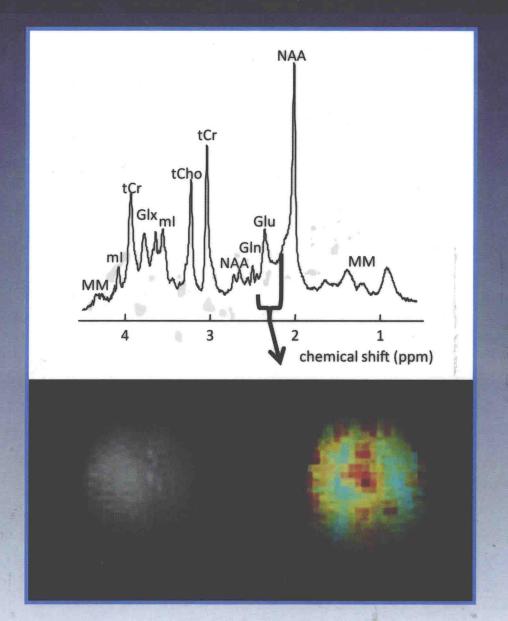
# HAGNETIC HESONANCE SPECTROSCOPY

TOOLS FOR NEUROSCIENCE RESEARCH AND EMERGING CLINICAL APPLICATIONS

EDITED BY CHARLOTTE J. STAGG, DOUGLAS L. ROTHMAN





## MAGNETIC RESONANCE SPECTROSCOPY

Tools for Neuroscience Research and Emerging Clinical Applications

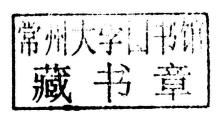
#### Edited by

#### CHARLOTTE J. STAGG

Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford

#### Douglas L. Rothman

Departments of Diagnostic Radiology, Magnetic Resonance Research Center, Yale University School of Medicine







Academic Press is an imprint of Elsevier 32 Jamestown Road, London NW1 7BY, UK 225 Wyman Street, Waltham, MA 02451, USA 525 B Street, Suite 1800, San Diego, CA 92101-4495, USA

Copyright © 2014 Elsevier Inc. All rights reserved

No part of this publication may be reproduced, stored in a retrieval system or transmitted in any form or by any means electronic, mechanical, photocopying, recording or otherwise without the prior written permission of the publisher.

Permissions may be sought directly from Elsevier's Science & Technology Rights Department in Oxford, UK: phone (+44) (0) 1865 843830; fax (+44) (0) 1865 853333; email: permissions@elsevier.com. Alternatively, visit the Science and Technology Books website at www.elsevierdirect.com/rights for further information.

#### Notice

No responsibility is assumed by the publisher for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions or ideas contained in the material herein. Because of rapid advances in the medical sciences, in particular, independent verification of diagnoses and drug dosages should be made.

#### British Library Cataloguing-in-Publication Data

A catalogue record for this book is available from the British Library

#### Library of Congress Cataloging-in-Publication Data

A catalog record for this book is available from the Library of Congress

ISBN: 978-0-12-401688-0

For information on all Academic Press publications visit our website at elsevierdirect.com

Typeset by MPS Limited, Chennai, India www.adi-mps.com

Printed and bound in United States of America

14 15 16 17 18 10 9 8 7 6 5 4 3 2 1



### MAGNETIC RESONANCE SPECTROSCOPY

此为试读,需要完整PDF请访问: www.ertongbook.com

### Acknowledgements

The editors would like to thank April Graham and Mica Haley for expertly steering this book from conception to production with constant good humor and seemingly endless patience. Thanks also go to our many colleagues around the globe who generously gave up so much of their time to contribute such

excellent chapters to the volume. CJS would like to thank Heidi Johansen-Berg, as always, for her generosity and support in allowing this book to be started and Emily Aspden for arriving just late enough to ensure it could be completed.

### Contributors

- **Prasanth Ariyannur** Uniformed Services University of the Health Sciences, Bethesda, MD, USA
- **Peethambaran Arun** Uniformed Services University of the Health Sciences, Bethesda, MD, USA
- Carles Arús Centro de Investigación Biomédica en Red en Bioingeniería, Biomateriales y Nanomedicina and Institut de Biotecnologia i de Biomedicina, Universitat Autònoma de Barcelona, Cerdanyola del Vallès, Spain
- Ian C. Atkinson Center for Magnetic Resonance Research, University of Illinois at Chicago, Chicago, IL, USA
- Velicia Bachtiar Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford, United Kingdom
- **Kevin L. Behar** Yale University School of Medicine, New Haven, CT, USA
- Jonathan G. Best Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford, United Kingdom
- Andrew Bivard University of Melbourne, Melbourne Brain Centre, Melbourne, Australia
- Vincent O. Boer University Medical Center Utrecht, Utrecht, The Netherlands
- Jennifer Brawn Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford, United Kingdom
- Dallas Card Department of Diagnostic Imaging, Hospital for Sick Children, Toronto, Ontario, Canada
- Kim M. Cecil Cincinnati Children's Hospital Medical Center at the University of Cincinnati College of Medicine, Cincinnati, OH, USA
- Olga Ciccarelli University College London Institute of Neurology, London, United Kingdom; National Institute for Health Research University College London Hospitals Biomedical Research Centre, London, United Kingdom
- Henk M. De Feyter Magnetic Resonance Research Center, Yale University School of Medicine, New Haven, CT, USA
- Robin A. de Graaf Magnetic Resonance Research Center, Yale University, School of Medicine, New Haven, CT, USA
- Nicola De Stefano University of Siena, Italy
- Andrea Dennis Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford, United Kingdom
- **Nicholas Gant** Centre for Brain Research, The University of Auckland, New Zealand
- Antonio Giorgio University of Siena, Italy

- Rolf Gruetter University of Geneva, Switzerland; Center for Biomedical Imaging and Laboratory of Functional and Metabolic Imaging, Ecole Polytechnique Federale de Lausanne, Switzerland; University of Lausanne, Switzerland
- **Hoby Hetherington** University of Pittsburgh Medical Center, Pittsburgh, PA, USA
- Amber Michelle Hill University College London Institute of Neurology, London, United Kingdom
- Christoph Juchem Magnetic Research Center, Yale University School of Medicine, New Haven, CT, USA
- Margarida Julià-Sapé Centro de Investigación Biomédica en Red en Bioingeniería, Biomateriales y Nanomedicina and Institut de Biotecnologia i de Biomedicina, Universitat Autònoma de Barcelona, Cerdanyola del Vallès, Spain
- **Dennis W.J. Klomp** University Medical Center Utrecht, Utrecht, The Netherlands
- **Hongxia Lei** University of Geneva, Switzerland; Center for Biomedical Imaging, Ecole Polytechnique Federale de Lausanne, Switzerland
- Joanne C. Lin Centre for Brain Research, The University of Auckland, New Zealand
- Carles Majós L'Hospitalet de Llobregat, Barcelona, Spain; Centro de Investigación Biomédica en Red en Bioingeniería, Biomateriales y Nanomedicina, Cerdanyola del Vallès, Spain
- **Vladimír Mlynárik** Center for Biomedical Imaging, Ecole Polytechnique Federale de Lausanne, Switzerland
- John R. Moffett Uniformed Services University of the Health Sciences, Bethesda, MD, USA
- **Aryan M.A. Namboodiri** Uniformed Services University of the Health Sciences, Bethesda, MD, USA
- Jamie Near McGill University and Centre d'Imagerie du Cerveau, Douglas Mental Health University Institute, Montreal, Quebec, Canada
- Jullie Pan University of Pittsburgh Medical Center, Pittsburgh, PA, USA
- Mark Parsons University of Newcastle, Newcastle, New South Wales, Australia
- **Brian D. Ross** Huntington Medical Research Institutes, Magnetic Resonance Spectroscopy Unit, Pasadena, CA, USA
- **Douglas L. Rothman** Magnetic Research Center, Yale University School of Medicine, New Haven, CT, USA
- Jun Shen Molecular Imaging Branch, National Institute of Mental Health, Bethesda, MD, USA

xii CONTRIBUTORS

Nicola R. Sibson CR-UK/MRC Gray Institute for Radiation Oncology and Biology, University of Oxford, Oxford, United Kingdom

- John G. Sled Research Institute, Hospital for Sick Children, Toronto, Ontario Canada; University of Toronto, Toronto, Ontario Canada
- Charlotte J. Stagg Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, University of Oxford, United Kingdom
- Peter Stanwell University of Newcastle, Newcastle, New South Wales, Australia
- Margot J. Taylor Research Institute, Hospital for Sick Children and Medical Imaging, Toronto, Ontario Canada; University of Toronto, Toronto, Ontario, Canada

- Matthew Taylor Institute of Psychiatry, London, United Kingdom
- Keith R. Thulborn Center for Magnetic Resonance Research, University of Illinois at Chicago, Chicago, IL, USA
- Clare E. Turner Centre for Brain Research, The University of Auckland, New Zealand
- Katy Vincent Oxford Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, and Nuffield Department of Obstetrics and Gynaecology, University of Oxford, United Kingdom
- **Lijing Xin** University of Lausanne, Switzerland; Laboratory of Functional and Metabolic Imaging, Ecole Polytechnique Federale de Lausanne, Switzerland

### Introduction

Magnetic resonance spectroscopy (MRS) is a noninvasive technique that uses the same physics principles and detection methods as magnetic resonance imaging (MRI) of H<sub>2</sub>O, but adds an additional dimension of information by also detecting the resonance frequencies of metabolites. From the resonance frequencies (referred to as chemical shift) and other properties of these resonances the identity, concentration, and stable isotope enrichment of biochemicals can be determined. <sup>1</sup>H MRS, which is the most widely used, was first performed on the brain by Behar and coworkers in the lab of Professor Robert G Shulman in 1983 (Behar et al., 1983). In this pioneering study, performed on a rat in a vertical bore magnet, resonances of N-acetylaspartate (NAA), glutamate, glutamine, choline, creatine, and lactate were assigned. These remain the only major metabolites studied using in vivo<sup>1</sup> H MRS. Within several years after this study the first highfield (1.5 T and above) human magnets were built by Oxford Instruments and in 1985 Bottomley and coworkers at General Electric published the first localized human brain <sup>1</sup>H MRS spectra (Bottomley et al., 1985). The first applications to human disease were presented at the meeting of the Society of Magnetic Resonance in Medicine in 1986 by den Hollander and colleagues working at Philips, and these were soon followed by several groups who performed pioneering studies in stroke, tumors, and other clinical conditions.

Following these and other pioneering studies <sup>1</sup>H MRS has been used for many years in clinical neuroscience as a method for investigating brain neurochemistry, critical to understanding neurological and psychiatric disease. However, a relatively low signal-to-noise (SNR) ratio has limited its use on standard clinical scanners. Over recent years, with the increasing availability of high and ultrahigh field scanners, as well as a much increased understanding of how metabolism plays a critical role in neuroenergetics and neurotransmission, MRS has undergone something of a renaissance and gained traction within the MR community for translational and clinical neuroscience. Improved acquisition and analysis approaches have increased interest in its use both for traditional clinical applications and also for neuroscience research. However, although excellent books exist covering the technical aspects of MRS for physicists, there is currently no book targeted at clinicians and neuroscientists that covers all aspects of the technique. In this book we attempt to address this need. It is organized as a reference text that is aimed not at physicists but at experts in the *application* of MRS such as neurologists, psychiatrists, radiologists, and neuroscientists. However, we hope that the coverage of applications and basic methodologies is complete enough that physicists entering the field may benefit and even experienced MR physicists working in other areas could use it to determine the state-of-the-art methodology used in the field.

To achieve these goals we have divided the book into three sections, which we outline below.

#### SECTION 1: HOW MRS IS ACQUIRED

In this section we have enlisted experts in the field of MRS data acquisition and processing to provide an introduction to the field and an overview of stateof-the-art methodology in these areas. Even though the mathematics is kept at a minimum, enough technical detail is included to allow readers to understand the principles and relative strengths and weaknesses of the different methods. In Chapter 1.1 Drs. Christoph Juchem and Douglas Rothman describe the basis of Magnetic Resonance focusing on basic principles but also give an overview of some of the most common methods used in MRS and chemicals measured. In Chapter 1.2 Drs. Hongxia Lei, Lijing Xin, Rolf Gruetter, and Vladimír Mlynárik describe the state of the art of single-volume <sup>1</sup>H MRS as well as novel recent approaches such as ultra short TE MRS. This chapter describes the methods needed to meet the stringent requirements for volume localization with MRS due to large resonances from water and scalp lipids. The critical importance and optimal methods for improving field homogeneity and suppressing intravoxel water are also described in detail as well as artifacts that may occur if adequate criteria are not met. MRS can both be obtained as information from a single volume (or several) in the brain or as a metabolic image. In Chapter 1.3 Drs. Vincent Boer and Dennis Klomp provide an introduction to magnetic resonance spectroscopic imaging (MRSI) including its application at ultrahigh fields such as 7 T. This chapter demonstrates the great potential of MRSI but also reviews its limitations, many of which relate to the need to establish xiv Introduction

adequate static and radiofrequency magnetic field homogeneity throughout the volume imaged (as compared to single-volume MRS where optimization is only required in a small region of the brain). With optimal B<sub>0</sub> homogeneity and higher B<sub>0</sub> fields more and more metabolites can be distinguished based on their resonance frequencies (chemical shift), but lower concentration metabolites such as  $\gamma$ -amino butyric acid (GABA) still cannot be resolved at clinical 3T fields. To overcome these spectral overlap limitations MRS methods that separate resonances based not just upon resonance frequency but also upon quantum J-coupling between resonances within a single molecule have been developed. These methods are often referred to as "spectral editing" because they edit out resonances from specific chemicals from overlapping resonances from other chemicals. In Chapter 1.4 Dr. Robin de Graaf provides a guide to modern editing and related 2D MRS methods. While these methods have largely been limited to specialized research MR systems recent developments in clinical 3T systems have greatly expanded their applicability. Even with the best data acquisition methodology the analysis and calibration methods used in MRS play a critical role in the accuracy and precision of the results obtained. In Chapter 1.5 Dr. Jamie Near covers in detail methods used to analyze and quantitate the in vivo MRS spectrum as well as the advantages and pitfalls of each.

### SECTION 2: BIOCHEMISTRY—WHAT UNDERLIES THE SIGNAL?

This section covers the biochemistry of the major neurochemicals, and what we can infer from increased or decreased levels of these in the brain and what cannot be elucidated provide a description of modern strategies for interpreting MRS results. In Chapter 2.1 Drs. John Moffett, Prasanth Ariyannur, Peethambaran Arun, and Aryan Namboodiri cover N-acetylaspartate (NAA) and N-acetylaspartylglutamate (NAAG) in central nervous system (CNS) health and disease. NAA was identified in the very first in vivo1 H MRS brain study performed and due to its high concentration and the presence of a singlet methyl group (which increases sensitivity by  $3 \times$  due to proton multiplicity) has been the major biochemical studied in clinical MRS. Despite its wide use there is considerable uncertainty about the function of NAA in the CNS and how to interpret changes seen in the MRS spectrum. In this chapter evidence for our present understanding of the roles of NAA and NAG and their underlying biochemistry are covered in detail along with implications for clinical MRS studies. In Chapter 2.2 Drs. Clare Turner and Nicholas Gant cover creatine, another

major metabolite measured in the MRS spectrum, again due to its high concentration and the presence of a methyl group. Creatine is often used as a concentration reference in the MRS spectrum (as described in Chapter 1.5) so that it is important to understand conditions where its concentration may change. Metabolites that have often been used in clinical MRS studies are the combined resonances of cholinecontaining compounds. In Chapter 2.3 Drs. Nicholas Gant and Joanne Lin cover in detail the biochemistry and functional roles of choline in the brain and how choline levels may reflect pathologies. Changes in choline, creatine, and NAA tend to be relatively slow, with the time for biosynthetic replacement of the pools (turnover time) on the order of days. However, MRS can also look at metabolites that are dynamically turning over through their involvement in energy metabolism and neurotransmission. In Chapter 2.4 Dr Jun Shen describes the role of glutamate in brain energy metabolism and neurotransmission and how these roles can be studied using <sup>1</sup>H MRS and <sup>13</sup>C MRS (which is followed up in more detail in Sections 3 and 4). This chapter also provides additional background on the MRS measurement of glutamate. In Chapter 2.5 Drs. Jonathan Best, Charlotte Stagg, and Andrea Dennis cover the biochemistry and functional roles of myo-inositol, GABA, glutamine, and lactate. GABA and glutamine provide, respectively, a measure of metabolism in GABAergic neurons and glial cells, which along with glutamatergic neurons (they contain the majority of the glutamate signal) account for the large majority of cells in the brain. The GABA signal measured by <sup>1</sup>H MRS is also related to tonic GABAergic inhibition, which opens up <sup>1</sup>H MRS to be applied to a variety of neuroscience-related applications as described in Section 3. Due to its production by nonoxidative glycolysis, lactate levels are highly sensitive to the oxygenation status of brain tissue and, as further described in Section 3, can be diagnostic for necrotic tumors and other conditions such as brain ischemia. Myo-inositol appears to be primarily localized to glial cells and the resonance is highly sensitive to the presence of neurodegenerative disease as well as alterations in brain osmotic levels like those found in ketoacidotic hyperglycemia and hyperammonemia.

### SECTION 3: APPLICATIONS OF PROTON MRS

MRS is theoretically feasible on any nucleus that possesses a magnetic moment; however, by far the most common nucleus for study is the proton. Protons have the greatest gyromagnetic ration of any nuclei seen in vivo and are also by far the most abundant nucleus in

INTRODUCTION XV

the brain. These two factors mean that <sup>1</sup>H MRS has a relatively high SNR and, because protons are found in all metabolically interesting compounds, there is great potential for the study of the brain. In addition, MRI is performed on H<sub>2</sub>O, meaning that clinical MR scanners can be used for <sup>1</sup>H MRS without buying expensive additional hardware, theoretically opening up its use to a wide range of users.

This section discusses the undoubted potential of <sup>1</sup>H MRS for both clinical and neuroscientific applications, as well as raising the limitations of the technique in the context of the conditions in which they have been applied. In Chapter 3.1 Drs. Carles Majós, Margarida Julià-Sapé, and Carles Arús discuss the clinical applications of MRS in tumor detection and management-exploring the application of MRS most commonly seen in clinical practice. MRS can be used to distinguish between tumors and non-tumors, can help the clinician to determine the nature of a tumor before pathology can be acquired, and can monitor the sequelae of treatment by distinguishing between tumor regrowth and post-treatment changes. In Chapter 3.2 Drs. Nicola De Stefano and Antonio Giorgio discuss the potential of MRS to determine pathology and monitor progression in inflammatory conditions, particularly in multiple sclerosis. Although MRS has undoubted potential to provide informative biomarkers for the development of potential treatments for MS, these are currently limited by the difficulty of acquiring reproducible data between different scanners, and possible solutions to this problem are discussed.

Chapter 3.3 focuses on epilepsy, a common neurological condition and one in which, as Drs. Julie Pan and Hoby Hetherington discuss, the ability of MRS to quantify neuroenergetics means that 'H MRS is invaluable in allowing underlying pathologies to be studied. They also describe complementary work using 31P and 13C MRS, nuclei covered in more detail in Section 4, to further assess altered energetics in epileptogenic tissue. In Chapter 3.4 Drs. Andrew Bivard, Peter Stanwell, and Mark Parsons review the potential of MRS to study the metabolic events in the hyperacute phases of stroke recovery and its developing use as a window into the changes underlying the recovery of function in the months that follow. As with all neurological conditions, however, there are significant challenges in acquiring and interpreting <sup>1</sup>H MRS data from these patients, particularly data acquired from within and surrounding the lesioned region where the tissue inhomogeneity leads to greatly broadened linewidths. Some potential solutions are discussed as well as the potential pitfalls associated with interpreting these data.

In Chapter 3.5 Dr. Kim Cecil discusses the unique potential of MRS in the study of pediatric conditions, where the acquisition of diagnostic information noninvasively (e.g., with no injected radioactive tracers) is perhaps of heightened importance. The study of inborn errors of metabolism such as the leukodystrophies and Canavan's disease in particular has shed considerable light on the pathology of these conditions. Chapter 3.6 highlights the potential of <sup>1</sup>H MRS to improve our understanding of psychiatric conditions. Dr. Matthew Taylor focuses particularly on psychotic conditions and mood disorders, where a range of abnormalities in neural metabolism and glutamatergic signaling have been identified. The final clinical MRS application to be considered is that of spinal MRS in Chapter 3.7, where Drs. Amber Hill and Olga Ciccarelli discuss the potential of this technologically challenging approach. Despite the difficulties of acquiring spectra of adequate quality from the cord, given its relatively small size and intrinsic movement, several preclinical and clinical studies have been performed, the results of which suggest that this may well become a much more widely used approach in the future.

Chapter 3.8 moves toward the application of <sup>1</sup>H MRS for neuroscientific questions. Drs. Velicia Bachtiar and Charlotte Stagg provide an overview of the potential of MRS studies focusing on GABA and glutamate in particular to increase our understanding of how differences in behavior between people may be driven by underlying physiology. In Chapter 3.9 Drs. Dallas Card, Margot Taylor, and John Sled discuss the use of MRS to study natural aging, where studies have been targeted in particular both at the rapid development occurring in the brains of infants during development *in utero* and in the elderly. The importance of these data, and the challenges of longitudinal studies, particularly in infants, are discussed.

In Chapter 3.10 Drs. Jennifer Brawn and Katy Vincent discuss the role of MRS in the study of hormonal influences in the brain. Until relatively recently the substantial effects of hormones on brain activity were not recognized, but there is now increasing evidence that hormones, their precursors, and their derivatives all have striking effects on neuronal metabolism and cell signaling. In particular, the role of the menstrual cycle is discussed in some detail, as this may be important to take into account when interpreting the results of MRS studies in other contexts. Finally, in Chapter 3.11, Drs. Nicola Sibson and Kevin Behar discuss the use of <sup>13</sup>C MRS to study brain biochemistry. Although not as widely available as <sup>1</sup>H MRS, <sup>13</sup>C MRS has a unique potential to study brain energetics and metabolism in vivo. 13C MRS has already provided many insights into brain function in animal models and, with the ongoing improvements in the technique, its potential for the study of brain function in humans is beginning to be realized, a topic covered in more detail in Section 4.

### SECTION 4: APPLICATIONS OF NON-PROTON MRS

Although the majority of in vivo MRS studies of the CNS have used the 1H nucleus there is a large amount of complimentary information that can be obtained using other nuclei, including <sup>13</sup>C, sodium (<sup>23</sup>Na), oxygen (<sup>17</sup>O), phosphorus (<sup>31</sup>P), and potassium (<sup>39</sup>K). Considerable insights into brain energetics and function have been obtained in research studies using these nuclei, several of which are described in Sections 2 and 3. Because these nuclei gain in sensitivity with field to a greater degree than the <sup>1</sup>H nucleus they may become standard as ultrahigh field systems such as 7 T become more common. In this section we cover present and potential uses of these nuclei and how they can add information to both clinical diagnosis and basic understanding of brain metabolism and function. In Chapter 4.1 Drs. Keith Thulborn and Ian Atkinson provide an introduction to state-of-the-art sodium, oxygen, and phosphorous MRS, MRSI, and MRI and show how at ultrahigh fields even imaging of potassium is possible. MRI of sodium and potassium has great potential for detecting clinical imbalances that may profoundly impact brain function as well as even a more direct form of functional imaging. Further, the authors introduce the concept of bioscales in the clinical applications of these measurements making a strong argument for going beyond standard MRI and <sup>1</sup>H MRS. In Chapter 4.2 Drs. Henk De Feyter and Douglas Rothman cover the methodology and applications of <sup>13</sup>C MRS in combination with <sup>13</sup>C-labeled brain substrates such as glucose and acetate. While this is one of the most challenging areas of MRS due to the need for stable isotope infusion, metabolic modeling and modified MR hardware, it has already provided novel insight into brain function and disease and has shown high sensitivity to a variety of clinical conditions including Alzheimer's disease and healthy aging, cancer, developmental disorders, diabetes, depression, and stroke. It is also the only method that can be used

to study cell type-specific metabolism and glutamate and GABA neurotransmission in humans.

An additional limitation of using <sup>13</sup>C MRS is its low sensitivity relative to <sup>1</sup>H MRS, which results in relatively course spatial resolution. While this can be recovered using inverse <sup>1</sup>H-[<sup>13</sup>C] MRS, particularly at high fields, the overall achievable spatial resolution is still well below PET scanning and other metabolic imaging methods. However, this limitation has been overcome recently through the development of hyperpolarized 13C MRS. The MRS signal is proportional to the difference in the number of nuclear spins pointing parallel versus antiparallel to the main magnetic field. Normally the excess is a small fraction of the total nuclei, but in hyperpolarized <sup>13</sup>C MRS the sensitivity of detection (and in principle spatial resolution) can be improved by over 10,000-fold due to prepolarization of the 13C-labeled precursor prior to injection. In Chapter 4.3 Dr. Brian Ross describes the state of the art of the use of hyperpolarized 13C MRS to study the brain. This application has been very challenging due to both the difficulties involved in performing conventional MRS and the additional challenges of delivering the hyperpolarized compound to the brain before it reverts through relaxation back to normal levels of polarization (losing the enhancement). However, recent breakthroughs described in this chapter make the prospects of performing these scans in patients much more promising.

#### References

Behar, K. L., den Hollander, J. A., Stromski, M. E., Ogino, T., Shulman, R. G., Petroff, O. A., & Prichard, J. W. (1983). High resolution <sup>1</sup>H nuclear magnetic resonance study of cerebral hypoxia in vivo. Proceedings of the National Academy of Sciences USA, 80(16), 4945–4948.

Bottomley, P. A., Edelstein, W. A., Foster, T. H., & Adams, W. A. (1985). In vivo solvent suppressed localized hydrogen nuclear magnetic resonance spectroscopy: a window to metabolism? Proceedings of the National Academy of Sciences USA, 82, 2148–2152.

### Contents

Acknowledgements ix Contributors xi Introduction xiii

1

#### TECHNICAL ASPECTS—HOW MRS IS ACQUIRED

1.1. Basis of Magnetic Resonance 3 Christoph Juchem and Douglas L. Rothman

Introduction 3 MRS Methods 4 Conclusions 12 References 12

> 1.2. Localized Single-Voxel Magnetic Resonance Spectroscopy, Water Suppression, and Novel Approaches for Ultrashort Echo-Time Measurements 15

> > HONGXIA LEI, LIJING XIN, ROLF GRUETTER AND VLADIMÍR MLYNÁRIK

Introduction 15
Instrumental Impacts on Volume Definition 16
Factors Affecting Spectral Quality 18
Basic Localization <sup>1</sup>H Mrs Methods 25
Acknowledgments 30
References 30

1.3. Technical Considerations for Multivoxel Approaches and Magnetic Resonance Spectroscopic Imaging 31

VINCENT O. BOER AND DENNIS W.J. KLOMP

Introduction 31
Multivolume Selection 31
Spatial Encoding 32
Fast Gradient-Encoding Methods 33
Encoding Based on Prior Knowledge 34
Water Suppression 35
Lipid Suppression 35
B<sub>0</sub> Shimming 37
Conclusions 38
References 38

1.4. Spectral Editing and 2D NMR 40 ROBIN A. DE GRAAF

Introduction 40 Scalar Coupling 40 In Vivo GABA Editing 44 2D NMR Spectroscopy 46 References 47

1.5. Spectral Quantification and Pitfalls in Interpreting Magnetic Resonance Spectroscopic Data: What To Look Out For 49

Introduction: A Simple Example of Spectral Quantitation 49
Measuring Peak Intensity 51
Nuisance Signals 56
Software Packages for Spectral Quantification 58
Signal Referencing and Absolute Quantification 61
Quality Control 63
Conclusions 65
References 66

2

### BIOCHEMISTRY – WHAT UNDERLIES THE SIGNAL?

2.1. N-Acetylaspartate and N-Acetylaspartylglutamate in Central Nervous System Health and Disease 71
IOHN B. MOFFETT, PRASANTH ARIYANNUR.

PEETHAMBARAN ARUN AND ARYAN M.A. NAMBOODIRI

Introduction 71 NAA 71 NAAG 79 References 85

2.2. The Biochemistry of Creatine 91 CLARE E. TURNER AND NICHOLAS GANT

Introduction 91
Creatine and High-Energy Phosphate Metabolism 91
CK within the CNS 94
Therapeutic Cr Supplementation 94
Quantification of Brain Cr with MRS 97
Conclusions 98
References 98

### 2.3. The Biochemistry of Choline 104 JOANNE C. LIN AND NICHOLAS GANT

Introduction 104
Biosynthesis 104
Biological Function 106
Indicators of Membrane Damage 106
The Cho Peak in MRS 106
Conclusions 107
References 108

### 2.4. Glutamate 111 JUN SHEN

Introduction 111
Roles of Glutamate in Brain 111
Regulation of Glutamate Concentration 116
Interpretation of Changes in Glutamate Concentration 117
Conclusions 119
References 120

### 2.5. Other Significant Metabolites: Myo-Inositol, GABA, Glutamine, and Lactate 122

JONATHAN G. BEST, CHARLOTTE J. STAGG AND ANDREA DENNIS

Introduction 122 Myo-Inositol 122 GABA 126 Glutamine 128 Lactate 129 Conclusions 134 References 134

3

#### APPLICATIONS OF PROTON-MRS

#### 3.1. Usefulness of Proton Magnetic Resonance Spectroscopy in the Clinical Management of Brain Tumors 141

CARLES MAJÓS, MARGARIDA JULIÀ-SAPÉ AND CARLES ARÚS

Introduction 141

<sup>1</sup>H MRS Acquisition and Normal Values 142

To be or not to be a Tumor, that is the Question 145

Proton MRS in the Classification of Brain Tumors 148

MRS in the Assessment of Glial Tumor Grade 157

Proton MRS in the Follow-up of Brain Tumors 158

Acknowledgments 159

References 159

### 3.2. Multiple Sclerosis and Inflammatory Diseases 162

NICOLA DE STEFANO AND ANTONIO GIORGIO

Introduction 162 MS 162 Conclusions 168 References 168

### 3.3. Epilepsy 172 JULLIE PAN AND HOBY HETHERINGTON

Introduction 172

31P Studies of High-Energy Phosphates in Epilepsy 172

1H Spectroscopy in Epilepsy 174

Conclusions 180

References 180

### 3.4. Stroke and Cerebral Ischemia 183 Andrew bivard, peter Stanwell and Mark Parsons

Introduction 183
Ischemic Stroke 183
Hemorrhagic Stroke 184
Magnetic Resonance Imaging in Stroke 184
Spectroscopy: Proton Mrs 186
MRS in the Penumbra and Infarct Core 189
Spectral Editing 189
Diaschisis 189
Stroke and Depression 191
Spectroscopy in the Recovering Brain 191
Problems with MRS Acquisition in Stroke 192
References 193

#### 3.5. Use of MRS in Inborn Errors of Metabolism: Canavan's Disease and MRS in Differential Diagnosis 196 KIM M. CECIL

Introduction 196
Primary Leukodystrophies 198
Lysosomal Storage Diseases 202
Peroxisomal Disorders Producing
Leukodystrophies 207
Amino Aciduria 209
Organic Acidurias 211
Conclusions 218
References 218

### 3.6. MRS of Psychiatric Disorders 222 MATTHEW TAYLOR

Introduction 222
Findings During Episodes of Illness 222
Abnormalities after Clinical Recovery 224
Abnormalities in High-Risk Groups 224
Clinical Role 226
Conclusions 226
References 226

### Preclinical and Clinical Applications of <sup>1</sup>H MRS in the Spinal Cord 229

AMBER MICHELLE HILL AND OLGA CICCARELLI

Introduction 229
Importance of <sup>1</sup>H MRS Advances 229
Methodological Challenges and Considerations 230

vii

Preclinical and Clinical <sup>1</sup>H MRS Applications in the Spinal Cord 233 Future Potential of <sup>1</sup>H MRS in the Spinal Cord 240 Conclusions 240 References 241

#### Interindividual Differences in Behavior and Plasticity 243

VELICIA BACHTIAR AND CHARLOTTE J. STAGG

Introduction 243
Summary of GABA Metabolism 243
Data from Animal Models of Plasticity Induction 244
Quantifying GABA in Humans in Vivo 244
Chapter Outline 246
Interindividual Differences in GABA can be Related to Behavior 246
GABA Changes in Clinical Populations 247
GABAergic Changes in Plasticity Induction 248
Relationship to MRS-Assessed GABA and Information Derived from Other Imaging Modalities 250
Conclusions and Outstanding Questions 251
References 252

#### 3.9. MRS in Development and Across the Life Span 254

DALLAS CARD, MARGOT J. TAYLOR AND JOHN G. SLED

Introduction 254
Overview 254
MRS in Early Brain Development 256
MRS Across the Life Span 260
The Way Forward 262
References 262

#### 3.10. Hormonal Influences on Magnetic Resonance Spectroscopy Measures 266 JENNIFER BRAWN AND KATY VINCENT

Introduction 266
Overview of Hormone Biology 266
Neurochemicals: Males Versus Females 268
Summary of Estradiol and Progesterone 271
Male Sex Steroidal Hormones and Neurochemical Changes 271
Conclusions 271
References 272

#### 3.11. Magnetic Resonance Spectroscopy in Neuroenergetics and Neurotransmission 274 NICOLA R. SIBSON AND KEVIN L. BEHAR

<sup>13</sup>C MRS Measurements of Cerebral Energy Metabolism MRS Measurements of Neurotransmitter Fluxes 280
 Application of <sup>13</sup>C MRS in Human Disease 285
 Summary 285
 References 285

4

### APPLICATIONS OF NON-PROTON MRS

4.1. Quantitative Metabolic Magnetic Resonance Imaging of Sodium, Oxygen, Phosphorus and Potassium in the Human Brain: A Rationale for Bioscales in Clinical Applications 291

KEITH R. THULBORN AND IAN C. ATKINSON

Introduction 291
Justification for Quantification of Mr Signals 292
Quantification of Mr Signals 293
Clinical Applications of Quantitative Sodium
Imaging 299
Potential Applications of Quantitative 17-Oxygen
Mr Imaging 308
Potential Applications of Quantitative 31-Phosphorus
Mr Imaging 308
Applications of Quantitative Potassium
Mr Imaging 308
Conclusions 309
Acknowledgments 310
References 310

#### 4.2. Carbon (<sup>13</sup>C) MRS 312 DOUGLAS L. ROTHMAN AND HENK M. DE FEYTER

Introduction 312
Studies in Animal and Cell Models of the Glutamate/Glutamine
Cycle and Neuronal and Glial Energetics 312
In Vivo<sup>13</sup>C MRS Studies of Human Brain 318
Future Prospects for <sup>13</sup>C MRS Studies
in Humans 323
Summary and Conclusions 326
References 326

## 4.3. Hyperpolarized Magnetic Resonance Imaging and Spectroscopy of the Brain 331 BRIAN D. ROSS

Introduction 331
Kinetic Analysis to Derive In Vivo Metabolic Rate(s) from
Hyperpolarized Mr Studies 344
Some Pointers to Future Exploitation of Hyperpolarized
Mr in Neuroscience and Neurology 346
Acknowledgments 348
References 348

#### Index 351