1

PROGRESS IN PROTEIN-LIPID INTERACTIONS

# Progress in Protein-Lipid Interactions

A.Watts and J.J.H.H.M. De Pont Editors

# **Progress in Protein-Lipid**Interactions

### **Editors**

# **A.Watts**

Biochemistry Department South Parks Road Oxford OX1 3QU England

and

# J.J.H.H.M. De Pont

Department of Biochemistry University of Nijmegen Postbus 9101 6500 HB Nijmegen The Netherlands



1985 ELSEVIER AMSTERDAM·NEW YORK·OXFORD All rights reserved. No part of this publication may be reproduced, stored in a retrieval system or transmitted in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without the prior written permission of the publisher, Elsevier Science Publishers B.V. (Biomedical Division), P.O. Box 1527, 1000 BM Amsterdam, The Netherlands.

Special regulations for readers in the USA: this publication has been registered with the Copyright Clearance Center Inc. (CCC), Salem, Massachusetts.

Information can be obtained from the CCC about conditions under which the photocopying of parts of this publication may be made in the USA. All other copyright questions, including photocopying outside of the USA, should be referred to the publisher.

ISBN 0-444-80630-X (volume) ISSN 0168-9614 (series)

Published by: Elsevier Science Publishers B.V. (Biomedical Division) P.O. Box 211 1000 AE Amsterdam The Netherlands

Sole distributors for the USA and Canada: Elsevier Science Publishing Company, Inc. 52 Vanderbilt Avenue New York, NY 10017 USA

# PROGRESS IN PROTEIN-LIPID INTERACTIONS VOLUME 1

An international review series designed to critically evaluate actively developing areas of research in protein-lipid interactions

#### **Series Editors**

## A.Watts

Biochemistry Department South Parks Road Oxford OX1 3QU England

and

## J.J.H.H.M. De Pont

Department of Biochemistry University of Nijmegen Postbus 9101 6500 HB Nijmegen The Netherlands



ELSEVIER AMSTERDAM NEW YORK OXFORD

# Progress in Protein-Lipid Interactions

此为试读,需要完整PDF请访问: www.ertongbook.com

# Preface

'Protein-lipid Interactions' has been initiated with the aim of allowing authors to present not a comprehensive review but a highly critical assessment of specific aspects of research into protein-lipid interactions in membranes. Active specialists have been asked to *freely* reappraise past work in the light of our present understanding, give new, possibly unpublished data, and then identify those areas which they believe show greatest potential for our future understanding of membrane biology.

The series will be aimed at established workers wishing to gain a fresh insight into their own interest in the field of protein-lipid interactions, or new researchers wanting to learn about other active areas of the topic. The editors have made an attempt to maintain a balance between the physical and biochemical approaches to the study of protein-lipid interactions.

In this first volume, one view of the many available theories used to describe protein-lipid interactions has been presented by Jim Abney and Jack Owicki. Magnetic resonance methods, probably still the most controversial, yet direct, approach to describing the dynamic interactions between proteins and lipids, are described by Myer Bloom and Ian Smith for deuterium nuclear magnetic resonance and Derek Marsh for spin-label electron spin resonance. Ben de Kruijff and his colleagues describe the way in which phosphorus nuclear magnetic resonance can identify lipid polymorphism in membranes and also monitor the modulation of these effects by proteins.

Bob Clegg and Winchil Vaz have described fluorescent methods, backed by some theoretical treatment, used to determine the relative movements of lipids and proteins in membranes. David Schachter has given his view of the function of intestinal plasma membranes in relation to lipid fluidity. Roberto Bisson and Cesare Montecucco conclude the first volume with a survey of the use of photoreactive phospholipids to study lipid-protein interactions. Although most work so far has approached protein-lipid interactions from the lipid viewpoint, as re-

flected in the composition of this volume, some attempt has been made to balance function and structure which loosely implies biochemical and biophysical studies. We hope to reach a broad readership and promote interdisciplinary discourse on this fascinating topic.

# List of contributors

#### J.R. Abnev

Department of Biophysics and Medical Physics, University of California, Berkeley, CA 94720, U.S.A.

#### R. Bisson

C.N.R. Center for the Study of the Physiology of Mitochondria and Laboratory of Molecular Biology and Pathology, Institute of General Pathology, University of Padova, Via Loredan 16, 35100 Padova, Italy

#### M. Bloom

Department of Physics, University of British Columbia, Vancouver, BC, Canada V6T 2A6

#### R.M. Clegg

Max Planck Institut für Biophysikalische Chemie, D-3400, Göttingen-Nikolausberg, F.R.G.

#### P.R. Cullis

Department of Biochemistry, University of British Columbia, Vancouver, BC, Canada V6T IW5

#### C.J.A. van Echteld

Department of Cardiology, University Hospital, State University of Utrecht, Catharijnesingel 101, Utrecht, The Netherlands

#### P. van Hoogevest

CIBA-Geigy Ltd., Agricultural Division, R & D Seeds/Biotechnology, Schwarz-waldallee 211, CH-4002 Basel, Switzerland

#### M.J. Hope

Department of Biochemistry, University of British Columbia, Vancouver, BC, Canada V6T IW5

#### J.A. Killian

Department of Biochemistry, State University of Utrecht, Padualaan 8, 3584 CH Utrecht, The Netherlands

#### B. de Kruijff

Institute of Molecular Biology, State University of Utrecht, Padualaan 8, 3584 CH Utrecht, The Netherlands

#### D. Marsh

Max Planck Institut für Biophysikalische Chemie, Abt. Spektroskopie, D-3400 Göttingen, F.R.G.

#### C. Montecucco

C.N.R. Center for the Study of the Physiology of Mitochondria and Laboratory of Molecular Biology and Pathology, Institute of General Pathology, University of Padova, Via Loredan 16, 35100 Padova, Italy

#### J.C. Owicki

Department of Biophysics and Medical Physics, University of California, Berkeley, CA 94720, U.S.A.

#### A. Rietveld

Department of Biochemistry, State University of Utrecht, Padualaan 8, 3584 CH Utrecht, The Netherlands

#### D. Schachter

Department of Physiology, Columbia University, College of Physicians and Surgeons, New York, NY 10032, U.S.A.

#### I.C.P. Smith

Division of Biological Sciences, National Research Council, Ottawa, Canada K1A OR6

#### A.T.M. van der Steen

Organon Teknika, Veedijk 58, B-2300 Turnhout, Belgium

#### T.F. Taraschi

Hahnemann Medical College, Department of Pathology, 230 N. Broad Street, Philadelphia, PA 19102, U.S.A.

#### W.L.C. Vaz

Max Planck Institut für Biophysikalische Chemie, D-3400 Göttingen-Nikolausberg, F.R.G.

#### A.J. Verkleij

Institute of Molecular Biology, State University of Utrecht, Padualaan 8, 3584 CH Utrecht, The Netherlands

# Contents

Pre <sub>s</sub> List	face t of (	Contributors
Chai	pter 1	
		s of protein-lipid and protein-protein interactions in membranes 1
by		
J.R.	Abı	ney and J.C. Owicki
I.	Intro	duction 1
II.	Effe	cts of proteins on membrane lipids
	1.	Summary of experimental results
	2.	Marčelja (1976)
		a. Description of theory
		b. Results
	3.	Schroeder (1977b)
		a. Description of theory
		b. Results
	4.	Landau theories: Owicki et al. (1978), Owicki and McConnell (1979) and Jähnig
		(1981a, b)
		a. Description of theories
		b. Results
	5.	Pink and Chapman (1979); Lookman et al. (1982)
		a. Overview
		b. Description of theory
		c. Results
	6.	Scott and Coe (1982)
		a. Description of theory
		b. Results
	7.	O'Leary (1982)
		a. Description of theory
		h Results 20

	8.	Pearson et al. (1984)	21
		a. Overview	21
		b. Description of theory	21
		c. Results	22
	9.	Lipid mixtures, Owicki et al. (1978)	23
		a. Description of theory	23
		b. Results	23
	10.	Summary	24
III.	Effe	cts of lipids on the activity of membrane proteins	25
	1.	Overview	25
	2.	Activation of proteins by binding of specific lipids	26
	3.	Effects of global state	26
		a. Overview	26
		b. Arrhenius analysis	27
		c. Jähnig and Bramhall (1982)	28
IV.	Prot	ein-protein interactions	31
	1.	Overview	31
	2.	Analysis of electron micrographs	32
		a. Statistical versus statistical-mechanical analyses	32
		b. The radial distribution function and the Percus-Yevick equation	32
		c. Historical development	36
		d. Higher-order distribution functions and the BGY equation	37
		e. Critique of EM analysis	37
	3.	Theoretical predictions of protein-protein interactions	38
		a. Lipid-mediated interactions	38
		b. Computer simulations	43
V.	Thei	rmodynamics of partition between membrane and water	49
	1.	Overview	49
	2.	Partitioning, Jähnig (1983)	49
		a. Description of theory	49
		b. Results	51
	3.	Asymmetry, Weinstein et al. (1982)	52
		a. Description of theory	52
		b. Results	53
	4.	Conclusions	53
VI.	Reco	ommendations for future work	54
		edgements	55
		es	56
Chap	oter 2		
		tations of lipid-protein interactions in deuterium NMR	61
by	nj es	initions of tipin protein timeractions in acaterium 111111	01
	Dlas	m and I.C.P. Smith	
IVI. I	0100	m and i.e.f. Smith	
		A series	
I.		oduction	61
	1.	Diffraction and nuclear magnetic resonance are complementary methods of deter-	
	_	mining protein structure	61
	2.	Deuterium nuclear magnetic resonance (3H-NMR) provides well defined, quan-	
		titative, local structural information on bilayer systems	64

	3. Nuclear magnetic resonance provides quantitative information on molecular mo-
	tion
II.	Review of <sup>2</sup> H-NMR studies of the effect of lipid-protein interactions on the acyl chains
	of phospholipid molecules
	1. The strategy underlying the first attempts to study lipid-protein interactions using
	<sup>2</sup> H-NMR
	2. Summary of <sup>2</sup> H-NMR results in the liquid crystalline region 6
	3. Comments on the <sup>2</sup> H-NMR results in the liquid crystalline region 6
	4. Summary of <sup>2</sup> H-NMR results in the gel region
	5. Comments on the <sup>2</sup> H-NMR results in the gel region
III.	Review of <sup>2</sup> H-NMR studies of the effect of lipid-protein interactions on the polar head-
	group region of lipid molecules
	1. Integral membrane proteins 7
	2. Peripheral membrane proteins
IV.	Relationship between the three-dimensional structure of integral membrane proteins
	and <sup>2</sup> H-NMR results on lipids
	1. The $\alpha$ -helix is a fundamental <i>trans</i> -membrane secondary structural element of
	integral membrane proteins
	2. A simple geometrical interpretation of orientational order in membranes
	3. Experiments with synthetic amphiphilic polypeptide molecules
V.	Review of <sup>2</sup> H-NMR measurements on integral membrane proteins and polypeptides . 7
	1. The study of <sup>2</sup> H-labelled amino acids incorporated biosynthetically into bacterio-
	rhodopsin
	2. <sup>2</sup> H-NMR studies of exchangeable hydrogen sites
VI.	Future prospects
	1. Limitations of the <sup>2</sup> H-NMR method sensitivity, time scale, specificity 8
	2. Thermodynamic studies
	3. Strategic approach to the search for selective lipid-protein binding
	4. More NMR tricks should be used
Ack	nowledgements
	erences
Ittic	Actives
Cha	pter 3
	dulation of lipid polymorphism by lipid-protein interactions 8
by	
B. d	e Kruijff, P.R. Cullis, A.J. Verkleij, M.J. Hope, C.J.A. van Echteld,
T.F	. Taraschi, P. van Hoogevest, J.A. Killian, A. Rietveld and A.T.M.
van	der Steen
Ť	Introduction
I.	
II.	Lipid polymorphism
	THE DESIGNATION OF THE PROPERTY OF THE PROPERT
***	3. The molecular shape concept: a rationale for lipid polymorphism
III.	Lipid-protein interactions and lipid polymorphism
	1. Extrinsic membrane proteins and polypeptides
	a. Poly-L-lysine
	b. Small amphipathic peptides

		c. Cytochrome c and apocytochrome c	106
	2.	Intrinsic membrane proteins and peptides	109
		a. Gramicidin	109
		b. Glycophorin	118
IV.	Fund	ctional aspects	125
	1.	Fusion	125
	2.	Transport	128
	3.	Protein insertion and translocation	130
	4.	Domains	133
V.	Con	cluding remarks	135
Refe	rence	28	135
ESF by	p <b>ter 4</b> R <i>spi</i> Marsl	in label studies of lipid-protein interactions	143
I.	Intr	oduction	143
II.		-component spectra from lipid-protein systems	145
III.		chiometry of lipid association	146
IV.		cificity of lipid-protein associations	154
V.		formational order of the lipids at the protein interface	160
VI.		hange of the lipids at the protein interface	163
		clusions	168
		edgement	170
		es	170
Reic	Tence		170
	pter 5 nslai	tional diffusion of proteins and lipids in artificial lipid bilayer	
mer		nnes. A comparison of experiment with theory	173
by R.M	1. Cle	egg and W.L.C. Vaz	
I.	Intro	oduction	173
II.	The	oretical considerations	174
	1.	Hydrodynamic theories	174
		a. General	174
		b. Three definitions of $f_T$	176
		c. Interpretation of the expression for $f_T$	180
		d. Critique of the hydrodynamic approach	181
		e. Application of the hydrodynamic models, and some general comments	183
	2.	Free-volume theory	184
		a. General	184
		b. Definition of the diffusion constant	186
		c. Extension to activated diffusion	188
		d. Appropriate application of the free-volume theory	188
		e. Critical examination of the two-dimensional free-volume formulation	188

			XIII
		f. Assumptions of the free-volume theory	190
	3.	Molecular models: flexible rod activated jump model for lipid diffusion	191
	5.	a. General	191
		b. Diffusion involving concerted movements of chain segments	192
		c. Selecting one degree of freedom important for diffusion	193
		d. Calculation of the diffusion constant	193
		e. Interchain interaction energy	195
		f. Application of the polymer diffusion model for self-diffusion in lipid bilayers.	199
	4.	Related topics, comparison of theories and suggestions for future work	202
	э.	a. Scope	202
		b. Polymers	202
			203
		c. Monolayers, surface viscosity and Eyring's two-dimensional theory d. Significant structure theory	205
		e. Interfacial diffusion	203
			207
	5	f. Computer molecular dynamics	
***	5.	Theoretical conclusions	207
III.		erimental results	208
	1.	Translational diffusion of integral proteins	208
	2.	Translational diffusion of extrinsic proteins	212
	3.	Translational diffusion of lipids and lipid derivatives	212
	4.	Comparison of experiment with theory	215
IV.		cluding comments	220
		breviations	221
		edgements	223
Refe	erence	es	223
Cha	pter 6		
	•	ynamics and lipid-protein interactions in intestinal plasma	
	,		231
540	nora	ines	231
by			
D. 8	Schac	chter	
I.	Plas	ma membranes of the intestinal enterocyte	231
II.	Lipi	d fluidity of intestinal plasma membranes	233
	1.	Plasma membranes as compared to cytosolic organelles	233
	2.	Microvillus as compared to basolateral membranes	233
	3.	Proximal as compared to distal intestinal segments	236
	4.	Intact membranes as compared to liposomes	236
III.	Lipi	d composition in relation to the fluidity of intestinal plasma membranes	238
	1.	Microvillus as compared to basolateral membranes	238
	2.	Differences in composition along the length of the intestine	240
	3.	Differences in composition of rat microvillus membranes with age	240
IV.		rmotropic transitions	241
	1.	Differential scanning calorimetry and fluorescence polarization studies	241
	2.	Temperature studies of intramolecular excimer fluorescence	244
	3.	Temperature studies of membrane enzyme and transport activities	245
	4.	Delipidation and relipidation studies of microvillus membrane p-nitrophenyl-	.a
		phosphatase	247

### XIV

V.	Regulation of enterocyte plasma membrane fluidity	248
	1. Role of cholesterol biosynthesis	248
	2. Dietary modulation of enterocyte membrane composition and fluidity	250
	3. Ionic regulation of enterocyte microvillus membrane fluidity	252
VI.	Summary and comments	252
Ack	nowledgements	254
Refe	erences	254
	*	
N. 1960		
	pter 7	
Use	e of photoreactive phospholipids for the study of lipid-protein	
inte	eractions	259
by		
R. I	Bisson and C. Montecucco	
I.	Introduction	259
II.	Photoreactive groups	260
III.	Small lipophilic photoreactive reagents	264
IV.	Photoreactive lipids	265
	1. Synthesis of photoreactive phospholipids	265
V.	Use	268
	1. Labelling of intrinsic proteins and determination of their lipid-exposed segments.	271
	2. Photoreactive phospholipids as chemical rulers	273
	3. Peptide folding in the membrane	276
	4. Half-bilayer labelling	280
	5. Photoactive phospholipids as photoaffinity reagents	282
VI.		283
Acknowledgements		
	erences	283 283
Sub	oject index	289

CHAPTER 1

# Theories of protein-lipid and protein-protein interactions in membranes

JAMES R. ABNEY and JOHN C. OWICKI

Department of Biophysics and Medical Physics, University of California, and Division of Biology and Medicine, Lawrence Berkeley Laboratory, Berkeley, CA, 94720 U.S.A.

#### I. Introduction

About 15 years ago it became apparent to a number of biochemists and biophysicists that biological membranes and model membranes behave in many ways like two-dimensional fluids. It also became apparent that a substantial fraction of membrane-associated proteins are integrally embedded in the bilayer membrane, and that some completely traverse it. These realizations were synthesized into the fluid-mosaic model of biological membranes (Singer and Nicolson, 1972), a paradigm that holds sway today with some modifications. One such modification, for example, is to include the immobilizing effects of cytoskeletal interactions.

The intervening years have brought substantial increases in our knowledge about membranes, due in large part to the application of new or refined experimental methods. There have been concomitant efforts on the part of theorists to construct detailed molecular or thermodynamic models that integrate and interpret the experimental work. The largest number of theoretical papers have dealt with pure lipid bilayers, especially the lipid bilayer phase transition. These have been reviewed well elsewhere (e.g., Nagle, 1980), and we do not consider them here except as they relate to our main topics: theoretical models of protein-lipid and protein-protein interactions in membranes.

Abbrevations: DMPC, dimyristoylphosphatidylcholine; DMR, deuterium magnetic resonance; DPPC, dipalmitoylphosphatidylcholine; DSC, differential scanning calorimetry; EPR, electron paramagnetic resonance; SPT, scaled particle theory;  $T_c$ , temperature of lipid bilayer phase transition.

We have focused on the influences that intrinsic membrane proteins have on membrane lipids, the effects of the lipids on the proteins, and the phase behavior and lateral distribution of both components. Unfortunately, spatial constraints have required the omission of a number of major topics. We do not discuss the mechanisms of lateral diffusion or the relationship of the diffusion coefficient to molecular geometry and environment. Nor do we discuss the behavior of cholesterol in membranes, in spite of the fact that cholesterol is much like a very small intrinsic membrane protein and has been the subject of a large amount of research.

We have concentrated on the most detailed statistical-mechanical and thermodynamic treatments. Our goal has been to give a reasonably comprehensive picture of the insights that theory has provided into a number of important membrane phenomena. We have also tried, in the spirit of this series, to assess deficiencies in the theoretical treatments and to recommend areas where more theoretical work seems most needed.

Finally, it is worth stating our opinion of the rôle of theory in this field. Clearly, predictive power and agreement with prior experiments are important for any scientific theory. Nevertheless, biological membranes – and even model membranes – are so complicated that any tractable theory must become only a caricature of the real system. If the caricature leads to recognizable consequences, then we hope that its few remaining features are indeed the most important ones for explaining the observations. Given the rampant approximations, conclusions must be rather tentative. It is then useful to examine several differing treatments of a phenomenon, to see whether some consensus emerges.

#### II. Effects of proteins on membrane lipids

#### 1. SUMMARY OF EXPERIMENTAL RESULTS

A substantial body of experiments, beginning with the electron paramagnetic resonance (EPR) results of Jost et al. (1973), has shown that intrinsic membrane proteins alter the properties of nearby phospholipids in the membrane. We will take the liberty of summarizing these findings below without detailed attribution, directing the interested reader to review articles for further information (Chapman et al., 1979; Jost and Griffith, 1980; Marsh and Watts, 1982; Seelig et al., 1982; Silvius, 1982; Marsh, 1983).

Magnetic-resonance experiments have shown remarkably similar results for different proteins. Typically, EPR experiments on spin-labeled lipids show distinguishable signals from bulk lipid and lipid associated with the protein as discussed in Chapter 4 of this Volume. The latter, usually called boundary lipid or annular lipid, shows little motional averaging on the EPR time scale (ca. 10–100 ns). In