

# The Design and Statistical Analysis of Animal Experiments

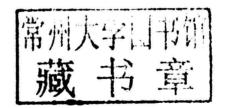
Simon T. Bate

GlaxoSmithKline, UK

and

Robin A. Clark

Huntingdon Life Sciences, UK







University Printing House, Cambridge CB2 8BS, United Kingdom

Published in the United States of America by Cambridge University Press, New York

Cambridge University Press is part of the University of Cambridge.

It furthers the University's mission by disseminating knowledge in the pursuit of education, learning and research at the highest international levels of excellence.

www.cambridge.org

Information on this title: www.cambridge.org/9781107030787

© S. T. Bate and R. A. Clark 2014

This publication is in copyright. Subject to statutory exception and to the provisions of relevant collective licensing agreements, no reproduction of any part may take place without the written permission of Cambridge University Press.

First published 2014

Printed in the United Kingdom by Clays, St Ives plc

A catalogue record for this publication is available from the British Library

Library of Congress Cataloguing in Publication data

Bate, Simon T., 1975-

The design and statistical analysis of animal experiments / Simon Bate, GlaxoSmithKline, UK, Robin Clark, Huntingdon Life Sciences, UK.

pages cm.

Includes bibliographical references and index.

ISBN 978-1-107-03078-7 (hardback) - ISBN 978-1-107-69094-3 (paperback)

Animal experimentation-Statistical methods.
Experimental design.
Clark, Robin A., software developer.
Title. HV4930.D47 2002

590.72'4-dc23

2013033851

ISBN 978-1-107-03078-7 Hardback

ISBN 978-1-107-69094-3 Paperback

Additional resources for this publication at www.cambridge.org/9781107030787

Cambridge University Press has no responsibility for the persistence or accuracy of URLs for external or third-party internet websites referred to in this publication, and does not guarantee that any content on such websites is, or will remain, accurate or appropriate.

# The Design and Statistical Analysis of Animal Experiments

Written specifically for animal researchers, this is the first book to provide a comprehensive guide to the design and statistical analysis of animal experiments. It has long been recognised that the proper implementation of these techniques can help to optimise the number of animals used in an experiment. By using real-life examples to make them more accessible, this book explains the statistical tools that are routinely employed by practitioners.

A wide range of design types are considered in detail, including block, factorial, nested, crossover, dose-escalation and repeated measures. Alongside each design, techniques are introduced to analyse the experimental data generated. Each analysis approach is described in non-mathematical terms, helping readers without a statistical background to understand key techniques such as: *t*-tests, ANOVA, repeated measures, analysis of covariance, multiple comparison tests, non-parametric methods and survival analysis.

This is also the first text to describe technical aspects of InVivoStat, a powerful open-source software package developed by the authors to enable animal researchers to analyse their data and obtain informative results. InVivoStat can be downloaded at www.invivostat.co.uk.

Simon T. Bate is a Principal Statistician at GlaxoSmith-Kline, supporting pre-clinical research. He has spent over 12 years supporting the design and statistical analysis of animal experiments, including drug discovery research, toxicology studies and safety assessment. He presents many statistics courses around Europe, including the statistics module of the British Association for Psychopharmacology's Pre-Clinical Certificate.

**Robin A. Clark** is a Senior Analyst Programmer at Huntingdon Life Sciences, Alconbury. Originally qualified as a marine ecologist, he is now a software architect producing data collection and statistical applications for the pharmaceutical industry. Robin produced the user interface and designed the system architecture for InVivoStat.

### To

### RB, NRB, MTB, EC, EJC, ZMC

### Preface

This book is aimed at practitioners who do not have a statistics degree and yet wish to apply statistics to help them arrive at valid and reliable conclusions while minimising the animal numbers required. Descriptions of the mathematical methods underpinning the topics covered in the book are purposefully kept to a minimum. If readers wish to gain a better understanding of the mathematics behind experimental design and statistical analysis then reading a more advanced textbook would help further their understanding.

The solutions to practical problems encountered when conducting animal experiments are explained using non-technical approaches. We believe that in many situations advanced statistical ideas can be employed successfully by researchers with no statistical qualification, using a combination of common sense and modern statistical analysis software packages. In our experience statistical ideas are often introduced to scientists using mathematical terminology. This can be off-putting to non-mathematicians and can leave researchers with, at best, only rudimentary statistical tools and at worst a fear of statistics.

To keep the descriptions of the statistical tools covered in this book as simple as possible, we shall occasionally give pragmatic explanations. While such explanations may not apply in all cases and in all scientific disciplines, this approach does allow us to introduce methods in a clear and concise way. By allowing ourselves the freedom to simplify the problems pragmatically, we aim to make statistical tools more accessible. The reader is invited, once they have familiarised themselves with (and hopefully found the benefit of using) the tools described in this book, to read more advanced texts on the subject.

This book is divided into seven chapters which loosely correspond to the procedure a researcher should take when planning the experimental design, running the experiment and evaluating the data generated. Following an introductory chapter and a second describing certain statistical concepts, the third chapter covers different types of designs. Designs are outlined, where possible, in simple non-technical language. This is followed by a chapter describing the randomisation of the experimental material. The fifth chapter discusses the statistical analysis of animal experiments and this is followed by a chapter describing how these methods can be applied within the statistical software package

InVivoStat. The final chapter draws some conclusions about the ideas contained within the text.

A scientist can apply all of the methodology described in this book. Certain topics covered are more advanced than others and while we aim to make all subjects accessible, the reader should be aware that the help of a professional statistician may be advisable when first implementing some of the more advanced tools. However, once the readers have familiarised themselves with the ideas contained within this book, we hope they will have a fuller appreciation of the help statistics can offer to improve the conclusions that can be made when running animal experiments.

## **Acknowledgments**

We would like to thank all our colleagues, both past and present, for providing so much inspiration for this book. We hope this text reflects the discussions, consultations and the range of examples that we have worked on together. There are simply too many scientists to mention, but you know who you are! We hope the wide variety of interesting practical problems we have faced together is reflected throughout this book.

Special thanks also to the statisticians who have advised us over the years and shaped our views on this subject matter. Simon would especially like to thank Philip Overend and Andrew Lloyd for their input and encouragement and Janet and Ed Godolphin for their help and support over the years.

We would like to give an extra special thanks to Gillian Amphlett, Gill Fleetwood, David Willé, Clare Stanford and Nathalie Percie du Sert for giving up their time to review sections of the book, and providing ideas, advice and feedback. Their input has been most helpful and improved the text considerably.

Finally we would like to thank our friends and family for putting up with the long unsociable hours we spent writing this book.

# Contents

Preface			page xii
Acknow	ledgmer	nts	x
	O		
1 In	troductio	on	1
1.	1 Struc	ture of this book	3
	1.1.1	Introductory sections	4
	1.1.2	Approaches to consider when	
		setting up a new animal model	4
	1.1.3	Approaches to consider when	
		generating hypotheses	5
	1.1.4	Approaches to consider when	
		testing hypotheses	5
1.3	2 Statis	tical problems faced by animal	
	resea	rchers	Ę
1.3	3 Pitfal	ls encountered when applying	
	statis	tics in practice	6
	1.3.1	Pitfalls with experimental design	6
	1.3.2	Pitfalls with randomisation	ç
	1.3.3	Pitfalls with statistical analysis	10
		Pitfalls when reporting animal	
		experiments	13
1.4	4 So wh	nere does statistics fit in?	15
1.5	5 The A	RRIVE guidelines	15
2 St	atistical c	concepts	18
2.	Decis	ion-making: the signal-to-noise rat	io 18
2.2	2 Proba	bility distributions	19
	2.2.1	The frequency distribution	20
	2.2.2	The density distribution	20
	2.2.3	The probability distribution	21
	2.2.4	The normal distribution	21
	2.2.5	The chi-squared distribution	22
	2.2.6	The <i>t</i> -distribution	22
	2.2.7	The F-distribution	23

			The hypothesis testing procedure		23		3.2.8		edly measuring the	
		2.3.1		ll and alternative				animal		45
			hypoth		23	3.3			esign types	46
		2.3.2	The $p$ -v		25			Block		46
		2.3.3	-	nificance level	25		3.3.2		al designs	47
		2.3.4	Signific	eant stars	26		3.3.3	Dose-re	esponse designs	47
		2.3.5	Type I a	and Type II errors	26		3.3.4	Nested	designs	47
	2.4	Explo	ratory vs	. confirmatory			3.3.5	Split-pl	ot designs	48
		exper	iments		28		3.3.6	Repeat	ed measures and	
	2.5	The es	stimation	process	29			dose-es	scalation designs	48
							3.3.7	Designs	s applied in practice	48
3	Expe	erimen	al design	1	30	3.4	Block	designs		49
	3.1	Why o	lesign ex	periments?	30		3.4.1	Practica	al reasons to block	49
		3.1.1	Practica	al reasons	30		3.4.2	Statistic	cal reasons to block	49
		3.1.2	Statistic	cal reasons:				3.4.2.1	Variance reduction	49
			variabil	ity, the signal and bias	31			3.4.2.2	Bias reduction	51
	3.2	What	does an e	experimental			3.4.3	How to	block	51
		design	involve	?	32		3.4.4	Comple	ete block designs	53
		3.2.1	Variable	es to be recorded	32			3.4.4.1	Efficiency	53
			3.2.1.1	Types of response	32			3.4.4.2	Randomisation	53
			3.2.1.2		34			3.4.4.3	Statistical analysis	
			3.2.1.3		34				of block designs	54
			3.2.1.4	175			3.4.5	Incomp	olete block designs	54
				during the experiment	35		3.4.6	_	ed incomplete	
		3.2.2	Set of tr	reatments	35			block d	-	55
		3.2.3		perimental unit and				3.4.6.1	Efficiency	55
				ervational unit	36			3.4.6.2	Randomisation	55
		3.2.4		and factors	37			3.4.6.3	Statistical analysis	55
			3.2.4.1	Defining factor			3.4.7		nan one block: the	
			J	level labels	39				lumn block design	56
			3.2.4.2	Defining the factors in				3.4.7.1	Efficiency	56
			0121112	an experimental design	39				Randomisation	56
		3.2.5	Fixed a	nd random factors	39				Statistical analysis	56
		0.2.0		Fixed factors	40		3.4.8		olumn block	50
			3.2.5.2	Random factors	40		3.1.0		based on Latin squares	57
				Random or fixed?	41				Efficiency	58
		3.2.6		rical factors and	11			3.4.8.2	Randomisation	58
		3.2.0	_	ious factors	42			3.4.8.3	Statistical analysis	58
		3.2.7		factors and	42		3.4.9		ver designs	59
		3.2.1	nested		42		3.4.3	3.4.9.1	Complete crossover	33
			3.2.7.1	Nested factors	42			J.4.J.1	designs	59
				Crossed factors	42			2400		39
			3.2.7.2					3.4.9.2	Incomplete crossover	co
			3.2.7.3	Partially crossed factors				2.4.0.0	designs	60
			3.2.7.4	Designs containing nest				3.4.9.3		
				and crossed factors	45				designs	61

		3.4.9.4	The issues with crossove	r			3.6.3.3	Adding an offset to	
			designs	62				the dose	88
		3.4.9.5	Treatment carry-over		3.7	Neste	d designs	3	90
			effects	62		3.7.1	Types o	f nested design	91
3.5	Facto	rial desig	n	63			3.7.1.1	Single-order nested	
	3.5.1	Randon	nisation	64				design	91
	3.5.2	Categor	rical factors and				3.7.1.2	Higher-order	
		interact	ions	64				nested design	91
	3.5.3	Small fa	ctorial designs	66		3.7.2	Sample	size and power	93
	3.5.4	Large fa	actorial designs	68			3.7.2.1	Factors that influence	
		3.5.4.1	Strategies when setting					sample size	93
			up a new animal model	68			3.7.2.2	Calculating sample	
		3.5.4.2	Graphical representation	1				sizes	95
			of large factorial				3.7.2.3	When not to calculate	
			designs	70				the statistical power	97
		3.5.4.3	Hidden replication	70		3.7.3	Higher-	order nested	
		3.5.4.4	Fractional factorial				designs		99
			designs to reduce				3.7.3.1	Identifying nested	
			animal use	72				factors	99
		3.5.4.5	Two-stage procedure to				3.7.3.2	Investigating the source	es
			reduce animal use	75				of variability in higher-	
	3.5.5	Factoria	al designs with					order nested designs	101
		continu	ous factors	77			3.7.3.3	Variance components:	
		3.5.5.1	Strategies for setting up a	a .				estimating the observa-	
			new animal model	78				tional unit variability	102
		3.5.5.2	Drug combination				3.7.3.4	Predicting the	
			studies	81				experimental unit	
		3.5.5.3	Continuous vs.					variability	103
			categorical factors	83			3.7.3.5	Investigating alternative	
	3.5.6		oughts on factorial					nested designs	105
		designs		83				Pseudo-replication	106
3.6		response		84	3.8	•		sures and dose-	
	3.6.1		r- and five-				ition desi		110
	121212		ter logistic curves	84		3.8.1		ed measures	
	3.6.2		nental design				designs		110
			erations	85				The repeated factor	110
		3.6.2.1	Increasing the number				3.8.1.2	The core experimental	
			of doses	86				design	112
		3.6.2.2	Decreasing the number				3.8.1.3	Nested repeated	
			of animals	86				measures designs	112
	3.6.3		ng the control group	87			3.8.1.4	More complex repeated	
		3.6.3.1	Analysing a change from			0.00	5	measures designs	114
		0.000	the control response	87		3.8.2		scalation designs	116
		3.6.3.2	Using a dual statistical				3.8.2.1	More complex	
			model	88				dose-escalation designs	s 117

	3.9	Split-p	olot desig	ns	117			5.2.1.3	The predicted mean	137
		3.9.1	Animals	s as whole plots	117			5.2.1.4	The geometric mean	137
		3.9.2	Animals	s as subplots	118		5.2.2	Parame	tric measures of	
	3.10	Exper	imental c	lesigns in practice	119			spread		138
	3.11	A goo	d design :	should result in	120			5.2.2.1	Variance	138
4	Done	lomisa	tion		122			5.2.2.2	Standard deviation	138
4	4.1			ns to randomise	122			5.2.2.3	Standard error of the	
	4.1		Bias red		122				mean	138
		4.1.1	4.1.1.1	Removing unforeseen	122			5.2.2.4	Confidence intervals	139
			1.1.1.1	trends	123			5.2.2.5	Coefficient of	
			4.1.1.2	Humans are systematic					variation	139
		4.1.2	Blinding		124		5.2.3		rametric measures	
	4.2			ons to randomise	124			of location		
		4.2.1		ing the variability	125		5.2.4		rametric measures	
		4.2.2		g upon the				of sprea		140
		242-0-2		al analysis strategy	125	5.3		ical tool		140
				Including interactions			5.3.1	Scatterp		140
				in the statistical model	126		5.3.2	Box-plo		142
			4.2.2.2	Including blocking			5.3.3	Histogra		143
				factors	127		5.3.4		rised case profiles	14 414
		4.2.3	Repeate	edly measured				plot	7.1 CF1.6 1	144
			respons	es	127		5.3.5		with SEMs plot	145
			4.2.3.1	Repeated factors and				5.3.5.1	Problems with the	1.45
				randomised factors	127			F 2 F 2	means with SEMs plot	145
			4.2.3.2	Block and dose-escalati	on			5.3.5.2	Benefits of the means	151
				designs	127	5.4	Daram	otrio one	with SEMs plot	151
			4.2.3.3	Crossover and		5.4		etric ana	tric assumptions	151 152
				dose-escalation designs	128		5.4.1	5.4.1.1	Numeric and	152
			4.2.3.4	Including interactions				3.4.1.1	continuous responses	152
				involving the repeated				5.4.1.2	Normally distributed	132
				factor	129			3.4.1.2	residuals	153
	4.3	What	to randor	nise	129			5.4.1.3	Homogeneity of	133
	4.4	How t	o randon	nise	130			5.4.1.5	variance	155
5	Statis	stical a	nalysis		132			5.4.1.4	Independence of the	133
3			uction		132			3.4.1.4	responses	158
	0.1		InVivoS	tat	133			5.4.1.5	Removal of outliers	159
				mended five-stage	100				Additivity	162
		0.1.12		tric analysis procedure	133		5.4.2	The <i>t</i> -te		163
	5.2	Summ	nary statis		135		0.1.2	5.4.2.1	The unpaired <i>t</i> -test	163
	10.000	5.2.1		tric measures of				5.4.2.2	When not to use an	
			location		135				unpaired <i>t</i> -test	165
			5.2.1.1	The true mean and the				5.4.2.3	The paired <i>t</i> -test	167
				sample mean	135			5.4.2.4	Randomisation and the	-
			5.2.1.2	The observed mean	136				paired t-test	168

5.4.3	Analysis	s of variance				5.4.6.5	Predicted group means	203
	(ANOVA	١)	168			5.4.6.6	Assumptions for	
	5.4.3.1	One-way ANOVA	169				ANCOVA	204
	5.4.3.2	Including the positive				5.4.6.7	Strategy for when the	
		control	173				independence	
	5.4.3.3	Two-way ANOVA	174				assumption does	
	5.4.3.4	Two-way vs. one-way					not hold	207
		ANOVA	176			5.4.6.8	ANCOVA and stratified	
	5.4.3.5	Dealing with missing					randomisation	208
		factor combinations	177			5.4.6.9	Change from baseline	
5.4.4	Repeate	ed measures					responses	208
	analysis		179		5.4.7	Regressi	on analysis	211
	5.4.4.1	Categorised case			5.4.8	Multiple	comparison	
		profiles plot	181			procedu	res	212
	5.4.4.2	Analysis of summary				5.4.8.1	The risk of finding	
		measures	181				false positives and false	
	5.4.4.3	Repeated measures					negatives	212
		analysis	189			5.4.8.2	Choosing the family	
	5.4.4.4	The mixed-model					of tests	214
		approach vs. the ANOVA	1-			5.4.8.3	Unadjusted tests	215
		based approach	191			5.4.8.4	Stepwise multiple	
	5.4.4.5	Advantages and					comparison procedures	3
		disadvantages of the					that control the FDR	218
		repeated measures				5.4.8.5	Simultaneous multiple	
		analysis	195				comparison procedures	3
5.4.5	Predicte	ed means from the					that control the FWE	218
	parame	tric analysis	196			5.4.8.6	Stepwise multiple	
		Least square (predicted)	)				comparison procedures	S
		means	196				based on group	
	5.4.5.2	Variability of the least					differences that control	
		square (predicted)					the FWE	222
		means	197			5.4.8.7	Stepwise-based multipl	e
	5.4.5.3	Geometric means and					comparison procedures	S
		confidence intervals	197				based on p-values that	
	5.4.5.4	Reliability of the					control the FWE	223
		predicted means	198			5.4.8.8	The gateway ANOVA	
5.4.6	Analysis	s of covariance					approach	224
	(ANCO	VA)	199			5.4.8.9	Multiple comparison	
	5.4.6.1	What is a covariate?	200				procedures in statistical	l
	5.4.6.2	Best-fit lines and					software packages	227
		predicted lines	201			5.4.8.10	Recommendations	228
	5.4.6.3	Categorised scatterplot	201	5.5	Other	useful an	alyses	228
	5.4.6.4	Predictions from			5.5.1		rametric analyses	228
		ANCOVA	202			-	When to use a	

				non-parametric test	229			6.3.3.5	Analysis of designs	
			5.5.1.2	Non-parametric tests	230				with missing factor	
		5.5.2	Testing	the difference					combinations	252
			betwee	n proportions	231	6.4	Repea	ted Mea	sures Parametric	
			5.5.2.1	Analysis procedure	232		Analys	sis modu	ile	252
			5.5.2.2	Chi-squared test	232		6.4.1	Analysi	s procedure	252
			5.5.2.3	Fisher's exact test	233		6.4.2	Worked	l example	255
		5.5.3	Surviva	l analysis	234		6.4.3	Technic	cal details	255
			5.5.3.1	The survival function	235	6.5	P-Valu	ie Adjust	ment module	258
			5.5.3.2	Comparing groups	236		6.5.1	Analysi	s procedure	259
							6.5.2	Worked	l example	259
6	Ana	lysis us	ing InViv	roStat	238	6.6	Non-F	arametr	ic Analysis module	260
	6.1	Gettir	ng started	I	238		6.6.1	Analysi	s procedure	260
		6.1.1	Data im	port	238		6.6.2	Worked	l example	262
			6.1.1.1	Single measure		6.7	Graph	ics mod	ule	262
				format	238		6.7.1	Analysi	s procedure	262
			6.1.1.2	Repeated measures			6.7.2	Exampl	e plots	263
				format	239	6.8	Power	Analysis	s module	263
		6.1.2	Importi	ing a dataset into			6.8.1	Analysi	s procedure	263
			InVivoS	Stat: Excel import	240		6.8.2	Worked	example	265
		6.1.3	Importi	ing a dataset into		6.9	Unpai	red t-tes	t Analysis module	267
			InVivoS	Stat: text file import	240		6.9.1	Analysi	s procedure	267
		6.1.4	Data ma	anagement	240		6.9.2	Worked	l example	271
		6.1.5	Running an analysis		240	6.10	Paired	t-test/w	rithin-subject	
		6.1.6	Warning	g and error messages	241		Analys	sis modu	le	272
		6.1.7	Log file		241		6.10.1	Analys	is procedure	272
		6.1.8	Exporti	ng results	241			Worke	d example	276
	6.2	Sumn	nary Stati	stics module	241	6.11	Dose-Response Analysis module			277
		6.2.1	Analysi	s procedure	242		6.11.1	Techni	ical details on	
		6.2.2	Worked	l example	243			curve f	itting	277
	6.3	Single	Measure	e Parametric			6.11.2	Fitting l	logistic curves to data	278
		Analy	sis modu	ıle	243		6.11.3	Analys	is of quantitative	
		6.3.1	Analysi	s procedure	243			assays		278
		6.3.2	Worked	l example	245		6.11.4	Analys	is procedure	279
		6.3.3		cal details	248		6.11.5	Worke	d example: a	
			6.3.3.1	Analysis of large				biologi	ical assay	281
				factorial experiments	248		6.11.6	User-d	efined equation	
			6.3.3.2	Analysis of small				option		282
				factorial experiments	248	6.12	Chi-sq	uared To	est and Fisher's	
			6.3.3.3	Analysis of experiment	S		Exact	Test mod	lule	282
				involving blocking			6.12.1	Analys	is procedure	283
				factors	249		6.12.2	Worke	d example	284
			6.3.3.4	Analysis of crossover		6.13	R-Run	ner mod	ule	285
				trials	251	6.14	Nestec	Design	Analysis module	285

	6.14.1 An	alysis procedure	286	7	Conclusion	293
	6.14.2 Wo	orked example	289			
6.15	Survival A	nalysis module	289	Glos	sary	295
	6.15.1 An	alysis procedure	289	Refe	rences	297
	6.15.2 We	orked example	291	Inde	X	303

### Introduction

Many researchers, either directly or indirectly, rely on statistical ideas when carrying out animal experiments. While some statistical tools are well known and are applied routinely, other tools are less well understood and so are less well used. The overall aim of this book is to discuss statistical methodologies that can be applied throughout the many stages of the experimental process. Researchers should be able to carry out most of the techniques described, although the advice of a professional statistician is advisable for some of the more advanced topics. Making use of these techniques will ensure that experiments are conducted in a logical and efficient way, which should result in reliable and reproducible decisions.

The particular types of study addressed in this book, as the title suggests, are studies involving animals. We attempt to cover all of the statistical tools that the animal researcher should use to run successful studies. Of course many of the problems faced by the animal researcher are common to other disciplines, and hence the ideas contained within this book can be applied to other areas. It should be noted that certain topics described in the text have been simplified to allow non-statisticians to apply the ideas without professional statistical support. Such pragmatic descriptions, while simplifying the technical details, are not universal and will not be applicable in all scientific disciplines.

There has been much interest in the use of statistics in animal research, in particular in the application of the 3Rs, replacement, reduction and refinement, as described by Russell and Burch (1959):

Every time any particle of statistical method is properly used, fewer animals are employed than would otherwise have been necessary.

Many authors have since highlighted how important the use of good experimental design is when conducting animal experiments; see Festing (1994, 2003a, 2003b) and the references contained within. Some of the more practical, as well as statistical, aspects of experimental design and statistics when applying the 3Rs are described in the book by Festing et al. (2002). There have also been surveys into the use of statistics in refereed journals; see McCance (1995) and more recently Kilkenny et al. (2009). The latter draws attention to some of the mistakes that can be made by researchers when designing and analysing animal experiments. The reliability of the reporting of animal experiments has been considered in, for example, Macleod et al. (2009) and Rooke et al. (2011). These articles highlight that papers describing experiments that do not employ suitable randomisation techniques and/or blinding may contain biased results.

The main goal of this text is to demonstrate how statistics can aid the reduction and refinement of animal studies. The efficient use of statistics, both in terms of complex experimental design and powerful statistical analysis, can reduce the number of animals required. Statistics can also help the researcher understand the processes that underpin the animal model and help identify factors that are influencing the experimental results. Such an understanding will inevitably lead to a refinement in the experimental process and a reduction in the total number of animals used.

Statistics, as a discipline, provides researchers with tools to help them arrive at valid conclusions. However, statistics, along with the application of some common sense, can also increase the understanding of the animal model through the application of graphical and mathematical techniques. For example, graphical tools play an important role in helping the researcher understand the effect of the features of the experimental design and also uncover overall patterns present in the data. The application of a formal statistical test, without first investigating the data graphically, can lead to the researcher drawing incorrect conclusions from the data. Consider the following real-life case study, which used graphical, as well as statistical, tools. If a conventional statistical analysis had been carried out, without first investigating all of the information gathered within the experiment, then the conclusions would have been misleading.

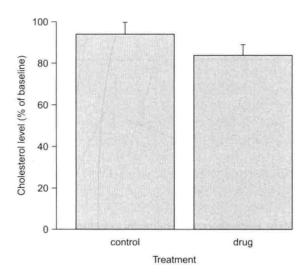
### Example 1.1: Reducing blood cholesterol levels in mice

A scientist wanted to test the hypothesis that a novel compound had a beneficial effect on reducing high-density lipoprotein (HDL) cholesterol levels in a transgenic C57Bl/6J strain of mice. A blood sample was taken pre-treatment and the baseline cholesterol level for each animal measured. The mice were then randomised to either the drug treatment group or the control group and dosed with either the drug treatment or vehicle twice daily for two weeks. At the end of this period, a terminal blood sample was taken and the HDL cholesterol level measured.

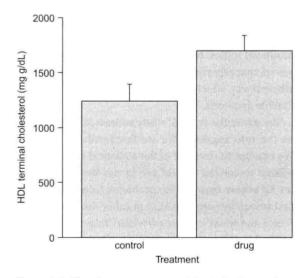
As the scientist wanted to make use of the baseline information in the statistical analysis, it was decided that the percentage change from baseline would be a suitable response to investigate. This would, the scientist hoped, effectively remove the animal-to-animal differences by normalising to the baseline level. While there was evidence of a decrease in HDL cholesterol level in the group of animals administered the drug treatment (a 20% decrease from baseline in the drug treatment group compared to a 10% decrease in the control group) this was not deemed statistically significant using an unpaired t-test (p = 0.191). A means with standard errors of the mean (SEMs) plot of the data (see Section 5.3.5) is presented in Figure 1.1.

As a follow-up the scientist also analysed the terminal HDL cholesterol level. From this analysis it appeared that there was a statistically significant increase in cholesterol level in the drugtreated group compared to the control. A plot of the means with SEMs of the terminal HDL cholesterol level is presented in Figure 1.2.

Based on the results of this experiment, should we conclude the drug increases cholesterol levels? And why did the two analyses give such different conclusions? These questions can be answered



**Figure 1.1.** Plot of treatment means with standard errors for the percentage of baseline cholesterol response for Example 1.1.



**Figure 1.2.** Plot of treatment means with standard errors for the terminal HDL cholesterol for Example 1.1.

by a simple scatterplot of the measured HDL cholesterol levels. If we plot terminal vs. baseline HDL cholesterol levels, an underlying problem with the experiment becomes clear. The scatterplot is presented in Figure 1.3.

From Figure 1.3 it can be seen that there are two distinct groupings along the X-axis. The plot reveals that, in terms of the HDL baseline cholesterol level, the animals belong to one of two