

Scholars'
Press

Debasish Ghosh

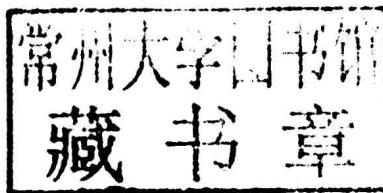
Molecular Cell Biology of Potato yellow dwarf virus N and P protein

Molecular and cellular characterization of PYDV nucleocapsid and phosphoprotein; Understanding the biology of PYDV

Debasish Ghosh

Molecular Cell Biology of Potato yellow dwarf virus N and P protein

**Molecular and cellular characterization of PYDV
nucleocapsid and phosphoprotein; Understanding
the biology of PYDV**



Scholar's Press

Impressum / Imprint

Bibliografische Information der Deutschen Nationalbibliothek: Die Deutsche Nationalbibliothek verzeichnet diese Publikation in der Deutschen Nationalbibliografie; detaillierte bibliografische Daten sind im Internet über <http://dnb.d-nb.de> abrufbar.

Alle in diesem Buch genannten Marken und Produktnamen unterliegen warenzeichen-, marken- oder patentrechtlichem Schutz bzw. sind Warenzeichen oder eingetragene Warenzeichen der jeweiligen Inhaber. Die Wiedergabe von Marken, Produktnamen, Gebrauchsnamen, Handelsnamen, Warenbezeichnungen u.s.w. in diesem Werk berechtigt auch ohne besondere Kennzeichnung nicht zu der Annahme, dass solche Namen im Sinne der Warenzeichen- und Markenschutzgesetzgebung als frei zu betrachten wären und daher von jedermann benutzt werden dürften.

Bibliographic information published by the Deutsche Nationalbibliothek: The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available in the Internet at <http://dnb.d-nb.de>.

Any brand names and product names mentioned in this book are subject to trademark, brand or patent protection and are trademarks or registered trademarks of their respective holders. The use of brand names, product names, common names, trade names, product descriptions etc. even without a particular marking in this works is in no way to be construed to mean that such names may be regarded as unrestricted in respect of trademark and brand protection legislation and could thus be used by anyone.

Coverbild / Cover image: www.ingimage.com

Verlag / Publisher:

Scholar's Press

ist ein Imprint der / is a trademark of

OmniScriptum GmbH & Co. KG

Heinrich-Böcking-Str. 6-8, 66121 Saarbrücken, Deutschland / Germany

Email: info@scholars-press.com

Herstellung: siehe letzte Seite /

Printed at: see last page

ISBN: 978-3-639-66426-3

Zugl. / Approved by: Lexington, University of Kentucky, Diss., 2008

Copyright © 2014 OmniScriptum GmbH & Co. KG

Alle Rechte vorbehalten. / All rights reserved. Saarbrücken 2014

Debasish Ghosh

Molecular Cell Biology of Potato yellow dwarf virus N and P protein

Molecular Cell Biology of Potato yellow dwarf virus N and P protein

Dr. Debasish Ghosh M.Sc., Ph.D.

Dedicated to the loving memory of my grandfather, Late Mr. Chitta Ranjan Ghose
and my mother, Late Mrs. Dipti Ghose

MOLECULAR CELL BIOLOGY OF THE N (NUCLEOCAPSID) AND P (PHOSPHOPROTEIN) PROTEIN OF THE SYDV (*POTATO YELLOW DWARF VIRUS – SANGUINOLENTA STRAIN*)

Potato yellow dwarf virus (PYDV) is the type member of the genus *Nucleorhabdovirus*. The virus replicates in the nuclei of infected cells and mature virions accumulate in the perinuclear space after viral cores bud through the inner nuclear membrane. The virus was first described as an extremely destructive pathogen of potato (*Solanum tuberosum*) and other members of family *Solanaceae*. There are two different strains of PYDV based on their insect-vector specificity, namely SYDV (*sanguinolenta* strain) and CYDV (*constricta* strain). PYDV is considered a model system to study virus-vector relationship, particularly for agriculturally harmful rhabdoviruses. However, very little is known about the molecular aspects and cell biology of PYDV. Preliminary studies showed that infection of transgenic *Nicotiana benthamiana* plants that constitutively express GFP targeted to endomembranes with SYDV and SYNV (*Sonchus yellow net virus*, another member of genus *Nucleorhabdovirus*) results in increased accumulation of GFP and membrane within the infected nuclei, though the pattern of GFP accumulation is completely different for the two viruses. GFP accumulation was found mainly in the external and internal loci of the nucleus in SYDV-infected cells, where as, in the case of SYNV infection, the GFP accumulation was scattered throughout the nucleus of the infected cell. Molecular characterization of SYDV was undertaken to better understand the cellular difference between these two members of *Nucleorhabdoviruses*. This dissertation describes the determination of the complete nucleotide and ORF (open reading frame) sequences of N (nucleocapsid) and P (Phosphoprotein) gene of SYDV from cDNA clones of both viral genomic and messenger RNAs. Analyses of sequence showed that SYDV-N mRNA contains an 11 nucleotide (nt) untranslated region followed by a 1416 nt ORF encoding a 472 amino acid (aa) protein and P-mRNA contains an 18 nt 5' untranslated region followed by 840 nt ORF encoding a 280 aa protein. Characterization of SYDV-N and P protein using bioinformatic algorithms predict basic hydrophilic and coiled coil regions that may posses the putative nuclear localization signal and protein-protein interaction domain, respectively. Comparison of the SYDV-N ORF with orthologous regions from other plant and animal rhabdoviruses showed

statistically significant identity. Phylogenetic analysis based on consensus N-ORFs placed SYDV into the same group with other *Nucleorhabdoviruses*. Localization studies of SYDV-N and P protein as autofluorescent protein fusions revealed that both proteins are exclusively nuclear localized. Taken together, this dissertation reports a detailed analysis of the biology of SYDV-N and P protein at the molecular and cellular level for the first time towards the long term goal to characterize the entire SYDV genome and to better understand SYDV-host interaction.

Keywords: rhabdovirus, SYDV, SYNV, nucleus, transgenic “16c” plant,

Dr. Debasish Ghosh M.Sc., Ph.D.

09/09/2014

ACKNOWLEDGEMENTS

The following book, while an individual work, benefited from the insights and direction of several people. First, my Dissertation Co-Chairs, Dr. Judith A. Lesnaw and Dr. Michael M. Goodin, exemplifies the high quality scholarship to which I aspire. In addition, they provided timely and instructive comments and evaluation at every stage of the dissertation process, allowing me to complete this project on schedule. Next, I wish to thank the complete Dissertation Committee, and outside reader, respectively: Dr. Randal A. Voss, Dr. Peter M. Mirabito, Dr. Thomas M. Chambers and Dr. Bruce Webb. Each individual provided insights that guided and challenged my thinking, substantially improving the finished product. I want to express my gratitude to Dr. Brian C. Rymond, Director of Graduate Studies (Biology Department, University of Kentucky, Lexington Kentucky, USA), Dr. Sheldon Steiner, Chair (Biology Department, University of Kentucky, Lexington Kentucky, USA) and Dr. David Smith, Chair (Department of Plant Pathology, University of Kentucky, Lexington Kentucky, USA) for their valuable advice and encouragement throughout the process. I must acknowledge and thank Dr. Renyuan Wang, who always been helpful to me and making sure I stay on track and sane. I also thank all my friends and well-wishers in the Biology and Plant Pathology department of the University of Kentucky for their continuous support.

In addition to the technical and instrumental assistance above, I received equally important assistance from my family and friend. I want to give special thanks to my uncle, Dr. Bidyut Ghosh, Scientific Manager, M.D. Anderson Cancer Research Center, Houston Texas, USA who always inspired me to achieve my educational dream and be there for me. My wife, Sourita, provided on-going moral support throughout the dissertation process, as well as technical assistance critical for completing the project in a timely manner.

My grandmother (s) (Mrs. Latika Ghose, Late Mrs. Bela Rani Ghosh), father (Mr. Jitesh Kanti Ghosh), brother (Mr. Subhasish Ghosh), aunt (Mrs. Samapti Ghosh, Mrs. Sutapa Ghosh), mother in law (Mrs. Nandita Das) and brother in law (Mr. Shubhasish Das) instilled in me, from an early age, the desire and skills to obtain the Ph.D. I must thank my friends (Sharon Yelton, Robbie Brooks, Dr. Anthony Clerk, George Chaffins, Sujit Dutta, Sushanta Saha, Shyamal Saha, Samarendra Saha) for their immense help to successfully finish this work. Thanks to Dr. Govinda Chandra Sadhukhan (Professor, Vidyasagar College, Calcutta, India), Dr. Sameer Banerjee, Dr. Dilip Nanda, and Dr. Dinendra Roychowdhury (Professor, Department of Zoology, University of Calcutta, India) for their precious guidance during my bachelors and masters days. Last but not the least, I am grateful to Dr. E. Patrick Heist, Mr. Shane Baker for believing in me and everyone from the FSI/ WTD family for their continuous support. Finally, I wish to thank the respondents of my study (who remain anonymous for confidentiality purposes). Their comments and insights created an informative and interesting project with opportunities for future work.

Table of Contents

Acknowledgements	iii
List of Tables.....	viii
List of Figures.....	ix
Chapter 1.....	1
1. Comprehensive literature review of rhabdoviruses.....	1
1.1 Introduction.....	1
1.2 Genomic organization of plant rhabdoviruses.....	2
1.3 Properties of rhabdoviral Nucleocapsid (N) and Phosphoprotein (P)...4	4
1.4 Biology of N and P protein in brief.....	6
1.5 Comparison of the replication cycle of <i>Nucleo-</i> and <i>Cytorhabdoviruses</i>	8
1.6 Vector specificity of plant rhabdoviruses.....	9
1.7 <i>Potato yellow dwarf virus</i> (PYDV) – type species of genus Nucleorhabdoviruses.....	10
1.8 Vector specificity, host range and strains of PYDV.....	10
1.9 Geographical distribution of PYDV.....	11
1.10 Serological relationship between PYDV strains.....	12
Chapter 2.....	27
2. Preliminary experiments: The infection pattern of SYDV in host cells.....27	27
2.1 Introduction.....	27
2.2 Plant material, growth conditions and virus inoculation procedures....28	28

2.3: SYDV infection induce increased accumulation of GFP in infected nuclei of Nicotiana benthamiana cells.....	29
2.4: GFP accumulates in both external and internal loci of nuclei in SYDV-infected tissue.....	30
2.5: Virus specific pattern of GFP and membrane accumulation in SYDV and SYNV-infected nuclei.....	30
2.6: Enlargement of nuclei occurs in only rhabdovirus-infected tissues.....	31
2.7 Hypothesis.....	31
Specific Aims.....	32
2.8 Significance.....	32
Chapter 3	40
3. Cloning and Characterization of SYDV N and P mRNA.....	40
3.1 Introduction.....	40
Materials and Methods.....	40
3.2 Virus maintenance and purification.....	40
3.3 SDS-PAGE analysis.....	42
3.4 RNA extraction and cloning of SYDV-specific PCR product.....	43
3.5 Cloning of 5' and 3' termini of SYDV Nucleocapsid (N) and Phosphoprotein (P) mRNA.....	43
3.6 Peptide sequencing and sequence analysis.....	44
3.7 DNA sequencing and sequence analysis.....	45
3.8 Northern blot hybridization analysis.....	46
3.9 Phylogenetic Analysis.....	47

3.10 Deposition of sequence data.....	47
3.11 Construction of pSITE expression vectors for <i>in planta</i> subcellular Localization.....	48
3.12 Cloning of SYDV-N mRNA.....	48
3.13 Cloning of SYDV-P mRNA.....	51
3.14 Structure, characterization and sequence analysis of SYDV-N mRNA..	52
3.15 Structure, characterization and sequence analysis of SYDV-P mRNA...	54
3.16 Phylogenetic relationship of SYDV with other <i>Rhabdoviruses</i> based on consensus sequences of the N ORF.....	57
3.17 Subcellular localization studies of SYDV-N and P protein.....	58
3.18 Time course systemic infection pattern of SYDV in <i>Nicotiana benthamiana</i>	60
3.19 Primers used in this study.....	62
Chapter 4.....	81
4.1 Discussion.....	81
4.2 Proposed strategy to complete the sequencing of SYDV genome.....	85
Appendices.....	87
5. Complete organization of all SYDV cDNAs, PCR products and northern blot hybridization data.....	87
5.1 Amplification of SYDV-N mRNA fragment.....	87
5.2 5' and 3' RACE of SYDV-N mRNA.....	88
5.3 Amplification of SYDV-N-G fragment.....	90
5.4 Amplification of SYDV-P mRNA.....	90

5.5 5' and 3' RACE of SYDV-P mRNA.....	91
5.6 Amplification of the region between N and P gene (Intergenic-region).....	92
5.7 Amplification of SYDV-G gene.....	94
5.8 Amplification of SYDV-L gene fragment.....	95
5.9 Amplification of SYDV-N-L fragment.....	96
5.10 Amplification of SYDV-specific 5.0 kb fragment.....	97
5.11 Amplification of SYDV-G-L fragment.....	98
References/ Bibliography.....	119

List of Tables

Chapter 1

Table 1.1: Examples of animal and-plant infecting rhabdoviruses.....	15
Table 1.2: Nuclear localization signal (NLS) of N and P protein of plant <i>Nucleorhabdoviruses</i>	16
Table 1.3: List of plant rhabdoviruses (<i>Nucleo-</i> and <i>Cytorhabdoviruses</i>) and their specific insect vector.....	17
Table 1.4: Comparison of gene junction sequences between plant and animal rhabdoviruses.....	18

Chapter 2

Table 2.1: SYDV cage condition in greenhouse.....	34
---	----

Chapter 3

Table 3.1: Degenerate primer sequences of SYDV-N and G peptide fragments.....	64
---	----

Appendices

Table 5.1: Complete chart of SYDV PCR product and cloned fragments.....	99
---	----

List of Figures

Chapter 1

Figure 1.1: Generalized morphology of rhabdoviruses.....	19
Figure 1.2: Comparison of negative-sense genomic organization between plant (SYNV) and animal (VSV) rhabdovirus.....	20
Figure 1.3: Structure of VSV-P protein.....	21
Figure 1.4: Comparison of multiplication cycle between <i>Nucleo-</i> and <i>Cytothabdoviruses</i> in the host cell.....	22
Figure 1.5: Symptoms of SYDV in infected leaves.....	23
Figure 1.6: Western immunoblots demonstrating serological relationship between the SYDV and CYDV.....	24
Figure 1.7: Serological western immunoblot analysis of SYDV and CYDV using different source of anti-SYDV and CYDV antibodies.....	25

Chapter 2

Figure 2.1: Detection of GFP fluorescence in SYDV infected “16c” <i>Nicotiana benthamiana</i> tissue.....	35
Figure 2.2: Increased accumulation of membrane-associated GFP in the nuclei of SYDV-infected cells.....	37
Figure 2.3: Comparison of nuclear area of <i>Nucleorhabdovirus</i> , SYDV and SYNV with cytoplasm replicating viruses e.g. TRV, TEV and INSV.....	39

Chapter 3

Figure 3.1: Flow chart of SYDV purification.....	65
Figure 3.2: Detection of purified SYDV from <i>Nicotiana benthamiana</i> leaf tissue.....	66

Figure 3.3: Strategy for cloning SYDV-N and P mRNA using gene-specific degenerate primers.....	67
Figure 3.4: Detection of N and P mRNA transcript in SYDV infected <i>Nicotiana benthamiana</i> by northern hybridization.....	69
Figure 3.5: Strategy for cloning 5' and 3' ends of SYDV-N and P mRNA.....	70
Figure 3.6: Nucleotide and deduced amino acid sequence of the SYDV-N mRNA.....	71
Figure 3.7: Nucleotide and deduced amino acid sequence of the SYDV-P mRNA.....	72
Figure 3.8: Hydropathy profile of SYDV-N and P protein.....	73
Figure 3.9: Prediction of coiled coil region in SYDV-N and P protein.....	74
Figure 3.10: Neighbor-joining phylogenetic tree using the complete SYDV-N amino acid sequence and orthologs from selected plant and animal rhabdoviruses.....	75
Figure 3.11: Confocal micrographs showing the subcellular localization of SYDV-N and P protein in <i>Nicotiana benthamiana</i> cells.....	76
Figure 3.12: Confocal micrographs showing the subcellular localization of SYDV-N and P protein in mock and SYDV-infected “16c” <i>Nicotiana benthamiana</i> cells.....	77
Figure 3.13: Time course virus infection in SYDV infected <i>Nicotiana</i> leaves.....	79
Appendices	
Figure 5.1: Amplification of SYDV-N gene.....	100
Figure 5.2: Northern hybridization showing the validation of SYDV-N clones.....	101