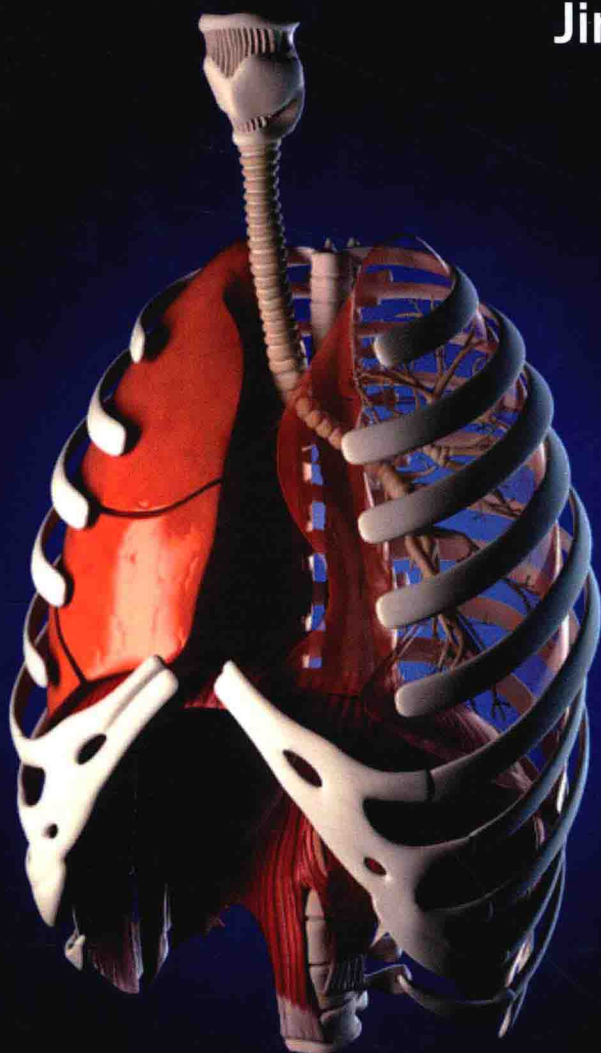


# **Pulmonary Hypertension**

Clinical Research and Challenges

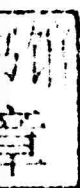
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**Jim Foster**



# **Pulmonary Hypertension – Clinical Research and Challenges**

**Edited by Jim Foster**



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## **Pulmonary Hypertension – Clinical Research and Challenges**

Edited by Jim Foster

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## Preface

The purpose of the book is to provide a glimpse into the dynamics and to present opinions and studies of some of the scientists engaged in the development of new ideas in the field from very different standpoints. This book will prove useful to students and researchers owing to its high content quality.

The book presents fundamental knowledge and current discoveries associated with the structure and cellular function of the pulmonary vasculature. This book elucidates the following topics: dysregulated cellular pathways observed in experimental and human pulmonary hypertension; structure and function of the normal pulmonary vasculature; introduction of numerous particular forms of this illness; clinical characteristics of the illness in general, and its management in special circumstances. It is a unique book which integrates cardiac and pulmonary physiology and pathophysiology with clinical aspects of this disease. Description of dysregulated pathways affected by pulmonary hypertension has also been provided. The book elucidates the effects of hypoxia on the pulmonary vasculature and the myocardium as well. An introduction of the techniques of assessing pulmonary hypertension has also been presented in this book. Numerous forms of pulmonary hypertension have been elucidated in this book, which are particularly challenging in clinical practice (like pulmonary arterial hypertension related to systemic sclerosis). Towards the end, the book discusses special considerations related to the management of this disease in certain clinical scenarios like pulmonary hypertension in the seriously ill.

At the end, I would like to appreciate all the efforts made by the authors in completing their chapters professionally. I express my deepest gratitude to all of them for contributing to this book by sharing their valuable works. A special thanks to my family and friends for their constant support in this journey.

**Editor**

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## **Permissions**

## **List of Contributors**

## **Part 1**

# **Pulmonary Vascular Function and Dysfunction**





# Pulmonary Hypertension: Endothelial Cell Function

Rajamma Mathew

*Dept of Pediatrics, Maria Fareri Children's Hospital at Westchester Medical Center,  
New York Medical College, Dept. of Physiology, New York Medical College, Valhalla, NY,  
USA*

## 1. Introduction

Pulmonary hypertension (PH) is a devastating sequel of a number of diverse systemic diseases including cardiopulmonary, autoimmune, inflammatory and myeloproliferative diseases, drug toxicity, acquired immunodeficiency syndrome, portal hypertension, sickle cell disease and thalassemia etc. Despite major advances in the field, precise mechanism/s of PH is not yet fully understood. In experimental models, endothelial dysfunction is reported to occur before the onset of PH. Therefore, it is not surprising that the clinical diagnosis is often made late during the course of the disease. The major features of PH are impaired vascular relaxation, smooth muscle cell hypertrophy and proliferation, narrowing of the lumen, elevated pulmonary artery pressure and right ventricular hypertrophy. As the disease progresses, neointima formation takes place leading to further narrowing of the lumen, worsening of the disease, right heart failure and death.

Endothelial cells (EC) maintain a balance between vasoconstriction and vasodilatation, and between cell proliferation and apoptosis. In addition, they provide barrier function, balance pro- and anticoagulation factors of the vessel wall, and participate in immune function. Plasmalemmal membrane of the EC have specialized microdomains such as caveolae, rich in cholesterol and sphingolipids that serve as a platform for a numerous signaling molecules and compartmentalize them for optimum function. Caveolin-1, a major protein constituent of caveolae maintains the shape of caveolae and interacts with numerous signaling molecules that reside in or recruited to caveolae, and stabilizes them and keeps these molecules in an inhibitory conformation. A large number of signaling pathways implicated in PH have been shown to interact with endothelial caveolin-1. Therefore, endothelial dysfunction including the loss of functional endothelial caveolin-1 induced by injury such as inflammation, toxicity, increased shear stress and hypoxia may be the initiating factor in the pathogenesis of PH and also contributing to the progression of the disease.

## 2. Pulmonary Hypertension

PH is a rare but a devastating disease with high mortality and morbidity rate. A large number of unrelated diseases are known to lead to PH. The current W.H.O. clinical classification of PH includes 5 groups: *Gr I*: Pulmonary arterial hypertension (PAH): This group comprises of idiopathic and heritable PAH, PAH secondary to drug toxicity and

associated with congenital heart defects, connective tissue diseases, portal hypertension, infection, chronic hemolytic anemia, and persistent pulmonary hypertension of the newborn. Recently, pulmonary veno-occlusive disease and pulmonary capillary hemangiomatosis have been added to this group as a subcategory. *Gr II*: PH due to left heart diseases, *Gr III*: PH due to lung diseases and hypoxia, *Gr IV*: Chronic thromboembolic PH, and *Gr V*: PH secondary to other systemic diseases such as sarcoidosis, myeloproliferative diseases, metabolic disorders and chronic renal failure on dialysis etc. (Simonneau 2004, Hoeper 2009). Regardless of the underlying disease; the major features of PH are endothelial dysfunction, impaired vascular relaxation, smooth muscle cell proliferation and impaired apoptosis, neointima formation, narrowing of the lumen, elevated pulmonary artery pressure and right ventricular hypertrophy, subsequently leading to right heart failure and death. Early changes that occur in the vasculature are not clinically apparent. The patients usually present with vague symptoms, therefore it is not surprising that the diagnosis is often made late. By the time the diagnosis is made, extensive vascular changes have already taken place, which makes the treatment a formidable challenge.

Although major advances have been made, the precise mechanism/s leading to PH is not yet fully elucidated. Multiple signaling pathways have been implicated in the pathogenesis of PH. Loss of nitric oxide (NO), prostacyclin (PGI<sub>2</sub>) and resulting impaired vascular relaxation is the hallmark of PH. Recent studies have revealed that certain genetic defects in humans increase the likelihood of developing PAH. Several members of transforming growth factor (TGF)  $\beta$  superfamily have been implicated in the pathogenesis of PAH; the most notable example being heterozygous germline mutations in bone morphogenic protein receptor type II (BMPRII). This mutation has been noted in approximately 70% of heritable PAH and 26% of idiopathic PAH. Importantly, only 20% of people with this mutation develop PAH. It has recently been shown that inflammation and serotonin increase susceptibility to develop PH in BMPRII<sup>+/–</sup> mice (Thomson 2000, Machado 2006, Long 2006, Song 2008, Mathew 2011b). Altered metabolism of estrogen resulting in low production of 2 methylestradiol is also thought to be a “second hit” for the development of PAH in females with BMPRII mutation (Austin 2009). Thus, environmental, metabolic and/or other genetic factors act as a “second hit” in the development of PAH in patients with BMPRII mutations.

Inflammation plays a significant role in the pathogenesis of clinical and experimental PH. PH has been reported in patients suffering from systemic inflammatory, autoimmune diseases and human immunodeficiency virus infection (Lespirat 1998, Dorfmueller 2003, Mathew 2010). In patients with idiopathic PAH, increased plasma levels of proinflammatory cytokines and chemokines such as interleukin (IL)-1, IL-6, fractalkine and monocyte chemoattractant protein-1 (MCP-1, currently known as CCL2) have been documented. Perivascular inflammatory cells, chiefly macrophages and monocytes, and regulated upon activation normal T-cell expressed and secreted (RANTES) have also been reported in the lungs of these patients [Tuder 1994, Humbert 1995, Dorfmueller 2002, Balabanian 2002, Itoh 2006, Sanchez 2007, Mathew 2010]. In the monocrotaline (MCT) model, early and progressive upregulation of IL-6 mRNA with increased IL-6 bioactivity, progressive loss of endothelial caveolin-1 coupled with activation (tyrosine phosphorylation, PY) of signal transducer and the activator of transcription (STAT) 3 have been shown to occur before the onset of PH; and the rescue of endothelial caveolin-1 inhibits PY-STAT3 activation and attenuates PH (Mathew 2007, Huang 2008). These observations not only underscore a role for inflammation in the pathogenesis of PH but also show the importance of endothelial cell membrane integrity in vascular health.

BMPRII is predominantly expressed in endothelial cells (EC). A part of BMPRII has been shown to colocalize with caveolin-1 in caveolar microdomain and also in golgi bodies. BMPRII signaling is essential for BMP-mediated regulation of vascular smooth muscle cell (SMC) growth and differentiation, and it also protects EC from apoptosis (Yu 2008, Teichert-Kuliszewska 2006). In some cell systems, persistent activation of PY-STAT3 leads to a reduction in the BMPRII protein expression, and BMP2 induces apoptosis by inhibiting PY-STAT3 activation and by down-regulating Bcl-xL, a downstream mediator of PY-STAT3 (Brock 2009, Kawamura 2000). In addition, the loss of BMPRII in in-vivo and in-vitro studies has been shown to increase the production of cytokines such as IL-6, MCP-1 and TGF $\beta$ ; and exogenous BMP ligand decreases these cytokines. Interestingly, reduction in the expression of BMPRII has been reported in patients with idiopathic PAH without BMPRII mutation and to a lesser extent in patients with secondary PH (Atkinson 2002, Mathew 2010). Furthermore, both MCT and hypoxia models of PH exhibit reduction in the expression of BMPRII (Murakami 2010, Reynolds 2009). Since there is a significant interaction and crosstalk between the BMP system and IL-6/STAT3 pathway, a reduction in the expression of BMPRII may exacerbate inflammatory response in PH.

### 3. Endothelial cell function

Endothelium, a monolayer lining the cardiovascular system, is a critical interface between circulating blood on one side, and tissues and organs on the other. EC form a non-thrombogenic and a selective barrier to circulating macromolecules and other elements. Vascular EC subjected to blood flow-induced shear stress transform mechanical stimuli into biological signaling. EC are a group of heterogeneous cells adapted to function for the underlying organs. They have numerous metabolic functions. Depending on the stimuli they are capable of secreting several transducing molecules for participation in vascular tone and structure, inflammation, thrombosis, barrier function, cell proliferation and apoptosis. The dominance of these various factors, determines whether the effect would be cytoprotective or cytotoxic. EC have specialized microdomains on the plasmalemmal membrane. Caveolae, a subset of these specialized microdomains are omega shaped invaginations (50-100 nm) found on a variety of cells including EC, SMC and epithelial cells. Caveolae serve as a platform and compartmentalize a number of signaling molecules that reside in or are recruited to caveolae. Caveolae are also involved in transcytosis, endocytosis and potocytosis. Three isoforms of caveolin proteins have been identified. Caveolin-1 (22kD) is the major scaffolding protein that supports and maintains the structure of caveolae. It interacts with numerous transducing molecules that reside in or are recruited to caveolae, and it regulates cell proliferation, differentiation and apoptosis via a number of diverse signaling pathways. Caveolin-2 requires caveolin-1 for its membrane localization and functions as an anti-proliferative molecule. However, unlike caveolin-1, caveolin-2 has no effect on vascular tone. Caveolin-3 is a muscle specific protein found predominantly in cardiac and skeletal muscle (Razani 2002, Mathew 2011b).

Caveolin-1 interacts, regulates and stabilizes several proteins including Src family of kinases, G-proteins ( $\alpha$  subunits), G protein-coupled receptors, H-Ras, PKC, eNOS, integrins and growth factor receptors such as VEGF-R, EGF-R. Caveolin-1 exerts negative regulation of the target protein within caveolae, through caveolin-1-scaffolding domain (CSD, residue 82-101). Major ion channels such as Ca<sup>2+</sup>-dependent potassium channels and voltage-dependent K<sup>+</sup> channels (Kv1.5), and a number of molecules responsible for Ca<sup>2+</sup> handling such as inositol triphosphate receptor (IP<sub>3</sub>R), heterodimeric GTP binding protein, Ca<sup>2+</sup>



ATPase and several transient receptor potential channels localize in caveolae, and interact with caveolin-1. Production of vasodilators such as nitric oxide (NO), prostacyclin (PGI<sub>2</sub>) and endothelium-derived hyperpolarizing factor [EDHF] within caveolae are dependent on caveolin-1-mediated regulation of Ca<sup>2+</sup> entry (Mathew 2011b).

EC have important cytoplasmic organelles such as Weibel Palade bodies, initially formed in trans-golgi network; as these organelles mature they become responsive to secretagogues such as thrombin and histamine. Weibel Palade bodies store a number of molecules that are necessary for hemostasis, inflammation, vascular proliferation and angiogenesis. These molecules including vWF, P-selectin, angiopoietin 2, ET-1 and endothelin converting enzyme, IL-8, calcitonin gene-related peptide and osteoprotegerin are readily available for the designated function (Metcalf 2008).

### **3.1 Vasomotor tone**

#### **3.1.1 Endothelial nitric oxide synthase (eNOS)/cyclic guanosine monophosphate (cGMP) pathway**

eNOS/cGMP pathway plays a major role in vascular tone and structure. In addition to vasodilatory function, it inhibits cell proliferation, DNA synthesis, platelet aggregation, and it modulates inflammatory responses. eNOS is tightly regulated by a variety of intracellular processes, post-translational modification and protein-protein interaction with caveolin-1 and Ca<sup>2+</sup>/calmodulin. For efficient synthesis, eNOS is associated with golgi bodies, and for optimum activation, eNOS is targeted to caveolae. An increase in intracellular Ca<sup>2+</sup> induced by shear stress and varying oxygen tension activate eNOS (Sessa 1995, Shaul 1996). NO, a short lived free radical gas is synthesized by the catalytic activity of eNOS on L-arginine in the vascular EC. NO activates the enzyme, soluble guanylate cyclase (sGC) that converts guanosine triphosphate (GTP) to cGMP.

cGMP through its protein kinase (PKG) causes vascular relaxation, inhibits cell proliferation and inflammation. It is thought that the extracellular L-arginine and its transport through cationic amino acid transporter-1 (CAT-1), localized in the caveolae, are available for eNOS activity. L-arginine found in different intracellular compartments may not be readily available for eNOS activity. This dependence on extracellular L-arginine for NO production has been termed "L-arginine paradox" (McDonald 1997, Zharikov 1998). In addition to CAT-1, tetrahydrobiopterin (BH<sub>4</sub>) and sGC are compartmentalized in caveolae with eNOS for optimum activation. BH<sub>4</sub> is an essential cofactor required for the activity of eNOS and is synthesized from GTP by a rate limiting enzyme, guanosine triphosphate cyclohydrolase 1 (GTPCH-1). Interestingly, GTPCH-1 also localizes in caveolar microdomain with caveolin-1 and eNOS. This spatial colocalization with eNOS may ensure NO synthesis (Peterson 2009). Caveolin-1 inhibits eNOS through protein-protein interaction, but it also facilitates the increase in intracellular Ca<sup>2+</sup>. HSP90 binds to eNOS away from caveolin-1 in Ca<sup>2+</sup>-calmodulin-depended manner and reduces the inhibitory influence of caveolin-1 to increase eNOS activity. Thus, caveolin-1 and eNOS have a dynamic interrelationship (Gratton 2000, Mathew 2007).

#### **3.1.2 Prostacyclin (PGI<sub>2</sub>)/cyclic adenosine monophosphate (cAMP) pathway**

PGI<sub>2</sub>, a potent vasodilator produced by EC is formed from arachidonic acid by the enzymatic activity of PGI<sub>2</sub> synthase, catalyzed by cyclooxygenase 2. PGI<sub>2</sub> synthase belongs

to a family of G-protein coupled receptors and it colocalizes with endothelial caveolin-1. PGI<sub>2</sub> binds to the receptor resulting in the stimulation of adenylyl cyclase which catalyzes the conversion of ATP to second messenger cAMP. In vascular system, PGI<sub>2</sub> via cAMP and cAMP-dependent protein kinase (PKA) promotes vascular relaxation, inhibits platelet aggregation, inflammation and cell proliferation. In addition, cAMP/PKA pathway activates NO production via phosphorylation of eNOS (Stitham 2011, Kawabe 2010, Zhang 2006). Unlike eNOS, PGI<sub>2</sub> synthase remains enzymatically active even when bound to caveolin-1. Furthermore, eNOS, PGI<sub>2</sub> synthase and vascular endothelial growth factor receptor (VEGFR) 2 colocalize with caveolin-1 suggesting a role for caveolin-1 in angiogenesis signaling pathways (Spisni 2001).

### 3.1.3 Endothelium-derived hyperpolarizing factor (EDHF)

An elevation of intracellular Ca<sup>2+</sup> is essential for EDHF-mediated responses; and the family of transient receptor potential cation (TRPC) channels participates in Ca<sup>2+</sup> entry. TRPC1 is associated with caveolae and a direct interaction with caveolin-1 is necessary for TRPC membrane localization, and Ca<sup>2+</sup> influx. Ca<sup>2+</sup> influx also occurs via TRPV4 channel that belongs to a subfamily of TRPC. TRPV4 channel is expressed in a variety of cells including EC, and is also linked to caveolin-1. Interestingly, arachidonic acid metabolites epoxyeicosatrienoic acids (5, 6-EET and 8, 9-EET) act as direct TRPV4 channel activators in EC. Furthermore, genetic deletion of caveolin-1 has been shown to abrogate EDHF-induced hyperpolarization by altering Ca<sup>2+</sup> entry, thus highlighting the role of caveolin-1 in EDHF regulation (Rath 2009, Vriens 2005, Saliez 2008).

### 3.2 Barrier function

Endothelial cytoskeleton maintains barrier integrity, and EC are linked with each other through tight junctions (TJ) and adherens junctions (AJ). EC control the passage of blood constituents to the underlying tissue. The solutes pass through transcellular or paracellular pathway. Transcellular permeability is regulated by signaling pathways responsible for endocytosis and vesicular trafficking. Paracellular permeability is the result of opening and closing of the endothelial cellular junction; it is governed by a complex arrangement of adhesion proteins and related cytoskeleton proteins organized in distinct structures such as TJ and AJ. Vascular endothelial (VE)-cadherin plays a critical role in integrating spatial signals into cell behavior. VE-cadherin interacts with  $\beta$ -catenin, p120 and plakoglobin, and binds to  $\alpha$ -catenin. Association of VE-cadherin with catenins is required for cellular control of endothelial permeability and junction stabilization. It is believed that the tyrosine phosphorylation of VE-cadherin and other components of AJ results in a weak junction and impaired barrier function (Dejana 2008, Mahta 2006). Furthermore, VE-cadherin is a link between AJ and TJ; it upregulates the gene encoding for the protein claudin-5, a TJ adhesive protein (Taddei 2008). RhoA is considered crucial for the endothelial contractile machinery. Basal activity of RhoA maintains EC junctions, but the induced activity mediates cell contraction, AJ destabilization, barrier disruption and increased permeability. Suppression of RhoA by the activation of p190RhoGAP (GTPase activating protein) reverses permeability. Interestingly, caveolin-1 deficiency impairs AJ integrity and reduces the expression of VE-cadherin and  $\beta$ -catenin. In caveolin-1 deficient EC, increased activity of eNOS accompanied by reactive oxygen species (ROS) generation leads to nitration; the

consequent inactivation of p190RhoGAP-A results in RhoA activation and increased permeability. Inhibition of RhoA or eNOS reduces hyper-permeability in caveolin-1<sup>-/-</sup> mice (van Nieuw Amerongen 2007, Siddiqui 2011, Schubert 2002). It has also been shown that NO-mediated s-nitrosylation of  $\beta$ -catenin is involved in the VEGF-induced permeability. Interestingly, blocking sGC improves high tidal volume ventilator-induced endothelial barrier function. These mice with ventilator-induced lung injury exhibit high cGMP and low cAMP levels, and treatment with iloprost improves vascular leak (Thibeuau 2010, Schmidt 2008, Birukova 2010). Thus, cGMP and cAMP levels appear to have opposing effects on endothelial barrier function.

Activated protein C (APC), a plasma serine protease that forms a complex with EC protein C receptor (EPCR) is a cytoprotective agent functioning as an anticoagulant and profibrinolytic factor, and it participates in anti-inflammatory responses. In addition, EPCR has been shown to support APC-induced protease-activated receptor (PAR)-1-mediated cell signaling. APC via EPCR inhibits RhoA activation, increases Rac1 expression and inhibits vascular permeability. In support of this view, recent studies have shown reduced expression of EPCR and reciprocal increase in the expression of Rho associated kinase (ROCK)1 in a mouse model of ventilation-induced lung injury; and the treatment with APC restored the EPCR expression, attenuated ROCK1 expression and inhibited capillary leak (Baes 2007, Sen 2011, Finigan 2009). Interestingly, both thrombin and APC activate PAR1 with opposing effects. APC-induced PAR1 is cytoprotective whereas thrombin-induced PAR1 activation stimulates RhoA/ROCK, actin stress fiber formation, and alters the integrity of EC layer. Localization of APC-activated PAR1 and EPCR in caveolae is essential for the cytoprotective effects, but for thrombin-activated PAR1 caveolar localization is not necessary. APC treatment inhibits thrombin-induced activation of ERK1/2, whereas in caveolin-1-deficient EC, APC treatment does not prevent thrombin-induced ERK1/2 activation (Russo 2009, Carlisle-Klusack 2007). These studies underscore the importance of EC including endothelial caveolin-1 in maintaining vascular health.

### 3.3 Inflammation

It is well established that inflammation plays a significant role in the pathogenesis of PH. Inflammation is an orchestrated process designed to combat injury/infection. The relevance of endothelium in controlling and modulating inflammatory responses in general is accepted. Under normal conditions, the apoptosis rate in EC is extremely low. Activated EC exhibit a reduction in the endothelial surface layer, glycocalyx, and increased rate of apoptosis. EC detached from the basement membrane appear in blood circulation. Therefore, it is not surprising that increased circulating endothelial cell levels in PH are indicative of poor prognosis (Grange 2010, Jones 2005, Smadja 2010). Both NO and ROS are implicated in the EC response to inflammation. Increased NO levels compared to ROS results in anti-inflammatory response via cGMP pathway, whereas, increased levels of ROS and/or the presence of reactive NO species activate proinflammatory transcription factors (Grange 2010).

In response to infection and inflammatory mediators, EC secrete increased amounts of Interleukin (IL)-6, and upregulate intracellular adhesion molecule (ICAM) and vascular adhesion molecule (VCAM), which spread over the surface of EC. ICAM, VCAM and also P-selectin released from Weibel Palade bodies allow rapid rolling and adhesion of leukocytes



on the EC surface; and biosynthesized E-selectin maintains this process. Interaction of leukocyte platelet endothelial cell adhesion molecule-1 (PECAM-1) and EC PECAM-1 leads to transmigration of leukocytes through the inter EC junction and possibly through EC as well. Furthermore, stimulation of ICAM leads to VE-cadherin phosphorylation resulting in destabilization of AJ, thus further facilitating transmigration of leukocytes (Jirik 1989, Grange 2010, Muller 2009, van Buul 2007). IL-6 plays an important role in inflammatory response, thus, is critical for the acute phase response. It is believed that IL-6 resolves acute phase response and promotes acquired immune responses, which is controlled by chemokine-directed leukocyte recruitment but also by efficient activation of leukocyte apoptosis. IL-6-driven STAT3 activation is thought to limit the recruitment of neutrophils as well as pro-inflammatory cytokine. However, IL-6 also rescues cells from apoptosis via the activation of STAT3, and increased expression of anti-apoptotic factors such as Bcl-xL and Bcl<sub>2</sub> (Jones 2005, Fielding 2008). In addition, the expression of isoforms of ROCK is increased. Inhibition of ROCK is thought to impair IL-6-mediated resolution of neutrophils-dependent acute inflammation (Mong 2009). Thus, IL-6 can function as an anti-inflammatory or a pro-inflammatory factor.

Deregulated IL-6/STAT3 pathway underlies a number of vascular diseases including PH, autoimmune diseases and cancer (Mathew 2004, Huang 2008, Hirano 2010, Yu 2009). In addition, the loss of caveolin-1 has been reported in these cases. Caveolin-1 is known to inhibit PY-STAT3 activation as well as the expression of Bcl-xL and Bcl<sub>2</sub>. Caveolin-1 also inhibits and degrades inflammatory and pro-neoplastic protein COX2 (Mathew 2004, Huang 2010, Mathew 2011b, Mathew 2007). Caveolin-1 modulates inflammatory processes via its regulatory effect on eNOS, and depending on the cell type and context of the disease, the effect can be positive or negative.

Hemoxygenase (HO)-1, one of the isoenzymes has emerged as an important player in cellular defense mechanism. HO-1 catalyzes the metabolism of free heme into equimolar ferrous iron, carbon monoxide (CO) and biliverdin. The latter is converted to bilirubin by biliverdin reductase. HO-1 suppresses inflammation by removing pro-inflammatory molecule, heme, and by generating CO. CO, biliverdin and bilirubin have cytoprotective function. HO-1/CO inhibits pro-inflammatory cytokines such as CCL2 and IL-6, and increases the production of IL-10 an anti-inflammatory cytokine. Interestingly, HO-1 and biliverdin reductase are compartmentalized in endothelial caveolae; and similar to eNOS, HO-1 activity is inhibited by caveolin-1. CO has been shown also to activate sGC (Durante 2011, Pae 2009, Liang 2011).

### 3.4 Coagulation and thrombosis

In health, endothelium prevents thrombosis via a number of endothelium-derived inhibitors of coagulation such as thrombomodulin, protein S, heparin sulfate proteoglycans and tissue plasminogen activator (tPA). In addition, PGI<sub>2</sub>, NO and CD39 inhibit platelet aggregation. Released tPA catalyzes the conversion of plasminogen to plasmin thus, facilitating proteolytic degradation of thrombus (Oliver 2005). Activation of coagulation cascade is necessary for normal hemostasis. Tissue factor (TF) is a transmembrane glycoprotein that initiates coagulation cascade; and thrombin is the key effector enzyme for the clotting process. The coagulation cascade is activated to stop the blood loss by forming a clot (Shovlin 2010). TF, a member of cytokine superfamily that functions as high affinity receptor



and a cofactor for plasma factors VII/VIIa, the initiator of blood coagulation. TF is not expressed in EC, but it is rapidly induced by infection and inflammatory cytokines (TNF $\alpha$ , IL-1 $\beta$ ). VEGF, a major stimulator of angiogenesis, is known to upregulate TF expression in EC (Mechtcheriakova 1999). Following injury/infection, Weibel Palade bodies fuse with endothelial cell membrane and release vWF, P-selectin and IL-8. Interestingly, capillary EC lack Weibel Palade bodies but they do express vWF, P-selectin, thus, are capable of participating in coagulation process. The inter-activation of vWF multimers with exposed subintimal matrix results in adherence to activated platelets and participation in clot formation. The release of P-selectin facilitates neutrophil adherence to EC and transmigration (Ochoa 2010).

It is well accepted that there are cross-talks between inflammatory responses and thrombosis. Coagulation has been shown to augment inflammatory responses, and anticoagulants blunt the coagulation-induced inflammatory responses. Furthermore, PGI<sub>2</sub> and APC inhibit injury-induced Ca<sup>2+</sup> flux and NF $\kappa$ B activation, and reduce significantly the expression of proinflammatory cytokines such as TNF $\alpha$ , IL-6 and IL-8. EPCR augments APC by thrombin/thrombomodulin complex; but EPCR is shed from EC by inflammatory mediators and thrombin, thus favoring thrombosis (Esmon 2001).

Under physiological state, circulating platelets are in a quiescent state, and the activation is inhibited by endothelium-derived NO and PGI<sub>2</sub>. Platelets are recruited early to the site of inflammation/injury to provide rapid protection from bleeding; however, they contribute both to coagulation and inflammation. Platelets form a layer, and vWF plays a critical role in the adherence of platelets to the injury site. At the site of adherence, platelets release platelet activating factors such as adenosine diphosphate (ADP), thromboxane A<sub>2</sub> (TxA<sub>2</sub>), serotonin, collagen and thrombin. Thrombin is the most potent thrombogenic factor. In addition, release of ADP and TxA<sub>2</sub> from platelets increases the expression of P-selectin and CD40 ligand (Angeolillo 2010). CD40, the receptor for CD40 ligand, is found on a number of cells including EC, macrophages, B-cells and vascular SMC. The interaction between CD40 and its ligand causes severe inflammatory responses, matrix degradation and thrombus formation; and it has been implicated in the pathogenesis of PH. Platelet-derived member of TNF superfamily “lymphotoxin-like inducible protein that competes with glycoprotein D for herpes virus entry mediator on T lymphocytes” (LIGHT) levels in serum are increased in patients with PAH; interestingly, LIGHT levels are not altered in PH secondary to left heart failure. LIGHT increases the expression of TF and plasminogen activator inhibitor (PAI)-1, and decreases thrombomodulin levels, thus, making EC pro-thrombogenic (Otterdal 2008). PAI-1, a potent endogenous inhibitor of fibrinolysis, is produced by several cells including EC. ROS has been shown to have a significant role in cytokine-induced increase in PAI-1 expression. Increased levels of PAI-1 enhance thrombosis and impair fibrinolysis. Recent studies suggest that PAI-1 regulates EC integrity and cell death. Increased levels are thought to confer resistance to apoptosis and facilitate cell proliferation (Jaulmes 2009, Balsara 2008, Schneider 2008).

### 3.5 Angiogenesis

The formation of new capillaries from a preexisting vessel is called angiogenesis. Angiogenesis plays a pivotal role in a numerous physiological and pathological processes such as organ development, tissue repair and carcinogenesis. Angiogenesis is controlled by