Lymphokines

edited by

Edgar Pick

Maurice Landy

ADVISORY EDITOR

ACADEMIC PRESS

SUBSIDIARY OF HARCOURT BRACE JOVANOVICH, PUBLISHERS

Lymphokines

A Forum for Immunoregulatory Cell Products

EDITED BY

EDGAR PICK

Department of Human Microbiology Sackler School of Medicine Tel-Aviv University Ramat-Aviv, Tel-Aviv, Israel

ADVISORY EDITOR MAURICE LANDY

Schweizerisches Forschungsinstitut Medizinische Abteilung Davos, Switzerland

VOLUME 2

1981



ACADEMIC PRESS A Subsidiary of Harcourt Brace Jovanovich, Publishers New York London Toronto Sydney San Francisco

Formerly titled

Lymphokine Reports

A Forum for Nonantibody Lymphocyte Products

Copyright © 1981, by Academic Press, Inc. all rights reserved.

No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopy, recording, or any information storage and retrieval system, without permission in writing from the publisher.

ACADEMIC PRESS, INC. 111 Fifth Avenue, New York, New York 10003

United Kingdom Edition published by ACADEMIC PRESS, INC. (LONDON) LTD. 24/28 Oval Road, London NW1 7DX

ISSN 0197-596X

ISBN 0-12-432002-3

PRINTED IN THE UNITED STATES OF AMERICA

81 82 83 84 9 8 7 6 5 4 3 2 1

LYMPHOKINES

A FORUM FOR IMMUNOREGULATORY CELL PRODUCTS

VOLUME 2

Formerly titled

Lymphokine Reports

A Forum for Nonantibody Lymphocyte Products

EDITORIAL BOARD

Barry R. Bloom Stanley Cohen John R. David Alain L. de Weck Dudley C. Dumonde Marc Feldmann Steven B. Mizel Joost J. Oppenheim Max H. Schreier Clemens Sorg John L. Turk Emil R. Unanue Byron H. Waksman

List of Contributors

Numbers in parentheses indicate the pages on which the authors' contributions begin.

- MIKE CECKA (125), ICRF Tumour Immunology Unit, Department of Zoology, University College, London WC1E 6BT, England
- Pojen Chen (273), Department of Microbiology and Immunology, University of California, San Francisco, California 94143
- Alain L. de Weck (1), Institute of Clinical Immunology, Inselspital, Bern 3010, Switzerland
- ANN M. DVORAK (203), Departments of Pathology, Beth Israel Hospital and Harvard Medical School, and the Charles A. Dana Research Institute, Beth Israel Hospital, Boston, Massachusetts 02215
- HAROLD F. DVORAK (203), Departments of Pathology, Beth Israel Hospital and Harvard Medical School, and the Charles A. Dana Research Institute, Beth Israel Hospital, Boston, Massachusetts 02215
- VICTOR EDY* (1), Theodor Kocher Institut, 3000 Bern 9, Switzerland
- Peter Erb (125), Institute for Microbiology, University of Basel, Basel, Switzerland
- Marc Feldmann (87, 125), ICRF Tumour Immunology Unit, Department of Zoology, University College, London WC1E 6BT, England
- George E. Gifford (235), Department of Immunology and Medical Microbiology, University of Florida College of Medicine, Gainesville, Florida 32610
- JOEL W. GOODMAN (273), Department of Microbiology and Immunology, University of California, San Francisco, California 94143
- J. TERRELL HOFFELD (63), Laboratory of Microbiology and Immunology, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20205

^{*} Present address: Flow Laboratories, Inc., 7655 Old Springhouse Road, McLean, Virginia 22102.

- EVA MARIE KNIEP (1), Max-Planck Institut für Immunbiologie, Freiburg, Federal Republic of Germany
- SIRKKA KONTIAINEN (87), Department of Bacteriology and Immunology, University of Helsinki, 00290 Helsinki 29, Finland
- Sandor Lazary (1), Institute of Clinical Immunology, Inselspital, Bern 3010, Switzerland
- George K. Lewis (273), Department of Microbiology and Immunology, University of California, San Francisco, California 94143
- David Y. Liu (153), Departments of Medicine, Harvard Medical School and the Robert B. Brigham Division of Brigham and Women's Hospital, Boston, Massachusetts 02115
- ZVI METZGER (63), Laboratory of Microbiology and Immunology, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20205
- Jun Minowada (1), Roswell Park Memorial Institute, Buffalo, New York 14263
- RUTH NETA (295), Department of Microbiology, University of Notre Dame, Notre Dame, Indiana 46556
- Ranka Odavic (1), Division of Hematology, Inselspital, Bern 3010, Switzerland
- JOOST J. OPPENHEIM (63), Laboratory of Microbiology and Immunology, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20205
- NEIL S. ORENSTEIN (203), Departments of Pathology, Beth Israel Hospital and Harvard Medical School, and the Charles A. Dana Research Institute, Beth Israel Hospital, Boston, Massachusetts 02215
- URSULA OTZ (1), Institute of Clinical Immunology, Inselspital, Bern 3010, Switzerland
- Daniele Primi (273), Department of Microbiology and Immunology, University of California, San Francisco, California 94143
- Heinz G. Remold (153), Departments of Medicine, Harvard Medical School and the Robert B. Brigham Division of Brigham and Women's Hospital, Boston, Massachusetts 02115

- Ross E. Rocklin (163), Department of Medicine, Tufts University School of Medicine-New England Medical Center Hospital, Boston, Massachusetts 02111
- MICHAEL R. RUFF (235), Department of Immunology and Medical Microbiology, University of Florida College of Medicine, Gainesville, Florida 32610
- S. B. Salvin (295), Department of Microbiology, School of Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania 15261
- LAWRENCE B. SCHOOK† (1), Institute of Clinical Immunology, Inselspital, Bern 3010, Switzerland
- MAX H. SCHREIER (31), Basel Institute for Immunology, 4005 Basel, Switzerland
- Kendall A. Smith (21), The Immunology Program, Dartmouth Medical School, The Norris Cotton Cancer Center, Hanover, New Hampshire 03755
- RICHARD P. TRIGLIA (273), Department of Microbiology and Immunology, University of California, San Francisco, California 94143
- Peter Vogt (125), Institute for Microbiology, University of Basel, Basel, Switzerland
- Larry M. Wahl (179), Laboratory of Microbiology and Immunology, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20205
- Sharon M. Wahl (179), Laboratory of Microbiology and Immunology, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20205

[†] Present address: Department of Microbiology and Immunology, University of Michigan Medical School, Ann Arbor, Michigan 48109.

PREFACE XV

apparent selectivity for neoplastic cells are exciting themes for future research, as is here pointed out.

In Chapter 12 Primi *et al.* present a major and exciting technical advance in lymphokine research—the design of a method for enumerating lymphokine-secreting cells. Making use of the increasing availability of anti-lymphokine antibodies, the authors are the first to offer us a long-awaited nonimmunoglobulin equivalent of the Jerne plaque assay. They rightly point out the practically unlimited potential of this and analogous techniques, especially in light of the methodological revolution initiated by hybridoma-derived monoclonal antibodies.

The closing chapter is devoted to a key issue in lymphokine studies, the significance of the presence of lymphokines in the tissues of animals and humans. Neta and Salvin discuss the experimental and clinical demonstration of lymphokines, mostly of the inflammatory type, in blood, lymph, serous cavities, and even in the aqueous humor of the eye.

This second volume of *Lymphokines* begins more fully than its predecessor to reflect the extraordinary diversity of lymphokine research and the impressive implications these concurrent developments portend for the immunological community. It is in this context that we hope it fulfills the expectations of our audience, while accurately projecting the intentions of the Editors. I warmly thank this volume's authors for their patient and effective cooperation and the staff of Academic Press for their sustained encouragement and support.

EDGAR PICK

Preface

Volume 2 of *Lymphokines*¹ is published at a time when research on immunoregulatory cell products is penetrating every aspect of immunology. A considerable portion of the papers now being published in the major immunological journals deal with lymphokines, monokines, and other cytokines; all recent textbooks of immunology devote entire sections to soluble mediators of cellular immunity.

This volume opens with a systematic screening for lymphokine activities detectable in the supernatants of continuous lymphoid and myeloid cell lines, as described in the chapter by Schook and colleagues. Nine distinct biologic activities were examined from 24 cell lines. The effectiveness of B-cell lines in producing MIF, interferon, and mitogenic and macrophage cytotoxicity factors clearly refutes the commonly accepted but thoroughly misleading idea that lymphokines are to be defined as soluble T-cell products.

In Chapter 2 Smith reviews our present knowledge of T-cell growth factor (TCGF) as well as the prospects for future lymphokine studies stemming from the discovery and rapidly expanding application of TCGF. The conceptual revolution, precipitated by the discovery of TCGF, has infiltrated many key areas of immunological research. The discovery of TCGF enables us to maintain selected T cells proliferating continuously, a matter of enormous significance.

In Chapter 3 Schreier analyzes the central role played by T- and B-cell growth factors in the primary antibody response *in vitro*. By using cloned helper T cells, the author proposes that specific antigenic stimulation of helper T cells generates nonantigen-specific growth factors acting upon B cells stimulated by the same or an unrelated antigen.

The immune response *in vitro* is also the subject of Chapter 4 by Hoffeld and colleagues. They propose that the suppressive activity of macrophages in this system is mediated by oxygen reduction products and derivatives of arachidonic acid oxidation.

Feldmann and Kontiainen (Chapter 5) provide us with an up-to-date

¹The main title of this serial has been simplified from *Lymphokine Reports* to *Lymphokines*. Our intent is to emphasize the orientation of this publication as a wide-ranging survey of progress and the definition of new directions in lymphokine studies, rather than the mere publication of current research reports. It bears special emphasis that *lymphokine* is utilized here in its broadest connotation, as covering the entire spectrum of soluble immunoregulatory products of lymphocytes, macrophages, and other cells.

xiv PREFACE

analysis of the role of antigen-specific T-cell factors in both help and suppression. They rightly emphasize the urgent need for more evidence in support of a role of such factors in the *in vivo* reality. They also propose a model for the structure of antigen-specific factors, the essence of which is the presence of a constant region, determining the functional property of the molecule (help or suppression), and a variable region which binds to antigen.

The role of soluble factors in macrophage—T-cell interaction is discussed in Chapter 6 by Erb et al. The monokine, known as genetically related macrophage factor (GRF), is released by macrophages incubated with antigen and is distinguished by its capacity to induce antigen-specific helper T cells and by its possession of Ia antigens. The authors point out that both the Ia and the immunogenic fragments present on GRF may act in a macrophage-bound form.

In Chapter 7, the as yet little explored domain of the action mechanism of lymphokines is examined by Liu and Remold in their treatment of membrane receptors for lymphokines. In Chapter 8 Rocklin discusses the mechanism of action of leukocyte migration inhibitory factor (LIF). Liu and Remold describe in detail their own studies on the receptor for MIF/macrophage activating factor (MAF), which stemmed from the important discovery that simple sugars can block lymphokine action.

Rocklin gives a personal account of his studies of the mode of action of LIF. The possibility that a leukocyte-derived polypeptide, released under the influence of LIF, is actually mediating the cell-immobilizing effect of the mediator is considered.

The biological universality of lymphokines is aptly affirmed in the chapter by Wahl and Wahl on the modulation of connective tissue metabolism by lymphokines and monokines. The lymphokine-mediated interaction between lymphocytes and macrophages, at one end, and fibroblasts, at the other, may offer an explanation for many hitherto unexplained aspects of granuloma formation and, possibly, for the pathogenesis of some fibrotic conditions of obscure etiology.

Further proof of the penetration of lymphokine studies into many other areas of medicine is offered in Chapter 10 by Dvorak and colleagues, who analyze cytokine-like mediators secreted by malignant cells. Such mediators provide the tumor with a suitable microenvironment that neutralizes host defense mechanisms.

In Chapter 11 Ruff and Gifford offer a detailed description of tumor necrosis factor (TNF), a mediator discovered 20 years ago but representing the culmination of a research effort which started more than 100 years ago. The elucidation of its mechanism of action and of its

Contents

LIST OF CONTRIBUTORS	ix xiii
Lymphokine and Monokine Activities in Supernatants from Hum Lymphoid and Myeloid Cell Lines	an
LAWRENCE B. SCHOOK, URSULA OTZ, SANDOR LAZARY, ALAIN L. DE WECK, JUN MINOWADA, RANKA ODAVIC, EVA MARIE KNIEP, AND VICTOR EDY	
I. Introduction	1 2 8 16 17 18
KENDALL A. SMITH	
I. Introduction II. The Functional Significance of TCGF III. TCGF Production IV. TCGF Action V. Therapeutic Implications of TCGF VI. The Prospects Ahead VII. Conclusions References	21 22 23 24 27 28 29 30
The Antibody Response in Vitro: Dissection of a Complex Syste	m
MAX H. SCHREIER	
I. Introduction	31
Response to Heterologous Erythrocytes	33
IV. Specific Induction of Unprimed B Cells in Vitro	43
V. Specific T-Cell Help and Clones of Helper T Cells	50
VI. The Nature of Antigen-Activated T-Cell Help	56
VII. Conclusions and Outlook	58
References	60

vi CONTENTS

Oxygen-Derived Metabolites as Suppressors of Immune Responses $in\ Vitro$

	J. TERRELL HOFFELD, ZVI METZGER, AND JOOST J. OPPENHEIM	
	Introduction	63
YTT	Radicals	67
	Oxygen Radicals and the Primary Antibody Response in Vitro	74
LV.	Discussion	83 84
	References	04
The	Role of Antigen-Specific T-Cell Factors in the Immune Respon	nse
	MARC FELDMANN AND SIRKKA KONTIAINEN	
I.	Summary	87
	Introduction	88
	Antigen-Specific Helper Factors	91
	Antigen-Specific Suppressor Factors	102
	Antigen-Specific Factors in the Cell-Mediated Response	107
	Analysis of Monoclonal Factors	108
VΠ.	A Scheme for the Nature of Antigen-Specific Factors	109
V 1.1.1.	Role for T-Cell Factors?	116
	References	119
Acti	vation of T Cells by I-Region Products Released by Macropha	ges
	Peter Erb, Peter Vogt, Mike Cecka, and Marc Feldmann	
I.	Introduction	125
11.	Production and Assay of GRF	127
Ш.	Nature of GRF	127
	Target of GRF Activity	132
	Role of GRF in T-Cell Activation	139
VI.	What Is the Biological Significance of GRF?	146
VII.	Conclusions	149
	References	150
	Cell Surface Receptors for Lymphokines	
	DAVID Y. LIU AND HEINZ G. REMOLD	
	Introduction	153
	Studies on the Possible Cell Surface Receptor Components	
11.	Studies on the Possible Cell Surface Receptor Components for MIF and MAF	153 154

CONTENTS	vii
IV. Interferon Receptors	159 161
Properties and Mechanism of Action of Human Leukocyte Migra Inhibitory Factor (LIF)	ition
ROSS E. ROCKLIN	
1. Cell Migration Inhibition Systems 11. Production and Assay of Human LIF 111. Physicochemical Properties of LIF 112. Esterase Nature of LIF 123. V. Mechanism of Action of LIF 124. VI. Summary 125. References 126. Modulation of Fibroblast Growth and Function by Monokines as Lymphokines	163 165 167 168 173 176 176
SHARON M. WAHL AND LARRY M. WAHL	
I. Introduction . II. Monokine Regulation of Fibroblasts . III. Lymphokine Regulation of Fibroblasts IV. Role of Fibroblasts in Chronic Inflammatory Diseases V. Summary . References	179 180 188 191 196 199
Tumor-Secreted Mediators and the Tumor Microenvironment: Relationship to Immunological Surveillance	
HAROLD F. DVORAK, NEIL S. ORENSTEIN, AND ANN M. DVORAK	
I. Immunological Surveillance: Where Are the King's New Clothes? II. The Pathological Anatomy of Tumors: A Reevaluation of the Tumor	203
Microenvironment	205 216
Fibroplasia V. Summary and Conclusions References	226 227 230
Tumor Necrosis Factor	
MICHAEL R. RUFF AND GEORGE E. GIFFORD	
I. Introduction	235

viii CONTENTS

III.	Definition of TNF							239
	Priming and Elicitation of TNF in Vivo							239
	Cells Responsible for TNF Production							240
	Assays for TNF							245
	Purification and Physicochemical Properties of TNF		*			0.000	OL	249
	Mechanism of Action and Cellular Effects							251
	Prospectus							267
1.71	References							269
	neietences							209
I	A Hemolytic Plaque Assay for the Estimation Lymphokine-Secreting Cells Daniele Primi, Richard P. Triglia, Pojen Chen, George K.				IS.	A	ND	
	Joel W. Goodman						. 14.5	
	Introduction						100	273
	Experimental Design							274
	Nature of the Plaque-Forming Cells							277
	T-PFC Response to Antigenic Stimulation							282
	Functional Properties of T-PFC							283
VI.	Plaque Formation by Human Lymphotoxin-Secreting Cells							288
VII.	Concluding Remarks	. 19		8	3 1	4	×	291
	References		į.	8	į.	*		293
	Production of Lymphokines in Vivo Ruth Neta and S. B. Salvin							
I.	Introduction					a i		295
	Serum							296
III.	Body Fluids							302
	Comments							305
	References							307
			5			*		
NDE	V.							300

LYMPHOKINES, VOL. 2

Lymphokine and Monokine Activities in Supernatants from Human Lymphoid and Myeloid Cell Lines¹

LAWRENCE B. SCHOOK,**.2 URSULA OTZ,* SANDOR LAZARY,*
ALAIN L. DE WECK,* JUN MINOWADA,† RANKA ODAVIC,‡
EVA MARIE KNIEP,§ AND VICTOR EDY**

*Institute of Clinical Immunology and ‡Division of Hematology, Inselspital, Bern, Switzerland; †Roswell Park Memorial Institute, Buffalo, New York; §Max-Planck Institut für Immunbiologie, Freiburg, Federal Republic of Germany; and **Theodor Kocher Institut, Bern, Switzerland

1. Introduction	1
II. Materials and Methods	2
A. Maintenance of Lymphoid and Myeloid Cell Lines and Production	
of Lymphokine- and Monokine-Containing Supernatants	2
B. Assay for Migration Inhibitory Factor (MIF)	4
C. Assay for Mitogenic Factor (MF) on Human PBL	5
D. Assay for Mitogenic Factor on Mouse Thymocytes	5
E. Assay for Lymphocyte Activating Factor (LAF) Activity	6
F. Assay for Colony Stimulating Activity (CSA)	6
G. Assay for Skin Reactive Factor (SRF)	6
H. Assay for Vascular Permeability Factor (VPF)	6
I. Assay for Interferon (IF)	7
J. Assay for Macrophage Cytotoxicity Factor (MCF)	7
K. Assay for Leukocyte Inhibition Factor (LIF)	7
L. Kinetic Study on Lymphokine Production by the B-Cell Line	
RPMI 1788	8
III. Results	8
IV. Discussion	16
V. Summary	17
References	18

I. Introduction

During the past decade, an increasing number of soluble products from stimulated lymphocytes and monocytes referred to as "lymphokines" and "monokines," respectively, have been described (Wolstencroft and Dumonde, 1970; de Weck *et al.*, 1980). Classical approaches in the production of human lymphokines (LK) and

¹ This work has been supported in part by the Swiss National Science Foundation (Grant No. 3.850.077).

² Recipient of Postdoctoral Research Fellowships awarded by the National Cancer Institute, NIH (CA-0628-01) and the Swiss National Fund (88.679.-0.78). Present address: Department of Microbiology and Immunology, University of Michigan Medical Center, Ann Arbor, Michigan 48109.

monokines (MK) have involved the stimulation of peripheral blood lymphoid and myeloid cells, either specifically by antigen or nonspecifically by mitogens (Rocklin et al., 1972; Remold et al., 1972). One of the problems in using such unfractionated cell populations in the production of human LK and MK is a "ping-pong" effect in which stimulated cells foster the indirect production of LK or MK from other cell types (Oppenheim et al., 1979). According to our experience (unpublished observations), this phenomenon leads to large variations in the production and pattern of LK and MK activities in supernatants from various individuals. Using Con A, one finds many activities in a given supernatant of human peripheral blood lymphocyte (PBL) stimulated cells such as migration inhibitory factor (MIF), mitogenic factor (MF), lymphocyte activating factor (LAF), and colonystimulating activity (CSA), and although produced with different kinetics, it is virtually impossible to use mitogen stimulation for production of a single activity (Schook et al., 1978). Second, the amounts of these biologically active products released from stimulated cells are extremely low and their frequently similar chromatographic and electrophoretic behavior creates difficulties during separation procedures (de Weck et al., 1976). Earlier reports (Granger et al., 1970; Papageorgiou et al., 1974; Yoshida et al., 1976) have shown that LK activities were found in supernatants from continuous lymphoid cell lines, which may be a source(s) of large amounts of LK. In an attempt to produce supernatants containing more restricted LK and MK activities, we have systematically assayed supernatants from 22 human lymphoid cell lines (LCL) and 2 myeloid cell lines (MCL) for nine various factors.

II. Materials and Methods

A. Maintenance of Lymphoid and Myeloid Cell Lines and Production of Lymphokine- and Monokine-Containing Supernatants

Twenty-two continuous lymphoid cell lines and two myeloid cell lines maintained at Roswell Park Memorial Institute were used in this study. A description of the individual line characteristics is given in Table I.

Supernatants for determination of LK and MK production were prepared as follows (Fig. 1): Initial stationary starter cultures (0.5×10^6 viable cells/ml) of RPMI 1640 supplemented with 10% heatinactivated fetal calf serum (FCS), penicillin (100 U/ml), and strep-