

EDITED

BY

J.B.C. FINDLAY

Membrane Protein Models

J.B.C. Findlay

Department of Biochemistry and Molecular Biology, University of Leeds, Leeds, UK

© BIOS Scientific Publishers Limited, 1996

First published 1996

All rights reserved. No part of this book may be produced or transmitted, in any form or by any means, without permission.

A CIP catalogue record for this book is available from the British Library.

ISBN 1 85996 080 4

BIOS Scientific Publishers Ltd 9 Newtec Place, Magdalen Road, Oxford OX4 1RE, UK Tel. +44 (0)1865 726286. Fax +44 (0)1865 246823

DISTRIBUTORS

Australia and New Zealand
DA Information Services
648 Whitehorse Road, Mitcham
Victoria 3132

Singapore and South East Asia
Toppan Company (S) PTE Ltd
38 Liu Fang Road, Jurong
Singapore 2262

India

Viva Books Private Limited 4325/3 Ansari Road, Daryaganj New Delhi 110002

USA and Canada

Books International Inc.
PO Box 605, Herndon, V A 22070

The colour section in this book is kindly sponsored by Orion-Farmos

Typeset by Euroset, Alresford, Hampshire, UK. Printed by Information Press Ltd, Oxford, UK.

Membrane Protein Models

Contributors

- **Argos, P.** European Molecular Biology Laboratory, Postfach 102209, Meyerhofstrasse 1, D-69012 Heidelberg. Germany
- **Bhogal, N.** Department of Biochemistry and Molecular Biology, University of Leeds, Leeds LS2 9JT, UK
- Blaney, F.E. SmithKline Beecham Pharmaceuticals, Computational Chemistry Department, New Frontiers Science Park (North), Third Avenue, Harlow, Essex CM19 5AW, UK
- **Bradshaw, C.** Glaxo Institute for Molecular Biology, Plan-les-Quates, CH-1228 Geneva, Switzerland
- Chollet, A. Glaxo Institute for Molecular Biology, Plan-les-Quates, CH-1228 Geneva, Switzerland
- Cocchi, M. Dipartimento di Chimica, Universita' di Modena, Via Campi 183, I-41100 Modena, Italy
- **De Benedetti, P.G.** Dipartimento di Chimica, Universita' di Modena, Via Campi 183, I-41100 Modena, Italy
- **Donnelly, D.** Department of Biochemistry and Molecular Biology, University of Leeds, Leeds LS2 9JT, UK
- Fanelli, F. Dipartimento di Chimica, Universita' di Modena, Via Campi 183, I-41100 Modena, Italy
- Finbow, M.E. Beatson Institute, Garscube Estate, Switchback Road, Bearsden, Glasgow G61 1BD, UK
- Findlay, J.B.C. Department of Biochemistry and Molecular Biology, University of Leeds, Leeds LS2 9JT, UK
- **Harrison, M.A.** Department of Biochemistry and Molecular Biology, University of Leeds LS2 9JT, UK
- **Hurrell, C.R.** Department of Biochemistry and Molecular Biology, University of Leeds, Leeds LS2 9JT, UK
- IJzerman, A.P. Leiden-Amsterdam Center for Drug Research, Division of Medicinal Chemistry, Gorlaeus Laboratories, P.O. Box 9502, NL-2300 RA, The Netherlands
- Jones, P.C. Department of Biochemistry and Molecular Biology, University of Leeds LS2 9JT, UK

x Contributors

Kim, Y.-I. Department of Biochemistry and Molecular Biology, University of Leeds LS2 9JT, UK

- **Kuipers, W.** Department of Medicinal Chemistry, Solvay Duphar B.V., P.O. Box 900, NL-1380 DA Weesp, The Netherlands
- **Lybrand, T.P.** University of Washington, Center for Bioengineering, P.O. Box 351750, Seattle, WA 98195-1750, USA
- Menziani, M.C. Dipartimento di Chimica, Universita' di Modena, Via Campi 183, I-41100 Modena, Italy
- **Milpetz, F.** European Molecular Biology Laboratory, Postfach 102209, Meyerhofstrasse 1, D-69012 Heidelberg. Germany
- Nemeth, K. Glaxo Institute for Molecular Biology, Plan-les-Quates, CH-1228 Geneva, Switzerland
- Oliveira, L. Department of Biophysics, Escola Paulista de Medicina, Sao Pualo 04043-971, Brasil
- Paiva, A.C.M. Department of Biophysics, Escola Paulista de Medicina, Sao Paulo 04043-971, Brasil
- **Persson, B.** Department of Medical Biochemistry and Biophysics, Karolinska Institute, S-17199 Stockholm, Sweden
- Rippmann, F. E. Merck, Preclinical Pharmaceutical Research, D-64271 Darmstadt, Germany
- Sander, C. European Molecular Biology Laboratory, BIOcomputing department, Meyerhofstrasse 1, D-69012 Heidelberg, Germany
- Sankararamakrishnan, R. Department of Physiology and Biophysics, Beckman Institute, University of Illinois at Urbana-Champaign, Urbana, IL 61801, USA
- Sansom, M.S.P. Laboratory of Molecular Biophysics, University of Oxford, The Rex Richards Building. South Parks Road, Oxford OX1 3QU, UK
- **Tennant, M.** SmithKline Beecham Pharmaceuticals, Computational Chemistry Department, New Frontiers Science Park (North), Third Avenue, Harlow, Essex CM19 5AW, UK
- **Thomas, P.** Department of Biomolecular Structure, Glaxo Research and Development Limited, The Medicines Research Centre, Gunnels Wood Road, Stevenage, Hertfordshire SG1 2NY, UK
- **Turcatti, G.** Glaxo Institute for Molecular Biology, Plan-les-Quates, CH-1228 Geneva, Switzerland
- **Vriend, G.** European Molecular Biology Laboratory, BIOcomputing department, Meyerhofstrasse 1, D-69012 Heidelberg, Germany
- **Zuurmond, H.M.** Leiden-Amsterdam Center for Drug Research, Division of Medicinal Chemistry, Gorlaeus Laboratories, P.O. Box 9502, NL-2300 RA, The Netherlands.

Abbreviations

5-CT 5-carboxamidotryptamine 5-hydroxytryptamine ASP atomic solvation parameter benzophenone-4-maleimide

bR bacteriorhodopsin
BSA bovine serum albumin
CFP channel-forming peptides
DCCD dicyclohexylcarbodiimide
DHA (-)-[³H]dihydroalprenolol

DOI 1-(2,5-dimethoxy-4-Br-phenyl)-2-aminopropane

8-OH-DPAT 2-(di-n-propylamino)-8-hydroxytetralin

ET endothelin

F-M fluorescein-5-maleimide FTIR Fourier transform infrared

GA gramicidin A

GPCR G protein-coupled receptors

HLA histocompatibility antigen molecules

IE interaction energies

LSD lysergic acid diethylamide MD molecular dynamics

nAChR nicotinic acetylcholine receptor NBD 7-nitrobenz-2-oxa-1,3-diazol-4-yl

NK neurokinin

PBS phosphate-buffered saline QMD quenched molecular dynamics

QSAR quantitative structure–activity relationship

SA/MD stimulated annealing via restrained molecular dynamics

SAR structure–activity relationship

SDS-PAGE sodium dodecyl sulphate-polyacrylamide gel electrophoresis

TCA trichloracetate TM transmembrane

Preface

Integral membrane proteins

The membrane has represented one of the most intractable elements of the biological cell. From the time when it was realized that there was a barrier within the plant cell wall and that this barrier represented the most important and fundamental regulator of the cellular environment, considerable effort has been expended to determine its nature and properties. The lipid fraction was the first to be described but although that was achieved some time ago and interest has waned somewhat since then, there are still fundamental mysteries to explain, not least the real link between function and composition. Knowledge of the protein fraction lagged some way behind, principally due to the intractability of these polypeptides to the methodologies then available. But with the advent/use of such simple molecules as detergents and organic solvents has come the dramatic advances in our understanding of membrane proteins from the primitive days of scaffold-like fragments to the tantalizing mechanisms suggested by the high-resolution 3-dimensional structures of the photosynthetic reaction complex and cytochrome oxidase.

Now that the molecular description of biology is in full flood, it is clearly important to be able to describe the structure and mechanisms of action of integral membrane proteins, both the types that are responsible for the translocation of material and those which mediate the transfer of information. But this urgent need has not so far been fulfilled because yet again, routine methodology is still not available. This has given rise to alternative strategies to gain impressions, albeit crude and unreliable ones, of the structures of integral membrane proteins. The usefulness of such representations in facilitating experimental design and in stimulating the development of new concepts should not be underestimated, however, for there is ample evidence of their predictive potential at least at a low resolution level. During the conference of which these are the proceedings, many of the strategies used to further our structural understanding of integral membrane proteins were described and discussed. Many models were presented and their strengths, weaknesses, contradictions and inconsistencies thoroughly explored.

xiv Preface

G protein-coupled receptors

Most attention was devoted to the G protein-coupled receptors (GPCRs), surely now the most avidly pursued and widespread family of proteins in eukaryotic biology. Little did those few groups who wrestled with the mechanism of action, sequence and topography of the visual pigment rhodopsin appreciate the avalanche of structural and functional relatives that lie in the various eukaryotic genomes. The elucidation of the seven transmembrane segments of opsin has given rise to one of the most potent signatures of any protein family. There are now in the database over 700 (and rising) sequences which are recognized by this composite signature. Clearly, this is still only a small proportion of the total if the estimates of the number of olfactory receptors alone are in any way accurate. Already, however, interesting divisions are appearing which in some ways reflect evolutionary distance. The classical sequence-based motifs are incapable of recognizing sub-families such as those for yeast mating factor receptors, the dictyostelium cAMP receptors, the secretin subgroup and receptors for glutamate, Ca, gonadotrophin hormones etc. All appear to be involved in some way with G proteins but perhaps the mechanism of interaction and activation differs.

The assumption, so far unproven, is that the basic framework of these receptors remains more or less intact but considerable functionality is incorporated, often in association with new structural elements or additional domains. Thus, as the ligand gets larger, as with the peptide receptors, the hydrophilic surface regions including the extended N-terminus are recruited in to generate specific binding pockets. This development reaches full expression in the large N-terminal extensions seen in the follicle-stimulating hormone, thyroid-stimulating hormone and luteinizing hormone receptors which have a major role in interacting with the protein ligand. Even larger domains also occur in the glutamate and calcium receptors but here it is harder to give a rational explanation for these regions given the small size of the ligand. Whatever their precise role, these large domains are nevertheless both structurally and functionally intimately associated with the transmembrane segments through which the information flow must pass. It is to be expected that additional elements of structure will also be found at the intracellular face of the receptor. This has already been seen in the cephalopod visual receptors which possess a very unusual region at the C-terminus, one that intriguingly occurs in other, unrelated proteins.

On the basis of limited biophysical evidence, bacteriorhodopsin was the template used for the first tertiary structure representations of GPCRs. This assumption was justified when the low resolution structure of rhodopsin appeared. But it is also clear that the template was only an approximate one for there is variation in the relative position and pitch of some of the transmembrane helices. It is reasonable to assume that rhodopsin is now a more accurate template but, at the same time, it would not

Preface xv

be unexpected to find that further differences occur throughout the family and more likely still amongst the more distant and less conserved sub-families.

This conference was designed as a discussion forum to explain and examine the various approaches to modelling integral membrane proteins in general and GPCRs in particular. It revealed wide differences in interpretation particularly of the mutagenesis data. The protein models themselves, despite having different origins and being generated by different methods, apparently had a surprising degree of convergence. The really striking differences were seen in the area of ligand docking where quite different interpretations were put on much the same sets of data. The inevitable conclusion from this was that models whilst stimulating, provocative and reasonably predictive required much firmer structure-based data before one could reliably move from low resolution representations to higher resolution 'structures'.

Finally, one should not overlook the evolving but still unconfirmed concepts in GPCR structure/function relationships. The ternary complex model and the suggestions of a range of conformational intermediates between the activated R* (agonist binding) and inactive R (antagonist binding) modes present new structural challenges. So too do the increasingly frequent suggestions of different binding epitopes for agonists and antagonists, particularly when the former is large and the latter is relatively small. The even more detailed models of GPCRs will have to take on board these subtle concepts and observations. Part of this appreciation must involve a rational description of the activated state, particularly as it applies to the fascinating array of constitutively active mutants.

Transporters/channels

Whilst there is some structural basis for modelling GPCRs, the situation for transporters/channels is much more desperate, at almost all levels of resolution and types of data. The best information so far comes from the structure of bacterial outer membrane porins but it is not at all clear how representative these will be of the majority of channels, since the entire membrane domain consist of an emaculate β -barrel. The suggestions made for the acetylcholine (nicotinic) receptor are intriguing in that a bundle of helices appear to provide the transport route but these may be embedded in a continuous shell of β -sheet. Such data as exist for other transporters/channels emphasize a substantial α -helical content, sometimes incorporating a channel-lining segment of unknown secondary structure. Thus, it looks as though we may have a fascinating mixture of types with much scope for imagination. However, there are both modelling and experimental approaches which might provide useful insights into the structural and functional properties of proteins which mediate material transport. A few examples of these are presented here to illustrate their potential application to model construction.

xvi Preface

The meeting was entitled: 'Membrane Protein Models: Experiment, Theory and Speculation' (Leeds, UK, March/April 1994, under the auspices of The Molecular Graphics Society). From the excellent presentations that are included in this text, the reader will quickly appreciate the lively and stimulating debates that took place on all three aspects of the structure and function of integral membrane proteins.

J.B.C. Findlay

Contents

Con	ntributors	ix
Abb	previations	xi
Pref	face selection setting and the	xiii
1.	Prediction of transmembrane segments in proteins using multiple	
	sequence alignments. B. Persson, F. Milpetz and P. Argos	1
	Introduction	1
	Present prediction methods	2
	Residue distributions in membrane proteins	2
	Propensity values	3
	Prediction algorithm	7
	Optimization of the method	8
	Computer program	9
	Electronic mail	9
	Evaluation of the new algorithm	17
	Incorrect predictions	21
	Comparisons with other prediction methods	23
	Conclusions	23
	References	23
2.	Sequence–function correlation in G protein-coupled receptors.	
	W. Kuipers, L. Oliveira. A.C.M. Paiva, F. Rippmann, C. Sander,	3
	G. Vriend and A.P. IJzerman	27
	Introduction	27
	Data and methods	28
	Results and discussion	33
	Conclusions	
	References	43
3.	Modelling α-helical transmembrane domains. D. Donnelly,	
	N. Bhogal, C.R. Hurrell and J.B.C. Findlay	47
	Introduction	47
	Predicting structure from sequence	47

Contents

	Models of GPCRs	49
	The peptide binding site of the NK2 receptor	50
	References	53
4.	α-Helix bundles and ion channels. R. Sankararamakrishnan	
	and M.S.P. Sansom	55
	Introduction	55
	Methods	56
	Results	56
	Conclusions	68
	References	70
5.		
	P. Thomas	73
	Introduction	73
	What do experimental studies tell us about the structure of	
	G protein-coupled receptors?	73
	Evidence from theoretical analysis of GPCR sequences	76
	Deriving a template for GPCR models Conclusions	79 87
	References	88
	Telefolious and the second sec	00
6.	Pride and prejudice and G protein-coupled receptor models. <i>F. Rippmann</i>	91
	G protein-coupled receptors	91
	Structures of integral membrane proteins	91
	Sequence analysis of GPCRs	92
	Three-dimensional models of GPCRs	93
	Conclusions	100
	References	102
	Colombia	105
	Colour section	105
7		
7.	The heuristic-direct approach to quantitative structure—activity relationship analysis of ligand—G protein-coupled receptor complexes. M.C. Menziani, F. Fanelli, M. Cocchi and	
	P.G. De Benedetti	113
	Introduction	113
	Application: the endothelin receptors	115
	Computational strategy	118

Contents	vii

	Results and discussion	121
	Conclusions	128
	References	129
8.	Molecular modelling of β-adrenoceptors. A.P. IJzerman and	
	H.M. Zuurmond	133
	Introduction	133
	Methods	134
	Results and discussion	135
	Conclusion	143
	References	143
9.	Three-dimensional models for β -adrenoceptor-ligand complexes.	
	T.P. Lybrand	145
	Introduction	145
	Methodology	146
	Application to adrenoceptor models	148
	Discussion	156
	Conclusions	157
	References	157
10.	Computational tools and results in the construction of	
	G protein-coupled receptor models. F.E. Blaney and M. Tennant	161
	Introduction	161
	Construction of three-dimensional GPCR models	162
	Methods	163
	Conclusion	175
	References	175
11.	Fluorescence studies of neurokinin receptors. A. Chollet,	
	G. Turcatti, K. Nemeth and C. Bradshaw	177
	Introduction	177
	Methods	178
	Fluorescent ligands for the NK2 receptor	180
	Ligand–receptor interactions	182
	Probing receptor function by fluorescence	188
	Conclusions and future directions	190
	References	191

viii Contents

12.	A structural model for the 16 kDa membrane sector of the vacuolar H ⁺ - ATPase. J.B.C. Findlay, M.E. Finbow, P.C. Jones,	
	YI. Kim and M.A. Harrison	193
	Introduction	193
	Testing the molecular model of the 16 kDa proteolipid	198
	Functional implications of proteolipid structure	204
	References	207
	Index	209

Prediction of transmembrane segments in proteins using multiple sequence alignments

Bengt Persson, Frank Milpetz and Patrick Argos

1. Introduction

Membrane proteins are important for several processes and functions in all biological systems. For example, they act as receptors for neurotransmitters or hormones (Savarese and Fraser, 1992; Stephenson, 1991), form a wide variety of ion channels (Barnard, 1992; Miller, 1991), or serve as the respiratory chain (Capaldi, 1991) and transport proteins for different molecules (Griffith *et al.*, 1992; Marger and Saier, 1993; Schloss *et al.*, 1992). Bacterial toxins which form membrane pores (Li, 1992) also belong to this large group of lipid-associated molecules. There are several surface molecules which are anchored to the membrane by one transmembrane segment, for example histocompatibility antigen molecules (HLA), neuraminidases and haemagglutinins (cf. Popot and de Vitry, 1990).

Despite their biological significance tertiary structures have been determined for only a few membrane proteins. Three-dimensional structures at medium to high resolution are available for bacteriorhodopsin (Henderson *et al.*, 1990), photosynthetic reaction centre (Deisenhofer *et al.*, 1985), light-harvesting complexes (Kühlbrandt *et al.*, 1994; McDermott *et al.*, 1995), prostaglandin H₂ synthase-1 (Picot *et al.*, 1994), porin (Weiss and Schulz, 1992) and cytochrome *C* oxidase (Iwata *et al.*, 1995). Other structures are not yet fully resolved, for example photosystem I (Krauss *et al.*, 1993) and nicotinic acetylcholine receptor (Unwin, 1993). Given the scarcity of tertiary structural information, many experimental methods have been applied to determine membrane topology (Jennings, 1989), including analyses of gene fusion proteins and studies of biochemically modified membrane proteins (cf. Traxler *et al.*, 1993).

2 Chapter 1

Membrane proteins constitute a ubiquitous group of structures with members representing several different types of molecular architecture. However, since each traverses the lipid bilayer once or several times, they generally possess hydrophobic sequence segments. Various prediction methods use this characteristic to determine the location of these membrane-spanning regions, albeit with varying degrees of accuracy.

2. Present prediction methods

Theoretical prediction algorithms have been shown to be useful in detecting membrane-spanning segments from the primary structure alone, especially as an aid to designing experiments investigating protein topology. One of the most widely used is that of Kyte and Doolittle (1982), where mean residue hydrophobicity values are calculated for consecutive 19-residue sequence spans. Segments with hydrophobicity above a certain threshold are predicted to be membrane-spanning. A similar approach is adopted by Rao and Argos (1986), who also considered residues that break the transmembrane helices in order to improve reliability of prediction. Different prediction methods were reviewed and evaluated in 1990 by Degli Esposti et al. They examined the correlations amongst the various amino acid hydrophobicity scales used and compared the accuracy of the various prediction approaches. They also calculated a new set of parameters derived from seven different scales (Degli Esposti et al., 1990). A trapezoidal sliding window was used by von Heijne in his hydrophobic analysis of the sequence together with a consideration of positively charged residues interior to the membrane (von Heijne, 1986) particularly with application to the topology of bacterial inner membrane proteins (von Heijne, 1992). These rules were also applied to a number of eukaryotic membrane proteins (Sipos and von Heijne, 1993). Several studies have also been effected regarding helix-helix interactions in membrane proteins (e.g. Lemmon and Engelman, 1992).

Here we present a transmembrane helix prediction algorithm, based upon multiple sequence alignments of related proteins. This method takes advantage of the extended information not found in analysis of a single sequence as is characteristically used by other approaches. The algorithm is described in detail elsewhere (Persson and Argos, 1994). Present primary structural databases are large and expanding at such a rate that homologous sequences are often found. We show that this technique has higher accuracy in predicting transmembrane segments than previous methods based on individual sequences.

3. Residue distributions in membrane proteins

The tertiary structure of only a few membrane proteins are known. However, other types of data exist from various experiments and predictions to deduce the location