bound in USA 13 14 15 10 9 8 7 6 5 4 3 2 1

> Working together to grow libraries in developing countries

www.elsevier.com | www.bookaid.org | www.sabre.org

ELSEVIER

#### LIQUID CHROMATOGRAPHY: FUNDAMENTALS AND INSTRUMENTATION

#### Contributors

- Z. Aturki Institute of Chemical Methodologies, Italian National Council of Research (IMC-CNR), Area della Ricerca di Roma I, Rome, Italy
- N. Bohni School of Pharmaceutical Sciences, EPGL, University of Geneva, University of Lausanne, Geneva, Switzerland
- A. Cavazzini Department of Chemistry and Pharmaceutical Sciences, University of Ferrara, Ferrara, Italy
- S. Crotti DAIS, Cà Foscari University, Venezia, Italy
- G. D'Orazio Institute of Chemical Methodologies, Italian National Council of Research (IMC-CNR), Area della Ricerca di Roma I, Rome, Italy
- J.W. Dolan LC Resources, Inc., Walnut Creek, CA, USA
- A.S. Edison Department of Biochemistry and Molecular Biology and National High Magnetic Field Laboratory, University of Florida, Gainesville, Florida
- S. Fanali Institute of Chemical Methodologies, Italian National Council of Research (IMC-CNR), Area della Ricerca di Roma I, Rome, Italy
- A. Felinger Department of Analytical and Environmental Chemistry, University of Pécs, Pécs, Hungary
- F. Foret Institute of Analytical Chemistry ASCR, Brno, Czech Republic
- T. Fornstedt Department of Engineering and Chemical Sciences, Karlstad University, Karlstad, Sweden; Analytical Chemistry, Department of Chemistry BMC, Uppsala University, Uppsala, Sweden
- P. Forssén Department of Engineering and Chemical Sciences, Karlstad University, Karlstad, Sweden
- M.C. García-Álvarez-Coque Department of Analytical Chemistry, University of València, Spain
- I. Isak CNR-ISTM, Corso Stati Uniti 4, Padova, Italy
- R. Kaliszan Department of Biopharmaceutics and Pharmacodynamics, Medical University of Gdańsk and Department of Biopharmacy, Collegium Medicum in Bydgoszcz, Nicolaus Copernicus University in Toruń, Gdańsk, Poland
- M. Kaspereit Friedrich-Alexander-Universität Erlangen-Nürnberg, Lehrstuhl für Thermische Verfahrenstechnik, Erlangen, Germany
- J. Kuligowski Department of Analytical Chemistry, University of Valencia, Edificio Jerónimo Muñoz, Burjassot, Spain
- S. Lamotte Competence Center Analytics, Ludwigshafen, Germany
- B. Lendl Institute of Chemical Technologies and Analytics, Vienna University of Technology, Vienna, Austria

- E. Machtejevas Product Managment Analytical Chemistry, Merck Millipore, Merck KGaA, Darmstadt, Germany
- M. Macka University of Tasmania, Hobart, Australia
- K. Ndjoko-Ioset School of Pharmaceutical Sciences, EPGL, University of Geneva, Geneva, Switzerland
- P.N. Nesterenko ACROSS, University of Tasmania, Hobart, Tasmania, Australia
- B. Paull ACROSS, University of Tasmania, Hobart, Tasmania, Australia
- J.A. Queiroz Department of Chemistry, University of Beira Interior, Covilhã, Portugal
- G. Quintás Leitat Technological Center, Bio In Vitro Division, Terrassa, Spain
- G. Ramis-Ramos Department of Analytical Chemistry, University of Valencia, Spain
- A. Rocco Institute of Chemical Methodologies, Italian National Council of Research (IMC-CNR), Area della Ricerca di Roma I, Rome, Italy
- J. Samuelsson Department of Engineering and Chemical Sciences, Karlstad University, Karlstad, Sweden
- A. Seidel-Morgenstern Otto-von-Guericke-Universität Magdeburg, Institut für Verfahrenstechnik: Max-Planck-Institut für Dynamik Komplexer Technischer Systeme, Magdeburg, Germany
- J.G. Shackman Bristol-Myers Squibb, New Brunswick, NJ
- P. Smejkal Institute of Analytical Chemistry ASCR, Brno, Czech Republic; University of Tasmania, Hobart, Australia
- L.R. Snyder LC Resources, Inc., Walnut Creek, CA, USA
- **A.M. Striegel** Analytical Chemistry Division, National Institute of Standards and Technology, Gaithersburg, MD
- C.T. Tomaz CICS-UBI—Health Sciences Research Centre, University of Beira Interior, Covilhã, Portugal
- J.R. Torres-Lapasió Department of Analytical Chemistry, University of València, Spain
- P. Traldi CNR-ISTM, Corso Stati Uniti 4, Padova, Italy
- K.K. Unger Institute of Inorganic Chemistry and Analytical Chemistry, Johannes Gutenberg-University, Duesbergweg, Germany
- J.-L. Wolfender School of Pharmaceutical Sciences, EPGL, University of Geneva, University of Lausanne, Geneva, Switzerland

#### Contents

Milestones in the Development of Liquid Chromatography 1
 L.R. SNYDER AND J.W. DOLAN

 1.1. Introduction 2
 1.2. HPLC Theory and Practice 4
 1.3. Columns 6
 1.4. Equipment 9
 1.5. Detectors 12
 Apologies and Acknowledgements 13
 References 14

#### Contributors xi

2. Kinetic Theories of Liquid Chromatography 19 A. FELINGER AND A. CAVAZZINI
2.1. Introduction 20
2.2. Macroscopic Kinetic Theories 20 2.3. Microscopic Kinetic Theories 30
2.4. Comparison of the Microscopic and Macroscopic Models 38
References 39
References 39
3. Column Technology in Liquid Chromatography 41
K.K. UNGER, S. LAMOTTE AND E. MACHTEJEVAS
3.1. Introduction 42
3.2. Column Design and Hardware 44
3.3. Column Packing Materials and Stationary Phases 49
3.4. Column Systems and Operations 63
3.5. Chromatographic Column Testing and Evaluation 67

3.6. Column Maintenance and Troubleshooting 70
3.7. Today's Column Market—an Evaluation, Comparison, and Critical Appraisal 74
References 83

4.	Secondary	Chemical	Equilibria	in	Reversed	l-Phase	Liquid
	Chromatography 87						

M.C. GARCÍA-ÁLVAREZ-COQUE AND J.R. TORRES-LAPASIÓ

4.1. Introduction 88

4.2. Acid-Base Equilibria 89

4.3. Ion-Interaction Chromatography 9

4.4. Micellar Liquid Chromatography

4.5. Metal Complexation 100

Further Reading 103

#### 5. Hydrophilic Interaction Liquid Chromatography 105 A. CAVAZZINI AND A. FELINGER

5.1. Introduction 105

5.2. Principles of HILIC 107

5.3. Mobile and Stationary Phases Commonly Employed in HILIC 110 5.4. Application Examples 114

References 116

#### 6. Hydrophobic Interaction Chromatography 121

C.T. TOMAZ AND J.A. QUEIROZ

6.1. Introduction 122

6.2. Principles of Hydrophobic Interaction Chromatography 122

6.3. Main Factors that Affect Hydrophobic Interaction Chromatography 12

6.4. Purification Strategies 130

6.5. Practical Aspects of Hydrophobic Interaction Chromatography Purification 131

6.6. Selected Applications 132

6.7. Future Trends 135

References 136

#### 7. Liquid-Solid Chromatography 143

L.R. SNYDER AND I.W. DOLAN

7.1. Introduction 143

7.2. Retention and Separation 144

7.3. Method Development 149

7.4. Problems in the Use of Normal-Phase Chromatography 154

References 155

#### 3. Ion Chromatography 157

B. PAULL AND P.N. NESTERENKO

8.1. Introduction 157

8.2. Basic Principles and Separation Modes 158

CONTENTS vii

8.3. Instrumentation 165 8.4. Applications 185 References 188

#### 9. Size-Exclusion Chromatography 193

9.1. Introduction 194

A.M. STRIEGEL

9.2. Historical Background 194
9.3. Retention In Size-Exclusion Chromatography 197
9.4. Band Broadening in Size-Exclusion Chromatography 202

9.5. Resolution in Size-Exclusion Chromatography 205

9.6. Size-Exclusion Chromatography Enters the Modern Era: The Determination of Absolute Molar Mass 207

9.7. Size-Exclusion Chromatography Today: Multidetector Measurements,
 Physicochemical Characterization, Two-Dimensional Techniques 217
 9.8. Conclusions 219

Acknowledgement and Disclaimer 220 References 220

#### Solvent Selection in Liquid Chromatography 225 G. RAMIS-RAMOS AND M.C. GARCÍA-ÁLVAREZ-COQUE

10.1. Elution Strength 226

10.2. Columns and Solvents in RPLC, NPLC, and HILIC 228

10.3. Assessment of the Elution Strength 229

10.4. Schoenmakers's Rule 231

10.5. Isoeluotropic Mixtures 233

10.6. Solvent-Selectivity Triangles 234
10.7. Practical Guidelines for Optimization of Mobile Phase Composition 241
10.8. Additional Considerations for Solvent Selection 246
References 248

#### 11. Method Development in Liquid Chromatography 251

11.1. Introduction 252
11.2. Goals 253
11.3. A Structured Approach to Method Development 253
11.4. Method Development in Practice 258
11.5. Prevalidation 263
11.6. Validation 264
11.7. Documentation 265
11.8. Summary 266
References 267

viii CONTENTS

12.	Theory and Practice of	of Gradient	Elution	Liquid
	Chro	matograph	y 269	

J.W. DOLAN AND L.R. SNYDER

12.1. Introduction 269

12.2. The Effects of Experimental Conditions on Separation 27212.3. Method Development 278

12.4. Problems Associated with Gradient Elution 281 References 282

#### 13. General Instrumentation 283

J.G. SHACKMAN

13.1. Introduction 284

13.2. Solvent Source 286

13.3. Pumping Systems 286

13.4. Gradient-Elution Mixing Systems 289

13.5. Sample Injection 291

13.6. Column Compartment 293

13.7. Tubings and Fittings 294

13.8. Detector Overview 296

13.9. Ultraviolet-Visible Absorbance Detectors 298

13.10. Refractive Index Detectors 300

13.11. Evaporative Light-Scattering Detectors 301

13.12. Charged Aerosol Detectors 301

13.13. Conductivity Detectors 302

13.14. Fluorescence Detectors 302

13.15. Electrochemical Detectors 303

13.16. Other Detection Methods 303

References 304

#### Advanced Spectroscopic Detectors for Identification and Quantification: Mass Spectrometry 307

S. CROTTI, I. ISAK AND P. TRALDI

14.1. Introduction 308

14.2. Ionization Methods Suitable for LC Coupling 309

14.3. How to Increase Specificity of MS Data 317

14.4. Micro- and Nano-LC-MS 321

14.5. Capillary Electrochromatography 325

References 327

#### Advanced Spectroscopic Detectors for Identification and Quantification: FTIR and Raman 333 J. KULIGOWSKI, B. LENDL, AND G. QUINTÁS

15.1. Introduction 333 15.2. Off-Line Hyphenation 337 CONTENTS ix

15.3. On-Line Hyphenation 339 15.4. Conclusions 345 References 346

#### Advanced Spectroscopic Detectors for Identification and Quantification: Nuclear Magnetic Resonance 349

J.-L. WOLFENDER, N. BOHNI, K. NDJOKO-IOSET AND A.S. EDISON

16.1. Introduction 350
16.2. Hyphenation of NMR with HPLC 351
16.3. Advances in NMR Sensitivity 353

16.4. Strategies for Obtaining NMR Information from a Given LC Peak 35916.5. Integration with a Multiple Detection System (LC-NMR-MS) 368

16.6. Quantification Capabilities 368
16.7. Fields of Application 370
16.8. Conclusions 379
Acknowledgments 380
References 380

#### Quantitative Structure Property (Retention) Relationships in Liquid Chromatography 385

R. KALISZAN

17.1. Introduction 386
17.2. Methodology and Goals of OSRR Studies 386

17.3. Applications of QSRR in Proteomics 392

17.4. Characterization of Stationary Phases 393

17.5. QSRR and Assessment of Lipophilicity of Xenobiotics 395

17.6. QSRR Analysis of Retention Data Determined on Immobilized-Biomacromolecule Stationary Phases 399

17.7. Quantitative Retention—(Biological) Activity Relationships 400

17.8. Chemometrically Processed Multivariate Chromatographic Data in Relation to Pharmacological Properties of Drugs and

"Drug Candidates" 400

17.9. Concluding Remarks 401 Acknowledgment 402

References 402

#### Modeling of Preparative Liquid Chromatography 407 T. FORNSTEDT, P. FORSSÉN AND I. SAMUELSSON

18.1. Introduction 408
18.2. Column Model 409
18.3. Adsorption Model 410
18.4. Process Optimization of Preparative Chromatography 415
18.5. Case Example 422
References 422

X CONTENTS

19.	Process Concepts in Preparative Chromatography	427		
	M. KASPEREIT AND A. SEIDEL-MORGENSTERN			

19.1. Introduction 428

19.2. Classical Isocratic Discontinuous Elution Chromatography 42919.3. Other Discontinuous Operating Concepts 432

19.5. Onter Discontinuous Operating Concepts 432

19.4. Continuous Concepts of Preparative Chromatography 437

19.5. Optimization and Concept Comparison 446

19.6. Conclusions 448

Acknowledgments 448

Acknowledgments 448 References 449

#### 20. Miniaturization and Microfluidics 453 F. FORET, P. SMEJKAL AND M. MACKA

20.1. Introduction, Definitions, and Scope 453
20.2. Microfluidic Systems for Separations 455
20.3. Commercial Instrumentation 459
20.4. Conclusion 465
Acknowledgment 465
References 465

#### 21. Capillary Electrochromatography 469 A. ROCCO, G. D'ORAZIO, Z. ATURKI AND S. FANALI

21.1. Introduction 469
21.2. Principles of Capillary Electrochromatography 470
21.3. Instrumentation 471
21.4. Method Optimization in CEC 478
21.5. Examples of Some Recent Applications 479
21.6. Conclusions and Future Trends 486
References 487

Index 493

#### 1

### Milestones in the Development of Liquid Chromatography

L.R. Snyder, J.W. Dolan LC Resources, Inc., Walnut Creek, CA, USA

OUTLINE		
1.1. Introduction 1.1.1. Developments before 1960 1.1.2. HPLC at the Beginning	2 2 3	
1.2. HPLC Theory and Practice 1.2.1. New HPLC Modes and Techniques 1.2.2. Selection of Conditions for the Control of Selectivity	4 5 5	
1.3. Columns 1.3.1. Particles and Column Packing 1.3.2. Stationary Phases and Selectivity	6 6 9	
1.4. Equipment	9	
1.5. Detectors	12	
Apologies and Acknowledgements		
References	14	

The importance of liquid chromatography (LC), and especially high-performance LC (HPLC), in today's world hardly needs stating. It is the most widely used technique for the analysis of chemical mixtures and has contributed in a major way to science (especially the biological sciences) and everyday laboratory practice. Liquid chromatography is primarily a practical technique, so our story is limited to those innovations that

contributed significantly to its present use in "working" laboratories. In reflecting on the history of LC, it appears to us that only a few "essential" actors are in this drama: single individuals whose absence might have delayed the technique by more than a year or two. Thus, the development of present-day LC has largely been an evolutionary, rather than a revolutionary, process. Furthermore, many important innovations within the past 50 years have occurred within industrial research and development (R&D) groups, where it is often not possible to assign credit for a final product to a single person. Finally, every attempt at history suffers from incomplete and conflicting accounts of who did what—and when. In the present "history," we try to emphasize "what" and "when" rather than "who."

#### 1.1. INTRODUCTION

Several important innovations in the history of liquid chromatography are reviewed by Ettre [1]:

- Invention of chromatography in the early 1900s [2].
- Invention of partition and paper chromatography in the early 1940s [3].
- Development of ion-exchange chromatography [4] and the amino-acid analyzer during the 1950s [5].
- Invention of gel-filtration chromatography in the late 1950s [6,7].
- Development of the gel-permeation chromatograph in the early 1960s [8,9].
- Development of high-performance LC in the mid-1960s [8,10-17].

The present chapter emphasizes work on HPLC, while noting major, prior contributions that made this technique possible. Most advances in HPLC can be organized as follows:

- Development or application of basic theory, combined with empirical observations of the separation process.
- Invention of new chromatographic modes (e.g., ion-pair chromatography, hydrophilic-interaction chromatography) and the development of HPLC columns for new applications (chiral separation, large biomolecules).
- Development and improvement of equipment and columns.

#### 1.1.1. Developments before 1960

A good account of the discovery of chromatography by Tswett is given in [2] and [14, pp. 4–6]. Despite a few subsequent applications of chromatography in other laboratories [14, pp. 7–9], this technique became generally accepted only after its reintroduction in 1931 by Kuhn,

Winterstein, and Lederer [18]. The invention of *liquid-partition* chromatography was reported by Martin and Synge in 1941, followed soon after by its extension to *paper* chromatography in 1944 and the first application of *two-dimensional* chromatography [3].

Significant work on ion-exchange separation began in the 1930s, with the subsequent development and application of ion-exchange chromatography (IEC) for separation of the rare earths and transuranium elements [4]. The extension of IEC to organic compounds was next, implemented by Cohn and Samuelson [14, pp. 17–21]. By 1958, Moore, Stein, and Spackman reported the automatic analysis of amino acid samples by means of IEC [5]. Their system was a precursor of HPLC that incorporated automatic pumping, efficient IEC columns, and continuous colorimetric detection. This system was later improved and commercialized as the Beckman-Spinco model 120B amino acid analyzer.

Still another major development, in the later 1950s, was the invention of gel filtration [6,7] for the separation of large biomolecules by molecular size; this was followed a few years later by the development of gelpermeation chromatography (GPC) for the similar separation of synthetic polymers [9]. The latter then led to the development of a commercial GPC system by Waters Associates (the GPC-100 [8]), which would morph into an early commercial HPLC system (the ALC-100).

#### 1.1.2. HPLC at the Beginning

Prior to the development of the first HPLC systems, gas chromatography (GC) provided an example of what HPLC might be capable of: automation, speed, and detection sensitivity, as well as the separation of higher-boiling and thermally unstable compounds. By the early 1960s, the automation of LC had been demonstrated for amino-acid analysis and gel-permeation chromatography (Section 1.1.2). By then, it was appreciated that smaller particles in well-packed beds could increase both separation speed and efficiency. Fast separations with small-particle columns also require higher pressures to pump the mobile phase through the column. Detectors that could be used with LC for most samples presented a major challenge at this time—and for several years thereafter (Section 1.5).

Before 1965, the possibility of using HPLC for separating samples other than amino acids or polymers had undoubtedly occurred to many people. However, the exploitation of this idea required its reduction to practice, followed by the production of commercially available equipment, as in the case of the amino-acid analyzer and the gel-permeation chromatograph (Section 1.1.2). Viewed in these terms, *high-performance* LC was first reduced to practice in ~1964, in the United States under the direction of Csaba Horváth [16] and in Holland by Josef Huber (see [10, pp. 159–166

and 209–217]). The system developed by Horváth was subsequently the basis for the LCS 1000 Nucleic Acid Analyzer sold by Picker Nuclear (later acquired by Varian) and contributed to the first general-purpose HPLC (Waters ALC-100) [8]. Jack Kirkland had visited Huber's lab in 1964 and subsequently began an HPLC program at DuPont [19–22], which culminated in the Model 820 at about the same time as the ALC-100. By 1970, sales of HPLC systems were dominated by Waters Associates and Du Pont. Superficially porous Zipax [22] was initially the most popular column packing. In our opinion, Horváth, Huber, and Kirkland can be considered the "fathers" of HPLC. Some closely related work at this time by others [23–29] is also relevant to the origin of HPLC. For a description of the columns, equipment, and practice at that time, see [30].

#### 1.2. HPLC THEORY AND PRACTICE

Separation as a function of experimental conditions was understood in general terms for GC, and similar principles were expected to apply to HPLC. Resolution,  $R_s$ , can be described by the Purnell equation [31] in terms of the plate number, N, separation factor,  $\alpha$ , and retention factor, k:

$$R_s = 0.25(N^{0.5})(\alpha - 1) [k/(k+1)]$$
(1.1)

A semi-quantitative understanding of column efficiency (*plate number*, *N*) existed prior to 1965, based on the further development of the van Deemter equation for GC [32] and its extension to LC by Giddings [23]. Later work resulted in the highly useful and widely applied Knox equation [33]:

$$h = Av^{0.33} + B/v + Cv (1.2)$$

where the reduced plate height, h, is related to the reduced velocity, v, of the mobile phase. The later development of "Poppe" (or "kinetic") plots further advanced our understanding and use of column efficiency [34]. For further details on Eq. (1.2) and values of N, see Chapter 2 and [35]. When developing an HPLC procedure, the main challenge has been the selection of separation conditions for the control of peak spacing, that is, "optimum" values of  $\alpha$  (Section 1.2.2).

Basic theory played an important role in the development of HPLC, but its implementation was primarily the result of (a) the introduction of new separation modes or techniques (Section 1.2.1), (b) a better understanding of how best to vary conditions for a satisfactory separation (Section 1.2.2), and (c) improved columns (Section 1.3) and hardware (Sections 1.4 and 1.5).