

# ADVANCES IN CANCER RESEARCH

*Edited by*

GEORGE KLEIN

SIDNEY WEINHOUSE

*Volume 29—1979*

# ADVANCES IN CANCER RESEARCH

*Edited by*

**GEORGE KLEIN**

Department of Tumor Biology  
Karolinska Institutet  
Stockholm, Sweden

**SIDNEY WEINHOUSE**

Fels Research Institute  
Temple University Medical School  
Philadelphia, Pennsylvania



**Volume 29—1979**



**ACADEMIC PRESS** New York San Francisco London

A Subsidiary of Harcourt Brace Jovanovich, Publishers

# ADVANCES IN CANCER RESEARCH

Edited by

GEORGE KLEIN

Department of Tumor Biology  
Karolinska Institute  
Stockholm, Sweden

COPYRIGHT © 1979, BY ACADEMIC PRESS, INC.

ALL RIGHTS RESERVED.

NO PART OF THIS PUBLICATION MAY BE REPRODUCED OR  
TRANSMITTED IN ANY FORM OR BY ANY MEANS, ELECTRONIC  
OR MECHANICAL, INCLUDING PHOTOCOPY, RECORDING, OR ANY  
INFORMATION STORAGE AND RETRIEVAL SYSTEM, WITHOUT  
PERMISSION IN WRITING FROM THE PUBLISHER.

ACADEMIC PRESS, INC.

111 Fifth Avenue, New York, New York 10003

*United Kingdom Edition published by*  
ACADEMIC PRESS, INC. (LONDON) LTD.  
24/28 Oval Road, London NW1 7DX

LIBRARY OF CONGRESS CATALOG CARD NUMBER: 52-13360

ISBN 0-12-006629-7

PRINTED IN THE UNITED STATES OF AMERICA

79 80 81 82 9 8 7 6 5 4 3 2 1



## CONTRIBUTORS TO VOLUME 29

Numbers in parentheses indicate the pages on which the authors' contributions begin.

- L. F. CHASSEAUD, *Department of Metabolism and Pharmacokinetics, Huntingdon Research Centre, Huntingdon, United Kingdom* (175)
- ARNOLD S. DION, *Institute for Medical Research, Camden, New Jersey* 08103 (347)
- A. CLARK GRIFFIN, *Department of Biochemistry, The University of Texas System Cancer Center, M.D. Anderson Hospital and Tumor Institute, 6723 Bertner Avenue, Houston, Texas* 77030 (419)
- ETIENNE Y. LASFARGUES, *Institute for Medical Research, Camden, New Jersey* 08103 (347)
- CAROLE A. LONG, *Department of Microbiology and Immunology, Hahnemann Medical College and Hospital, Philadelphia, Pennsylvania* 19102 (347)
- J. F. A. P. MILLER, *The Walter and Eliza Hall Institute of Medical Research, P.O. Royal Melbourne Hospital, Victoria, Australia* (1)
- DAN H. MOORE, *Department of Microbiology and Immunology, Hahnemann Medical College and Hospital, Philadelphia, Pennsylvania* 19102 (347)
- DAVID NAOR, *Lautenberg Center for General and Tumor Immunology, The Hebrew University-Hadassah Medical School, Jerusalem, Israel* (45)
- ERKKI RUOSLAHTI, *Division of Immunology, City of Hope National Medical Center, Duarte, California* 91010 (275)
- MARKKU SEPPÄLÄ, *Department of Obstetrics and Gynecology, University Central Hospital and Department of Bacteriology and Immunology, University of Helsinki, Helsinki* 00290, Finland (275)
- JOEL B. SHEFFIELD, *Department of Biology, Temple University, Philadelphia, Pennsylvania* 19122 (347)
- JOSÉ URIEL, *Institut de Recherches Scientifiques sur le Cancer, B.P. N°8, 94800 Villejuif, France* (127)
- AKHIL B. VAIDYA, *Department of Microbiology and Immunology, Hahnemann Medical College and Hospital, Philadelphia, Pennsylvania* 19102 (347)

# CONTENTS

CONTRIBUTORS TO VOLUME 29 .....	ix
---------------------------------	----

## Influence of the Major Histocompatibility Complex on T-Cell Activation

J. F. A. P. MILLER

I. Introduction .....	1
II. The Major Histocompatibility Complex: A Brief Description .....	2
III. T-Lymphocyte Subsets .....	3
IV. MHC Gene Products Influence T-Cell Activities .....	4
V. H-2 Restriction Phenomena .....	5
VI. Models Offered to Explain H-2 Restriction .....	11
VII. Level at Which H-2 Restriction Is Imposed .....	14
VIII. H-2 Restriction: Relationship to MHC-Linked <i>Ir</i> Genes .....	21
IX. Possible Models for T-Cell Recognition and <i>Ir</i> Gene Effects .....	29
X. H-2 Restriction and Resistance to Tumors .....	36
XI. Summary and Conclusions .....	39
References .....	39

## Suppressor Cells: Permitters and Promoters of Malignancy?

DAVID NAOR

I. Background and Introduction .....	45
II. Effect of External Intervention on Antitumor Resistance: A Historical Perspective .....	52
III. Are Suppressor Cells the Initiators of "Immunostimulation" and "Sneaking through" Phenomena? .....	60
IV. Relationships between Malignant Cells and Suppressor Cells .....	64
V. Suppressor Cells Induced by Nontumorigenic Stimuli or by Tumor Excision .....	104
VI. Conclusions .....	106
VII. Appendix .....	114
References .....	119

## Retrodifferentiation and the Fetal Patterns of Gene Expression in Cancer

JOSÉ URIEL

I. Introduction .....	127
II. Fetal Patterns in Malignant Tumors .....	130



III. Fetal Patterns in Noncancerous Growth .....	147
IV. The Plasticity of the Differentiated State .....	152
V. Retrodifferentiation and Cancer .....	166
References .....	169

## The Role of Glutathione and Glutathione S-Transferases in the Metabolism of Chemical Carcinogens and Other Electrophilic Agents

L. F. CHASSEAUD

I. Introduction .....	176
II. Glutathione .....	176
III. Glutathione S-Transferases .....	177
IV. Compounds that Conjugate with Glutathione .....	200
V. Discussion .....	251
References .....	255

## $\alpha$ -Fetoprotein in Cancer and Fetal Development

ERKKI RUOSLAHTI AND MARKKU SEPPÄLÄ

I. Introduction .....	276
II. Detection and Measurement of AFP and Its Immunochemical Properties .....	276
III. Sites of Physiological AFP Synthesis .....	282
IV. Physiological Concentrations of AFP in Body Fluids .....	284
V. Purification and Chemical Properties of AFP .....	290
VI. Biological Properties of AFP .....	305
VII. AFP in Liver Disease .....	313
VIII. Germ Cell Tumors .....	323
IX. Other Clinical Conditions with Informative AFP Levels .....	327
X. Tolerance to AFP and Its Abrogation .....	331
XI. AFP as a Possible Target Molecule for Tumor Immunoprevention and Immunotherapy .....	333
XII. Molecular Basis of Regulation of AFP Synthesis .....	334
XIII. Areas of Future Research and Concluding Remarks .....	335
References .....	336

## Mammary Tumor Viruses

DAN H. MOORE, CAROLE A. LONG, AKHIL B. VAIDYA, JOEL B. SHEFFIELD,  
ARNOLD S. DION, AND ETIENNE Y. LASFARGUES

I. Introduction .....	347
II. Morphology .....	350
III. Structural Proteins and RNA-Directed DNA Polymerase of MuMTV .....	356

IV. Synthesis and Assembly of MuMTV Proteins .....	364
V. Genome of MuMTV .....	367
VI. Biology of MuMTV .....	370
VII. Antigens of MuMTV .....	386
VIII. Interaction of MuMTV with the Host's Immune System .....	395
IX. Comments on the Question of a MuMTV-Related Virus in Humans .....	405
X. Concluding Remarks .....	408
References .....	410

## Role of Selenium in the Chemoprevention of Cancer

### A. CLARK GRIFFIN

I. Introduction .....	419
II. Selenium Toxicology and Pathology .....	420
III. Nutritional Aspects of Selenium .....	421
IV. Selenium and Cancer .....	423
V. Biological Functions of Selenium .....	429
VI. Possible Mechanisms of Action of Selenium in the Inhibition of Carcinogenesis .....	432
VII. Summary .....	438
References .....	440
SUBJECT INDEX .....	443
CONTENTS OF PREVIOUS VOLUMES .....	447

# INFLUENCE OF THE MAJOR HISTOCOMPATIBILITY COMPLEX ON T-CELL ACTIVATION

J. F. A. P. Miller

The Walter and Eliza Hall Institute of Medical Research  
Post Office Royal Melbourne Hospital  
Victoria, Australia

I. Introduction .....	1
II. The Major Histocompatibility Complex: A Brief Description .....	2
III. T-Lymphocyte Subsets .....	3
IV. MHC Gene Products Influence T-Cell Activities .....	4
V. H-2 Restriction Phenomena .....	5
A. Cytotoxicity .....	6
B. Delayed-Type Hypersensitivity .....	6
C. Helper Functions .....	7
D. Suppression .....	8
E. Possible Exceptions to H-2 Restriction .....	9
VI. Models Offered to Explain H-2 Restriction .....	11
A. Intimacy Model .....	11
B. Altered Self-Model .....	12
C. Two Receptor Model .....	13
VII. Level at Which H-2 Restriction Is Imposed .....	14
A. Priming of T Cells .....	15
B. Differentiation of T Cells in Thymus .....	16
VIII. H-2 Restriction: Relationship to MHC-Linked <i>Ir</i> Genes .....	21
A. Formation of Complex of Antigen and MHC Product Immunogenic for T Cells .....	21
B. Generation of the T-Cell Repertoire .....	26
IX. Possible Models for T-Cell Recognition and <i>Ir</i> Gene Effects .....	29
A. Positive Selection for T Cells with Anti-Self-H-2 Receptors .....	29
B. Selection against T Cells with High-Affinity Anti-Self-H-2 Receptors .....	32
C. MHC Gene Products on Antigen-Presenting Cells Are Obligatory for T-Cell Activation .....	34
X. H-2 Restriction and Resistance to Tumors .....	36
XI. Summary and Conclusions .....	39
Abbreviations Used .....	39
Note Added in Proof .....	39
References .....	40

## I. Introduction

The major stimulus for the activation of most T lymphocytes does not seem to be antigen alone, but antigen in association with one or the other of the gene products of the major histocompatibility complex.



These products are present on the surface of the body's own cells and some of these, for example, cells of the monocyte-macrophage series, are particularly well equipped to present antigen in the appropriate form to T lymphocytes and to deliver the activating signal. Once activated, the T cells have their specificities directed, not to antigen alone, but to both the antigen and some histocompatibility gene product ("H-2 restriction," Zinkernagel and Doherty, 1974a). This discovery has stimulated a vast number of investigations aimed at defining the precise mechanism by which T cells are activated, the specificities of the receptors that enable T cells to recognize antigenic determinants, and the means by which the repertoire of T-cell reactivities may be generated in the genome. This work has led to a better understanding of the function of histocompatibility gene products and of the advantages of polymorphism of the loci which code for these products. It is the purpose of this review to consider some of these investigations and to offer certain models which may elucidate the role of the histocompatibility gene products in T-cell activation. The effect these genes may have in determining resistance to certain tumors will be briefly considered.

## II. The Major Histocompatibility Complex: A Brief Description

The features of the major histocompatibility complex (MHC)<sup>1</sup> relevant to this discussion are shown in Fig. 1. The MHC, H-2 in mice or HLA in man, codes for antigens (glycoproteins) many of which are expressed on the surface membrane of most cells. If we disregard S and G, we can divide H-2 antigens into two major classes: those controlled by the peripheral K or D regions (or classical H-2 antigens) and those controlled by the central I region [or I-associated (Ia) antigens]. The genetic control of the classical H-2 antigens is determined by the loci H-2K in the K region and H-2D in the D region. Due to polymorphism, there is a large number of alleles of H-2. Each allele is designated by a small letter as a superscript: e.g., H-2K<sup>b</sup>, H-2K<sup>d</sup>, H-2K<sup>k</sup>, etc., for the H-2K locus, and H-2D<sup>b</sup>, H-2D<sup>d</sup>, H-2D<sup>k</sup>, etc., for the

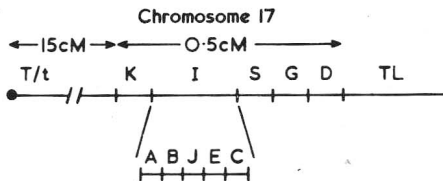


FIG. 1. The major histocompatibility complex or H-2 in the mouse.

H-2D locus. Each haplotype (a particular combination of H-2 antigens controlled by a single chromosome) is designated also by a small letter as a superscript: e.g., H-2<sup>b</sup>, H-2<sup>d</sup>, etc.

I-associated antigens or Ia antigens are genetically controlled by the I region and are distributed among five loci (so far detected): I-A, I-B, I-J, I-E, and I-C. Antigens coded by different I regions behave as on separate molecules. In contrast to H-2K and H-2D products which are represented on most cells, I-region products are found predominantly on cells of the lymphoreticular system (subsets of T cells, B cells, macrophages, etc.) and also on the epidermal cells of Langerhans. There are differences in the tissue distribution of individual Ia antigens coded by different I subregions (e.g., Ia antigens coded by the I-J locus are found predominantly on suppressor T cells). Many of the MHC-linked *Ir* genes have been mapped in the I-region.

On the right of D is the TL locus which codes for antigens expressed on thymocytes and on cells of some thymus-derived lymphomas. On the left of K is the T/t complex. The genes of this complex mediate a series of essential steps in embryogenesis probably by coding for cell surface components at relevant stages in development. There is a reciprocal relationship between one of these antigens, F9, and the H-2 antigens. F9 but not H-2 is present on sperms and on cells of the very early embryo. Later H-2 is present but not F9. TL has also some reciprocal relationship with H-2D: less H-2D is present when TL antigens are expressed on thymocytes. TL is not expressed on mature T cells. F9, H-2K, H-2D, and TL antigens are glycoproteins of similar molecular weight and three of these have the small molecule  $\beta_2$  m associated with them. It is therefore of great interest that four loci, within a region of some 15 cM on chromosome 17, specify analogous cell membrane molecules playing an important role in cell to cell interactions. This has led to the hypothesis that this family of four loci are the logical descendants of an ancestral locus which acquired the property of cell surface perception as a condition for metazoan evolution.

Further details on the genetic organization of the MHC will be found in Jan Klein's excellent monograph (1975) and in a number of reviews (e.g., Shreffler, 1977; McKenzie, 1977; Peterson *et al.* 1977; Artz and Bennett, 1975).

### III. T-Lymphocyte Subsets

T cells act in different ways in immune responses and may be divided into various subsets (Simpson and Beverley, 1977). Some T cells are directly cytotoxic ( $T_c$ ) if they can make intimate contact with ap-

appropriate targets. Some are involved in inducing the inflammatory lesions of delayed-type hypersensitivity (DTH,  $T_D$  cells). Some help B cells produce IgG antibody ( $T_H$ ) and some regulate the immune response by suppressing the activities of other T and B cells ( $T_S$ ). The various subsets can be distinguished by cell surface markers, such as the differentiation antigens of the Ly series and the differential expression of antigens coded by the MHC (Vadas *et al.*, 1976; Cantor and Boyse, 1977). Thus, in general,  $T_H$  and  $T_D$  cells are  $Ly-1^+$  and  $Ia^-$ ;  $T_C$  cells are  $Ly-2, 3^+$  and usually  $Ia^-$ , whereas  $T_S$  cells are  $Ly-2, 3^+$  and express Ia antigens coded by the I-J locus of the MHC. The exact way in which the various T-cell subsets exert their effect is not known but evidence points to the production, after antigenic stimulation, of factors which may be antigen specific or nonantigen specific. For example,  $T_H$  cells are thought to influence IgG-producing B cells either by a nonantigen-specific factor acting indirectly via macrophages (Miller, 1975), or by releasing antigen-specific factors of various types (Feldmann and Nossal, 1972; Munro and Taussig, 1975). Activated  $T_D$  cells release lymphokines such as the migration inhibitory factor (MIF), which are nonantigen specific and influence the recruitment of cells of the mononuclear-macrophage series (Bloom and Bennett, 1970).  $T_C$  cells presumably kill by some nonantigen-specific influence against target cells for which they have antigen-specific receptors (Cerottini and Brunner, 1974). Finally, antigen-specific helper or suppressor factors have been obtained from  $T_H$  and  $T_S$  cells and carry determinants coded by the I-region of the MHC. Unfortunately, none of the factors thought to be elaborated as a result of stimulation of the various T-cell subsets has yet been precisely characterized chemically.

#### IV. MHC Gene Products Influence T-Cell Activities

The MHC exerts a profound influence on T-cell activities. This is clear from the following.

(1) The frequency of alloreactive T cells (i.e., those directed against H-2 antigens) is generally thought to be 100 to 1000 times as high as the frequency of T cells reactive to non-H-2 antigens (Simonsen, 1967; Lindhal and Wilson, 1977).

(2) Different subsets of T cells exhibit the phenomenon of H-2 restriction, i.e., identity at certain MHC gene products is required between cells responsible for immune induction of T cells and targets of immune T cells (Zinkernagel and Doherty, 1974a; Bevan, 1975; Miller *et al.*, 1975; Katz and Benacerraf, 1975; Section V).

(3) The *Ir* genes, linked functionally to the MHC, influence the responsiveness of T cells or T-cell dependent functions (Benacerraf and Katz, 1975; Benacerraf and Germain, 1978; Section VII).

(4) There are I-region coded determinants on antigen-specific "helper" and "suppressor" factors which can be released or extracted from T<sub>H</sub> and T<sub>S</sub> cells, respectively (Feldmann *et al.*, 1977; Tada *et al.*, 1977).

The first three phenomena can all be accommodated in one general model of T-cell recognition and activation. This will be attempted in Section IX. On the other hand, the antigen-specific helper and suppressor factors will not be considered in detail in this review. It is felt that much more work is required to characterize these factors physiologically and biochemically to appreciate their special role in the network of immune interactions.

To account for the high frequency of alloreactive T cells, Jerne (1971) proposed that the repertoire of T-cell reactivities is coded by a set of germ-line *v* genes (genes coding for the *variable* region of T-cell antigen-binding units) which code for structures essentially complementary to the H-2 alleles of the species. After entering the thymus, potentially alloreactive T cells, which form a relatively large proportion of the T-cell pool, need not be influenced. By contrast, T cells with anti-self-H-2 reactivities would proliferate in response to H-2 structures present in the thymus. This must not be allowed to continue, for it was argued that such cells could kill self-H-2-bearing cells. Random somatic mutations in the genes coding for structures complementary to self-H-2 would thus be allowed to accumulate. This would decrease the strength of anti-self-H-2 binding and finally, only those T cells without anti-self-H-2 reactivities would mature. Such cells would have their reactivities directed to non-H-2 antigens and each specific set would thus constitute a much smaller proportion of the total T-cell pool than alloreactive T cells. As will be discussed later (Section IX), the discovery of the phenomenon of H-2 restriction has rekindled interest in this negative selection theory of Jerne. It has in fact been reformulated to account for a variety of the new findings (Langman, 1978; Blanden and Ada, 1978; von Boehmer *et al.*, 1978).

#### V. H-2 Restriction Phenomena

Most immune phenomena associated with T cells are subject to H-2 restriction. This has been well documented for T cells involved in cytotoxicity, delayed-type hypersensitivity (DTH), and helper func-

tions. By contrast, there is no evidence for H-2 restriction of B cell or natural killer (NK) cell activity.

### A. CYTOTOXICITY

T<sub>C</sub> cells may have their reactivities directed to H-2 antigens (other than self) or directed to non-H-2 antigens. For example, CBA (H-2<sup>k</sup>) mice immunized to DBA/2 (H-2<sup>d</sup>) produce T<sub>C</sub> cells which kill any targets bearing either H-2K<sup>d</sup> or H-2D<sup>d</sup>, no matter what other cell surface antigens these targets may have. Alloreactive T cells are thus clearly not self-H-2 restricted but rather "allo-H-2" restricted in view of their specificities to alloantigens. On the other hand, CBA mice recovering from infection by the virus of lymphocytic choriomeningitis (LCM) have T<sub>C</sub> cells which attack LCM-infected H-2<sup>k</sup> targets, not uninfected H-2<sup>k</sup> targets or LCM-infected targets of any other H-2 type. There thus appears to be a requirement for H-2 matching between T<sub>C</sub> cells and targets. This constitutes the phenomenon of "self-H-2 restriction" (hereinafter referred to as H-2 restriction) as originally described by Zinkernagel and Doherty (1974a). By using inbred strains of mice, congenic lines and recombinant lines, the genes imposing H-2 restriction of T<sub>C</sub> cells could be mapped. The H-2K and the H-2D genes were found to impose H-2 restriction of T<sub>C</sub> cells generated against virus-specified antigens (Doherty *et al.*, 1976a), hapten-modified antigens (such as trinitrophenyl, TNP) (Shearer *et al.*, 1976), and minor histocompatibility (H) antigens (non-H-2 H antigens designated H<sub>1</sub>, H<sub>3</sub>, H<sub>4</sub>, etc., perhaps up to H<sub>100</sub>, and the H-Y antigen which is a sex-linked minor H antigen) (Bevan, 1975; Gordon *et al.*, 1975).

T<sub>C</sub> cells have clonally distributed receptors (the word receptor is used loosely to describe a cell surface structure with a site having some degree of complementarity to a determinant on an antigen molecule). In other words, one particular T<sub>C</sub> cell has a receptor endowed with a unique specificity directed against either H-2K or H-2D gene products, but not both. There is therefore genic and allelic exclusion of receptors on the T cell (in contrast to the presence of both H-2K and H-2D gene-coded antigens on the same cell) (Zinkernagel and Doherty, 1975).

### B. DELAYED-TYPE HYPERSENSITIVITY

To achieve successful transfer of DTH, there is a requirement for H-2 matching between donors of sensitized T cells and naive recipients (Miller *et al.*, 1975). In the case of protein and polypeptide anti-



gens, the MHC region involved in restriction was I-A, but in the case of contact chemicals such as dinitrofluorobenzene, the genes were either H-2K, H-2D, or in the I region (Vadas *et al.*, 1977). These restrictions were interpreted as a requirement for H-2 matching between T<sub>D</sub> cells and cells, such as macrophages or epidermal cells, responsible for presenting antigen to the T cells. More formal evidence for this was obtained in experiments which showed the necessity for Ia matching between antigen-pulsed macrophages used for sensitization and for elicitation of sensitivity in an *in vivo* system (Miller *et al.*, 1978). This is in line with observations of other investigators showing a requirement for Ia matching between antigen-pulsed macrophages and sensitized T cells *in vitro* to allow these to proliferate (Yano *et al.*, 1977; Thomas *et al.*, 1977b). The *in vitro* system enabled the investigators to use specific antibodies to determine the role played by antigen and MHC components in stimulating the proliferation of sensitized T cells. Incubation of T cells, themselves, with anti-Ia antibody had no effect. Antibody directed to Ia antigens, of the appropriate specificities, present on macrophages blocked the ability of the antigen-pulsed macrophages to stimulate proliferation (Shevach *et al.*, 1972; Thomas *et al.*, 1977a). The blocking was haplotype-specific in the F<sub>1</sub>, indicating that it did not result from some nonspecific effect of the binding of antibody to a cell surface component (Schwartz *et al.*, 1976a). By contrast, antibody to the native antigen failed to block proliferation (Ellner *et al.*, 1977; Thomas *et al.*, 1978). The antigenic determinants which stimulate T cells therefore appear to do so in association with Ia determinants but are not readily accessible to antibodies directed against *native* antigen. Similar observations have been made in cytotoxic systems: the specific inhibition of lysis of target cells by T<sub>C</sub> cells was achieved by antibody directed to the H-2K and H-2D gene products of the targets (Germain *et al.*, 1975; Schmitt-Verhulst *et al.*, 1976). Antiviral antibodies, on the other hand, consistently failed to protect the target (Blanden *et al.*, 1976b; Doherty *et al.*, 1976a; Braciale, 1977), except in one case (Koszinowski and Ertl, 1976). The implications of these observations are that native antigen is not recognized as such by T lymphocytes.

### C. HELPER FUNCTIONS

It was initially observed that T<sub>H</sub> cells and B cells must share I-A gene products for successful cooperation in antibody responses (Katz and Benacerraf, 1975). The restriction of T<sub>H</sub> cells occurred both at the level of induction (presumed to reflect activation of T<sub>H</sub> cells by



macrophage-associated antigen) and during T- and B-cell cooperation (Sprent, 1978a). T-cell recognition of MHC-associated antigen on macrophages and on specific B cells may thus be either identical or very similar. If, as discussed above, the antigen recognized by T cells on macrophage surfaces is not native antigen, the same must be true of antigen presented by B cells.  $T_H$  cells may recognize the B cells which they help, only if these can associate antigen with their surface Ia determinants in the same way as the antigen-pulsed macrophages. T- and B-cell cooperation thus might not depend on carrier-hapten "focusing" between T- and B-cell receptors specific for determinants of the carrier and the hapten, respectively. The T cells specific for Ia and "processed" antigen would seek out those B cells displaying identical structures. Of course, only B cells with specific receptors for either hapten or carrier determinants should "capture" sufficient hapten-carrier conjugates via their surface immunoglobulin receptors. Only these B cells should be able to "process" carrier determinants and associate them with Ia determinants on their cell surface, thus displaying the correct structure for specific carrier-reactive  $T_H$  cells. Binding of the specific T-cell receptors to such structures on the surface of the B cells may be associated with the delivery of an activating signal (nonantigen specific) from the  $T_H$  cell to the B cell, and the induction of synthesis of the corresponding specific antibody by the B cell. Hence, antibodies to both carrier and hapten determinants would be produced by carrier-specific and hapten-specific B cells.

There may be a requirement for H-2 matching (also in the I-A region) between  $T_H$  cells and  $T_C$  cells, as suggested by recent work in chimeric mice (Zinkernagel *et al.*, 1978a) (see Section VII).

#### D. SUPPRESSION

The antibody response to protein antigens may be suppressed by an antigen-specific factor, bearing an I-J-coded determinant and obtained from  $T_s$  cells. Identity at the I-J subregion between donor and recipient resulted in more effective suppression in some (Tada *et al.*, 1977) but not all cases (Kapp, 1978). When  $T_s$  cells were induced to hapten-modified cell membranes as, for example, when mice were tolerized with dinitrophenyl (DNP)-modified lymph node cells, the  $T_s$  cells suppressed recipient mice only if these shared H-2D with the strain providing the lymphoid cells (S. D. Miller *et al.*, 1978). The reactivity of the  $T_s$  cells in this system thus appeared to be directed to DNP-modified H-2D products. Somewhat similar results were obtained with soluble suppressor factors released from  $T_s$  cells

(Moorhead, 1977). A requirement for H-2D compatibility was also reported for T<sub>s</sub> cells active in virus-infected systems (Pang and Blanden, 1976; Kumar and Bennett, 1977).

### E. POSSIBLE EXCEPTIONS TO H-2 RESTRICTION

A number of phenomena have not exhibited the classical H-2 restriction described above. They are listed under the following headings and some will be referred to again in Section X.

#### 1. T<sub>c</sub> Cells for I-Region Determinants

I-region determinants can serve both as stimulator and as targets of T<sub>c</sub> cells (Wagner *et al.*, 1975; J. Klein *et al.*, 1977; Billings *et al.*, 1977a). H-2K or H-2D compatibility is not required for the activity of these T<sub>c</sub> cells. In addition, I-region determinants are susceptible to the same type of antigen modification (e.g., by TNP) as H-2K or H-2D gene products. In this case, the modified Ia determinants serve as targets for specific T<sub>c</sub> cells raised against I-region determinants (Billings *et al.*, 1977b).

#### 2. T<sub>c</sub> Cells for F9 Antigen

OTT 6050 is a teratocarcinoma line, derived from strain 129 mice. It does not express H-2K or H-2D antigen specificities but produces an antigen, F9, genetically linked to the T/t locus of the MHC (Jacob, 1977). Although virus-infected F9<sup>+</sup> cells cannot serve as targets for T<sub>c</sub> cells generated *in vivo* by infection with the virus (Zinkernagel and Oldstone, 1976; Doherty *et al.*, 1977), *in vivo* priming of mice with H-2 negative F9<sup>+</sup> cells followed by *in vitro* restimulation with F9<sup>+</sup> cells triggers the generation of anti-F9 immune T<sub>c</sub> cells able to lyse specifically H-2 negative F9<sup>+</sup> cells (Wagner *et al.*, 1978). Since there may be some evolutionary relationship between the H-2 and T/t complexes, both located on chromosome 17 (Artz and Bennett, 1975), F9<sup>+</sup> cells may code for a functional analog of the H-2K and H-2D gene products. Whether it is possible to induce F9-restricted virus-specific or hapten-specific T cells has yet to be demonstrated.

#### 3. Cross-Reactive Lysis by T<sub>c</sub> Cells

T<sub>c</sub> cells activated to allogeneic cells *in vivo* or *in vitro* in mixed lymphocyte cultures are very heterogeneous as evidenced by their cross-reactivity patterns. The effector cells kill not only targets syngeneic to stimulator cells but also third-party allogeneic targets with different H-2 antigens (Lindahl *et al.*, 1975). Cross-reactive lysis has

also been demonstrated both against minor H-antigens (Bevan, 1977) and when normal spleen cells were cultured with irradiated TNP-conjugated syngeneic spleen cells. This generated T<sub>C</sub> cells which lysed syngeneic TNP-targets efficiently and, to a lesser extent, allogeneic TNP targets (Burakoff *et al.*, 1976). Cold target inhibition experiments indicated the existence of clones of cross-reactive T<sub>C</sub> cells and treatment of targets with anti-H-2 antibody blocked lysis thus demonstrating the requirement for recognition of H-2 on targets. If the repertoire of T-cell reactivities evolved to recognize modified self-H-2 antigens (Jerne, 1971), T<sub>C</sub> cells with reactivities to alloantigens or xenoantigens might consist of clones of cells with specificities directed to modified autologous MHC products which cross-react with allogeneic and xenogeneic antigens. In an experimental investigation of this possibility, alloreactivity generated by xenogeneic stimulation was shown to result from the activities of separate T<sub>C</sub> cell clones, each specific for an allogeneic target (Burakoff *et al.*, 1977). Moreover, alloreactive T<sub>C</sub> cells exhibited cross-reactivities with chemically modified target cells syngeneic to the responders (Lemonnier *et al.*, 1977). Such cross-reactive T<sub>C</sub> cells may account for the apparent breakdown of H-2 restriction observed in some tumor immunity systems (see Section X).

#### 4. Anomalous Cytotoxicity

"Anomalous" cytotoxicity has been detected in a variety of experimental systems. Thus, for example, heterospecific cytotoxic cells were generated in the first 3 to 5 days of acute LCM infection in mice. These cells killed many types of infected or uninfected cultured cells, including syngeneic cells (Pfizenmaier *et al.*, 1975; Blanden and Gardner, 1976; Welsh and Zinkernagel, 1977). Likewise, Epstein-Barr virus (EBV)-genome carrying lymphoblastoid cell lines (LCL) stimulated autologous lymphocytes to generate cytotoxic cells reactive not only against the autologous cell line but also against other unrelated cell lines, whether or not these carried the EBV genome (Viallat *et al.*, 1978a,b). A somewhat analogous situation was described in human mixed lymphocyte cultures (Seeley and Golub, 1978). Here, two distinct types of cytotoxic activities were generated during sensitization to normal allogeneic peripheral blood lymphocytes: (a) allospecific cytotoxicity directed against alloblasts only; (b) "anomalous" cytotoxicity directed against serologically unrelated LCL and against the autochthonous LCL. The peak anomalous cytotoxicity occurred 1-2 days earlier than the peak allocytotoxicity and declined earlier. The anomalous cytotoxicity was not limited to targets sharing HLA