

STUDIES IN INTERFACE SCIENCE

25

SERIES

EDITORS:

- D. MÖBIUS
- R. MILLER

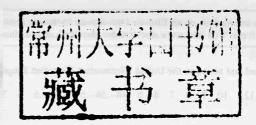
Stable Nanoemulsions

Self-Assembly in Nature and Nanomedicine

Joseph D'Arrigo

Stable Nanoemulsions: Self-Assembly in Nature and Nanomedicine

Joseph D'Arrigo





AMSTERDAM • BOSTON • HEIDELBERG • LONDON NEW YORK • OXFORD • PARIS • SAN DIEGO SAN FRANCISCO • SINGAPORE • SYDNEY • TOKYO Elsevier Radarweg 29, PO Box 211, 1000 AE Amsterdam, The Netherlands The Boulevard, Langford Lane, Kidlington, Oxford OX5 1GB, UK

Copyright @ 2011 Elsevier B.V. All rights reserved.

No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording, or otherwise, without the prior written permission of the publisher.

Permissions may be sought directly from Elsevier's Science & Technology Rights Department in Oxford, UK: phone: (+44) 1865 843830, fax: (+44) 1865 853333, E-mail: permissions@elsevier.com. You may also complete your request online via the Elsevier homepage (http://elsevier.com), by selecting "Support Contact" then "Copyright and Permission" and then "Obtaining Permissions."

Library of Congress Cataloging-in-Publication Data A catalog record for this book is available from the Library of Congress

British Library Cataloguing-in-Publication Data A catalogue record for this book is available from the British Library

ISBN: 978-0-444-53798-0

ISSN: 1383-7303

For information on all Elsevier Publications visit our Web site: www.elsevierdirect.com

Printed and bound in the United Kingdom

11 12 10 9 8 7 6 5 4 3 2

Working together to grow libraries in developing countries

www.elsevier.com | www.bookaid.org | www.sabre.org

Stable Nanoemulsions

Studies in Interface Science

- Vol. 1 Dynamics of Adsorption at Liquid Interfaces. *Theory, Experiment, Application*. By S.S. Dukhin, G. Kretzschmar and R. Miller
- Vol. 2 An Introduction to Dynamics of Colloids. By J.K.G. Dhont
- Vol. 3 Interfacial Tensiometry. By A.I. Rusanov and V.A. Prokhorov
- Vol. 4 New Developments in Construction and Functions of Organic Thin Films. Edited by T. Kajiyama and M. Aizawa
- Vol. 5 Foam and Foam Films. By D. Exerowa and P.M. Kruglyakov
- Vol. 6 Drops and Bubbles in Interfacial Research. Edited by D. Möbius and R. Miller
- Vol. 7 Proteins at Liquid Interfaces. Edited by D. Möbius and R. Miller
- Vol. 8 Dynamic Surface Tensiometry in Medicine. By V.M. Kazakov, O.V. Sinyachenko, V.B. Fainerman, U. Pison and R. Miller
- **Vol. 9** Hydrophile-Lipophile Balance of Surfactants and Solid Particles. *Physicochemical Aspects and Applications.* By P.M. Kruglyakov
- Vol. 10 Particles at Fluid Interfaces and Membranes. Attachment of Colloid Particles and Proteins to Interfaces and Formation of Two-Dimensional Arrays. By P.A. Kralchevsky and K. Nagayama
- Vol. 11 Novel Methods to Study Interfacial Layers. By D. Möbius and R. Miller
- Vol. 12 Colloid and Surface Chemistry. By E.D. Shchukin, A.V. Pertsov, E.A. Amelina and A.S. Zelenev
- Vol. 13 Surfactants: Chemistry, Interfacial Properties, Applications. Edited by V.B. Fainerman, D. Möbius and R. Miller
- Vol. 14 Complex Wave Dynamics on Thin Films. By H.-C. Chang and E.A. Demekhin
- Vol. 15 Ultrasound for Characterizing Colloids. Particle Sizing, Zeta Potential, Rheology. By A.S. Dukhin and P.J. Goetz
- Vol. 16 Organized Monolayers and Assemblies: Structure, Processes and Function. Edited by D. Möbius and R. Miller
- Vol. 17 Introduction to Molecular Microsimulation of Colloidal Dispersions. By A. Satoh
- Vol. 18 Transport Mediated by Electrified Interfaces: Studies in the linear, non-linear and far from equilibrium regimes. By R.C. Srivastava and R.P. Rastogi
- Vol. 19 Stable Gas-in-Liquid Emulsions: Production in Natural Waters and Artificial Media. Second Edition. By J.S. D'Arrigo
- Vol. 20 Interfacial Separation of Particles. By S. Lu, R.J. Pugh and E. ForssbergVol. 21 Surface Activity in Drug Action. By R.C. Srivastava and A.N. Nagappa
- Vol. 22 Electrorheological Fluids: The Non-aqueous Suspensions. T. Hao
- Vol. 23 Nanocomposite Structures and Dispersions Science and Nanotechnology - Fundamental Principles and Colloidal Particles. Edited by: I. Capek
- Vol. 24 Characterization of Liquids, Nano- and Microparticulates, and Porous Bodies Using Ultrasound. Second Edition. By Andrei S. Dukhin and Philip J. Goetz

To Sachie, Paul, and Marie

With the growth of complex system science and the expansion of nanotechnology, there is increased need to distinguish between two related mechanisms, "self-organization" and "self-assembly," occurring in physical and biological systems. Basically, as pointed out in a recent issue of the journal *Complexity*, self-organization is a nonequilibrium process; in contrast, self-assembly leads toward equilibrium. Nevertheless, self-organization and self-assembly are regularly used interchangeably, as both explain how collective order is developed from dynamic small-scale interactions [J.D. Halley, D.A. Winkler, Consistent concepts of self-organization and self-assembly, *Complexity* 14 (2008) 10–17]. Hence, in this book, all use of the term "self-assembly" (which some chemists classify as either static or dynamic [*Science* 295 (2002) 2418–2421]) is here only intended within a "dynamic" sense; specifically, "dynamic" self-assembly corresponds to what biologists understand as self-organization [*Complexity* 14 (2008) 10–17].

Stable gas nanoemulsions, existing in natural waters, represent self-assembled coated microbubbles (also known as "gas-in-liquid emulsions"). Similarly, in certain artificial media (namely, lipid dispersions modeled from natural microbubbles), stable nanoemulsions are also able to self-assemble (self-organize) readily. (Consequently, the first (1986) and expanded second (2003) editions of a related earlier book were entitled Stable Gas-in-Liquid Emulsions (with the subtitle Production in Natural Waters and Artificial Media). Yet, this much-expanded current book, that is, 12 chapters longer than the 2003 monograph, is more inclusive in its scope and accordingly entitled Stable Nanoemulsions.) In this specific case, the nanoemulsions comprise both "lipid-coated microbubbles (LCM)" (i.e., the gas-emulsion subpopulation) and "related lipid nanoparticles" (i.e., a particle-like subpopulation including mostly colloidal liquid crystals). Various measurements and other published findings indicate that the LCM's structural characteristics help drive and govern a continual and reversible (molecular and/ or supramolecular) lipid interchange, with the nanoparticle subpopulation, in these self-assembling lipid nanoemulsions.

The term "LCM" is utilized, in this multidisciplinary book, to accurately trace the chronological development (and functional conversion) of the "LCM/nanoparticle-derived" colloidal system: (1) from its (modeling after natural microbubble surfactant and) early biomedical application as an imaging agent (in Chapters 1–12, which focus mainly on the less numerous micron-scale colloidal species) into (2) the later adaptation of exactly the same mixed-lipid (e.g., Filmix®) colloidal system (in Chapters 13–27, which

focus more upon the vastly more numerous nanoscale colloidal species) for nanomedical application as a (LCM/nanoparticle-derived) drug-delivery vehicle. In addition, as explained in the chapters, newer models of several selected particle-size-analysis instruments have revealed that approximately 90% of these LCM/nanoparticle-derived colloidal species are actually smaller than 200 nm in diameter, while over 99% of the same mixed-lipid colloidal species (detectable via optical-particle-counter data) are documented to be smaller than 300 nm in diameter.

In this book, much experimental data are reviewed in detail and updated, along with the relevant current literature, which collectively demonstrate that this type of stable lipid nanoemulsion (upon intravenous injection) is capable of "active targeting" to tumors, and to certain lesion sites, via the process of receptor-mediated endocytosis. Hence, this LCM/nanoparticle-derived lipid formulation has been used successfully, in animals, as a drug-delivery agent that actively targets antineoplastic drug (e.g., paclitaxel) against tumor cells that commonly overexpress certain surface receptors, which fall within the category known as "lipoprotein receptors." Moreover, this LCM/nanoparticle-derived lipid nanoemulsion contains no phospholipids, proteins, peptides, and carbohydrates, and no chemical modification of the drug (paclitaxel) is required. Hence, this category of parenteral lipid nanoemulsion avoids various past problems reported for earlier versions of (actively) targeted drug-delivery agents utilizing such lipoprotein-receptor-mediated endocytic pathway(s). (Consequently, a human clinical trial is now in preparation, by a pharmaceutical company, for targeted drug delivery of paclitaxel to tumors in patients using an LCM/nanoparticle-derived drug-delivery agent.) In addition, as detailed in later chapters of the book, there are several noncancerous lesion/injury sites involving certain proliferative processes (e.g., atherosclerosis) which include overexpression of cellsurface lipoprotein receptors. Therefore, the scope of potential clinical trials, which are applicable to the pharmaceutical category referred to as LCM/nanoparticle-derived lipid nanoemulsions, can now include the targeted chemotherapy of hyperproliferative diseases, for example, atherosclerosis and CNS-injury sites. In these last few chapters, several sections detail how one particular lipidnanoemulsion agent (Filmix®) in this pharmaceutical (LCM-related) category, as well as a few other closely related protein-free parenteral lipid nanoemulsions, accordingly exhibit much (literature-supported) potential for providing "actively targeted" chemotherapy of atherosclerotic lesions in human subjects. (Such targeted chemotherapy is also in harmony with goals of the current U.S. National Nanotechnology Initiative, which include nanomedical approaches to drug delivery that focus on developing nanoscale particles (or macromolecules) to improve drug bioavailability, that is, often using targeted nanoparticles for delivering drugs with cell precision and less side effects.)

The book has been organized into six parts. Parts I and II (Chapters 1–8) describe coated microbubbles in the biosphere, as well as various biochemical, geochemical, surface, and structural properties of natural microbubble

surfactant. Next, artificial LCM and related lipid nanoparticles are described in Part III (Chapters 9–11), while their utilization in biomedical studies with animals is examined in detail in Part IV (Chapters 12–15).

Parts V and VI consist of completely new chapters (i.e., Chapters 16–27) that contribute to a strong nanomedicine focus. These 12 chapters further analyze and characterize this type of self-assembling mixed-lipid nanoemulsion, regarding LCM and especially its predominant mixed-lipid nanoparticle subpopulation. In addition, recent clinical studies with related parenteral (lipid) nanoemulsions are described; this limited clinical review provides added understanding of the development path leading to the human clinical trials—evaluating these parenteral lipid nanoemulsions as new, (actively) targeted, drug-delivery agents. Finally, throughout Parts V and VI, extensive cross-references to the earlier chapters are provided in the text. Furthermore, over 500 new literature references have been added by Parts V and VI, many of which are very recent.

The underlying chemical and biomedical principles covered in each chapter are presented in sufficient detail for this book to be useful to all interested readers worldwide with a working knowledge of chemistry, physics, and biology. Accordingly, the level of readership is intended to include graduate students, researchers, and professional people from widely varying fields. Furthermore, due to the many current and potential applications of stable lipid nanoemulsions, the appropriate readership of this book is likely to be found in industry, universities, government laboratories, and clinical facilities alike.

Thanks are due to the following colleagues for their collaboration on some of the original investigations described in this book and/or their generous help with various experimental measurements: Elisa Barbarese, William Barker, J. Howard Bradbury, Kai-Fei Chang, Stephanie A. Ching, Michael A. Davis, John F. Dunne, Donald C. Grant, Richard J. Guillory, Brendon C. Hammer, Shih-Yieh Ho, Toyoko Imae, Jacob N. Israelachvili, Inam U. Kureshi, Kathleen M. Nellis, Barry W. Ninham, Noboru Oishi, Richard M. Pashley, Neil S. Reimer, P. Scott Rice, Cesareo Saiz-Jimenez, Richard H. Simon, Kent Smith, Candra Smith-Slatas, Charles S. Springer, Ourai Sutiwatananiti, and Linda Vaught. Finally other acknowledgments, in addition to those appearing in the chapters, include permission for using quoted material appearing on p. 15, Copyright[©] 1981 by the AAAS; p. 26, Copyright[©] 1972 by the ASME; pp. 9, 12, 18, and 98, Copyright 1975, 1978, 1978, and 1974, respectively, by the Pergamon Press, Ltd.; p. 271, Copyright[©] 1973 by Springer-Verlag; pp. 271-272, Copyright[©] 1993 by the American Chemical Society; and the reprinting of Figure 12.1 on p. 216, Copyright[©] 1991 by Sage Publications, Inc. Joseph S. D'Arrigo

Contents

T.	Contents)
	sal romagnal murobundle lesidues le sei und Agartise Povoler	
	1.2 Adiperion of Hillearth Agreeus and Extrems for Use with the	
	3.2 Constituents to militario deliberation traits abbyinks E1	
Pre	ace / Ebinia lipe ainouna line	vii
		VII
	rt I	
Na	itural Coated Microbubbles in the Biosphere	1
1	0	
aa'	Occurrence of Dilute Gas-in-Liquid Emulsions in Natural Waters	2
	Natural vvaters	3
	1.1 Practical Importance of Stable Microbubbles	3
	1.1.1 Hydrodynamic Cavitation, Hydraulic and	
	Ocean Engineering	3
	1.1.2 Acoustic Cavitation	5
	1.1.3 Waste-Water Treatment: Microflotation	7
	1.1.4 Marine Biology, Chemical Oceanography 1.1.5 Meteorology	8
	1.2 Background Observations	9
	1.2.1 Problems with the Crevice Model for Bubble Nuclei	11
	1.2.2 Reduction of Gaseous Diffusion Across the Air/Water	. 11
	Interface by Selected Surfactant Monolayers	12
	1.3 Demonstration of Film-Stabilized Microbubbles in Fresh Water	15
	1.3.1 Acoustical Measurements	15
	1.3.2 Light-Scattering Measurements	19
	1.3.3 Gas-Diffusion Experiments	21
	1.4 Demonstration of Film-Stabilized Microbubbles in Sea Water	22
	1.4.1 Acoustical Measurements	22
	1.4.2 Light-Scattering Measurements	23
	1.4.3 Photographic Identification	23
2.	Early Work with Aqueous Carbohydrate Gels	29
	2.1 Development of the Agarose-Gel Method for Monitoring	
	Bubble Formation	29
	2.2 Results from Dilute Electrolyte Additions and pH Changes in	
	Agarose Gels	33
	2.3 Results from Concentrated Electrolyte Additions and	
	1% Phenol in Agarose Gels	34
	2.4 Detailed Comparison with Published Data in the	
	Physicochemical Literature for Salting Out of Identified Nonionic Surfactants	A.,
	2.5 Concluding Remarks	41
	- Continue of the state of the	4.3

3.2 Adaptation of (Filtered) Aqueous Soil Extracts for Use with the Agarose Gel Method 3.3 Ninhydrin Effect on Bubble Formation in Commercial Agarose and Aqueous Soil Extracts 3.4 Photochemical Experiments using Methylene Blue 3.5 2-Hydroxy-5-Nitrobenzyl Bromide Experiments 3.6 Conclusions 4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	46 47 48 50 52 54 55 55 55
Agarose Gel Method 3.3 Ninhydrin Effect on Bubble Formation in Commercial Agarose and Aqueous Soil Extracts 3.4 Photochemical Experiments using Methylene Blue 3.5 2-Hydroxy-5-Nitrobenzyl Bromide Experiments 3.6 Conclusions 4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	48 50 52 54 55 55
and Aqueous Soil Extracts 3.4 Photochemical Experiments using Methylene Blue 3.5 2-Hydroxy-5-Nitrobenzyl Bromide Experiments 3.6 Conclusions 4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	50 52 54 55 55
3.4 Photochemical Experiments using Methylene Blue 3.5 2-Hydroxy-5-Nitrobenzyl Bromide Experiments 3.6 Conclusions 4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	50 52 54 55 55
3.5 2-Hydroxy-5-Nitrobenzyl Bromide Experiments 3.6 Conclusions 4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	52 54 55 55
3.6 Conclusions4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	54 55 55
4. Characteristic Glycopeptide Fraction of Natural Microbubble Surfactant	55 55
Microbubble Surfactant 5	55
	55
4.1 Applytical Mothoda	55
4.1 Analytical Methods	
4.1.1 Isolation of Microbubble Glycopeptide Surfactant from	
	:6
	57
4.1.5 Carbohydrate Analyses of Partially Purified Glycopeptide	58
	61
4.1.6 Sephadex Column Chromatography of Dansylated	
	54
	66
	69 69
4.2.2 Amino Acid Composition of Microbubble Glycopeptide	
	69
	69
4.2.4 HPLC Determination of Carbohydrate Content 4.2.5 Gel-Filtration Column Chromatography: Determination	73
	75
4.3 Review of Natural-Product Literature and Possible	
Animal Sources of the Glycopeptide Fraction	
of Microbubble Surfactant	77
0	79
Part II	
Physicochemical Properties of Natural	
Microbubble Surfactant 8	1
5. Ecological Chemistry of Microbubble Surfactant	33
	83
	83

		5.1.2 Elemental, Infrared, and X-Ray Diffraction Measurements	84
		5.1.3 Pyrolysis Mass Spectrometry	84
		5.1.4 Isolation of Microbubble Surfactant	-84
		5.1.5 Gel-Filtration Column Chromatography, Amino Acid	
		Analysis, and Carbohydrate Determination	85
	5.2	Experimental Results	85
		5.2.1 Abundant Mineral Content and Characteristic IR	
		Absorption Bands	85
		5.2.2 Comparison of Pyrolysis Mass Spectra for Aqueous Soil	00
		Extract, Fulvic Acid, and Water-Soluble Humic Acid	86
		5.2.3 Further Purification of the Microbubble Surfactant Mixture	00
		by Gel-Filtration Column Chromatography	88
		5.2.4 Amino Acid Composition of the Main Glycopeptide	00
		Subfraction from Microbubble Surfactant	89
	5.3	Biochemical/Geochemical Considerations	91
		5.3.1 Interaction of Forest Soil Organic Matter with Abundant	21
		Mineral Content	91
		5.3.2 Dispersal of Microbubble Surfactants in Natural Waters	93
		5.3.3 Bonding Within the Microbubble Surfactant Complex	93
		5.3.4 Probable Biological Source of the Glycopeptide Fraction	23
		of Microbubble Surfactant	94
		of Microbabble Surfactant	94
6	SII	rface Properties of Microbubble-Surfactant	
v.	Me	onolayers	97
	1416	onolayers	9/
	6.1	Modified Langmuir Trough Method	97
		6.1.1 Surface Pressure Measurements with a Cylindrical Rod	97
		6.1.2 Advantages of Method When Testing Complex	
		Biochemical Mixtures	99
		6.1.3 Langmuir Trough Apparatus and Solutions	99
	6.2	Surface Pressure–Area (II–A) Curves	99
		6.2.1 Initial Compression–Expansion Cycle	99
		6.2.2 Effect of Salt Concentration, pH, and Selected	
		Nonelectrolytes	100
		6.2.3 <i>ПА–П</i> Plots	101
	6.3	Selective Desorption from Compressed Monolayers	102
	6.4	Bonding within Compressed Microbubble-Surfactant	
		Monolayers	103
	6.5	Glycopeptide:Acyl Lipid Area Ratio and Association	197
		of Complexes within Monolayers	104
	6.6	Conclusions	105
			† 19g
7.	Str	ucture of Predominant Surfactant Components	
	Sta	bilizing Natural Microbubbles	107
		13.1.7 LUDY for Grand tradition in condition mire blighted to a	
	7.1	¹ H NMR Spectroscopy of Isolated Microbubble Surfactant	107
	1.2	Langmuir-Trough Measurements and Collection of Monolayers	108

	7.3 ¹ H NMR Spectroscopy of Compressed Monolayer Material 7.4 Chemical Similarities Between Microbubble-Surfactant	110
	Monolayers and Lipid Surface Films at the Air/Sea Interface	111
0	Stable Migrobubbles in Physical sign Fluids	
0.	Stable Microbubbles in Physiological Fluids: Competing Hypotheses	113
		-113
	8.1 Comparison of Different Decompression Schedules: Correlation between Bubble Production in Agarose Gels and	
	Incidence of Decompression Sickness	114
	8.1.1 Background Observations	114
	8.1.2 Methods	115
	8.1.3 Experimental Results	115
	8.1.4 Water Depth at First Stop and Total Decompression Time8.2 Comparison of Cavitation Thresholds for Agarose Gels and	117
	Vertebrate Tissues	118
	8.3 Contradictory Findings	118
	8.4 Homogeneous Nucleation Hypothesis	121
	8.5 Clinical Use of Injected Gas Microbubbles: Echocardiography;	121
	Potential for Cancer Detection	121
Pa	t III	
Ph	ysicochemical Properties of Artificial	
	ated Microbubbles and Nanoparticles	125
C	ated wirerobubbles and wanoparticles	125
9.	Concentrated Gas-in-Liquid Emulsions in Artificial Media. I. Demonstration by Laser-Light Scattering	127
	9.1 Physiological Hints for the Production of Artificial Microbubbles	127
	9.2 Laser-Based Flow Cytometry and Forward-Angle	127
	Light Scattering	128
	9.3 Synthetic Microbubble Counts Versus the Control	129
	9.4 Microbubble Flotation with Time	131
	9.5 Microbubble Persistence with Time	132
10.	Concentrated Gas-in-Liquid Emulsions in Artificial Media. II. Characterization by Photon Correlation	
	Spectroscopy	135
	10.1 Brownian Motion and Autocorrelation Analysis of	
		135
	10.2 Parkers of Observations and the H. C. of	100
	10.3 Solubilization of Gases in Micelles	140
	10.4 Size Distribution of Synthetic Microbubbles: Formation,	140
	Coalescence, Fission, and Disappearance	141
	10.4.1 Bimodal Size Distribution of the Microbubble-Surfactan	
	Particle Population	141

	 10.4.2 Combined Evidence that the Larger-Diameter Filmix Particles (i.e., Subpopulation) are Surfactant-Stabilized Gas Microbubbles 10.4.3 Apparent Reversible and/or Cyclical Behavior: Microbubble Formation and Coalescence versus Microbubble Fission and Disappearance 	146
11.	Concentrated Gas-in-Liquid Emulsions in Artificial Media. III. Review of Molecular Mechanisms Involved in Microbubble Stabilization	169
	 11.1 Microbubble Longevity and Interaggregate Interactions 11.2 Molecular Packing within the Microbubble's Surfactant Monolayer 11.3 Repulsive Head-Group Interactions and Monolayer Curvature 11.4 Microbubble Fission, Collapse, and Reemergence 	169 169 170 171
Lip	rt IV pid-Coated Microbubbles and Related Lipid anoparticles in Biomedical Studies on Animals	175
12.	Targeted Imaging of Tumors, and Targeted Cavitation Therapy, with Lipid-Coated Microbubbles (LCM)	177
	 12.1 Description of the LCM Agent (Filmix®) 12.2 Targeted Ultrasonic Imaging of Tumors with LCM as a Contrast Agent 12.3 Tumor Detection Versus Tumor Therapy with LCM 12.4 Use of LCM as a Targeted, Susceptibility-Based, MRI Contrast Agent for Tumors 	177 178 182 184
	12.5 LCM-Facilitated Ultrasonic Therapy of Tumors Therapy beat MD	186
13.	Targeted Drug-Delivery Therapy of Tumors Using LCM	189
	13.1 Internalization of LCM by Tumor Cells <i>In Vivo</i> and <i>In Vitro</i> 13.1.1 LCM Reach Tumors Within Minutes After I.V. Injection: Light- and Fluorescence-Microscopy Data	189
	13.1.2 LCM Preferentially Interact with Tumor Cells In Vivo:	190
	Data from Confocal Laser Microscopy 13.1.3 LCM Are Found Inside Tumor Cells <i>In Vivo</i> : Serial	191
	Optical Sections 13.1.4 LCM Are Endocytosed by Tumor Cells in Culture:	192
	Kinetics of Uptake and Temperature Dependence 13.1.5 LCM Are Found in Acidic Compartments in Tumor Cells in Culture: Confocal Microscopy Using Dual-Channel	193
	Recording 13.1.6 Concluding Remarks	195 195
	If 2 YEAR HOREIDEN CHARGE CHARLES CHARLES AND MICE AND MICE AND COLORS	193

	13.2 Evaluation of LCM as a Delivery Agent of Paclitaxel (Taxol®)	
	for Tumor Therapy	197
	13.2.1 Experimental Methods	197
	13.2.2 Pharmacological Results	198
14.	Proposed Mechanism of Selective LCM Uptake by Tumor Cells: Role of Lipoprotein Receptor-Mediated Endocytic Pathways	207
	Notice of the late	207
	14.1 Low-Density Lipoprotein Receptors, on Tumor Cells, and LCM	207
	14.2 Multiligand Lipoprotein Receptors	207
	14.2.1 LDL Receptor-Related Protein, on Tumor Cells,	11209
	and LCM	209
	14.2.2 Scavenger Receptors on Tumor Cells as well as "Activated" Macrophages: LCM Binding, and Its Relation to Certain Disease Sites	211
	Ad Principles and Valuation Dynamics	
15.	Endocytotic Events Versus Particle Size: Multidisciplinary Analyses Demonstrate LCM Sizes	
	are Mostly Submicron	217
	15.1 Chylomicron Remnant-Like Particle Sizes	217
	15.2 Comparison with LCM Sizes: Proportion of LCM Population between 0.1 and 0.2 μm	218
D	("mind to see A local and to readynastic F.	
	rt V If-Assembling Mixed-Lipid Microbubbles and	
	moparticles for Clinical Applications	221
	Contract Agent (ig Timers) Salahed Agent (ig Timers)	
16.	LCM and Nanoparticle Subpopulations for	
	Drug Delivery	223
	16.1 Stable Nanoemulsions	223
	16.2 Mixed-Lipid "Microbubble Versus Nanoparticle"	
	Interrelationships in Filmix®	224
	16.2.1 Nanoparticles Based on Solid Lipids: Background	
	Literature	225
	16.2.2 "Dispersed LMN" (or Colloidal Liquid Crystals) and	
	Targeted Drug Delivery	225
	16.2.3 Self-Assembly and Interplay of LCM, Dispersed LMN, and Mixed Micelles: Correlations with Bile Colloids	226
17	Comparition of ICM Comparing Internal Management	
17.	Composition of LCM Governing Interplay with Nanoparticle Subpopulation	229
	Comment of the contract of the	
	17.1 Patented LCM Components	229
	17.2 LCM Structural Characteristics Affecting Molecular Interchange with Dispersed LMN and Mixed Micelles	230

15	17.3 Film-Shedding Transitions and/or Collapse in Lipid Monolayers Coating Microbubbles	230
	17.3.1 Dissolution of Microbubbles in Degassed Media 17.3.2 Dissolution of Microbubbles via Ultrasound 17.4 Concluding Remarks	230 231 233
18.	Targeted Nanoparticle Subpopulation: Comparison with Self-Nanoemulsifying Drug-Delivery Systems in Pharmaceutical Research	235
	 18.1 Small Energy Input for Production: Self-Nanoemulsification 18.2 Medium-Chain, and Long-Chain, Glycerides 18.3 Nonpolar Core of Nanoemulsion Particles 	235 236 237
19.	Clinical Development of an "LCM/Nanoparticle-Derived Formulation: A Nanoemulsion Based Upon "Dispersed LMN"	239
	19.1 Details of Chemical Composition of the Clinical-Grade "Non-Gas Containing, Lipid Nanoparticles" (or Clinical-Grade	233
	"Dispersed LMN") 19.2 Particle Size Distribution of Clinical-Grade Dispersed LMN Prepared with, or without, Incorporated Drug and/or Additives	240241
	19.3 Targeting Properties of Clinical-Grade Dispersed LMN: Drug Delivery to Tumor Cells19.4 Targeted Drug-Delivery by Clinical-Grade Dispersed LMN:	243
	Effects of Different Additives	246
20.	Selected Parenteral Lipid Nanoemulsions Under Clinical Study: Comparison Concerning Passive Accumulation in Tumors, Active Targeting of Tumors, and Validation Status	by'
	20.1 Tocol Nanoemulsions for Solubilizing, and Drug Delivery, of	247
	Paclitaxel: Passive Accumulation in Tumors 20.2 Cholesterol-Rich/Phospholipid Nanoemulsions Containing Derivatized Paclitaxel: Active Uptake into Tumors via	248
	20.3 Stable (Nonphospholipid, Nonprotein) Lipid Nanoemulsions for Targeting Tumors: Active Uptake of (Unmodified)	249
11		251
41.	Supplementary Operational Benefits Concerning "LCM/Nanoparticle-Derived" Formulations:	ji3a
		255
	21.2 Filmix® Chemical Composition Supports Long-Term Stability	255 257

21.3 04.5 04.5	Rapid (Nondestructive) Determination of Particle Size Distribution, of the Parenteral Nanoemulsion, by Standard Light-Scattering Methods	260
Biolog Recep	/I /Nanoparticle-Derived" Nanoemulsions: gical Lipid Polymorphism, and otor-Mediated Endocytosis, used for cal Study	265
22. Bio "D	ological Lipid Polymorphs: Preferred Cubic Phase of ispersed LMN"	267
	1 Biological Importance of Lipid Polymorphism: Focus on Cubic Phases 2 Inverse Micellar Cubic Phase: Special Relevance to	267
	"Dispersed LMN" 3 Physicochemical Tendency of the Dispersed LMN to Adopt "Nonlamellar" Mesostructural Topology: Roles of	270
	Head-Group Hydration, Acyl Chain Length, and Cholesterol Content	272
	onlamellar Phase(s) Facilitating Membrane Fusion: docytosis of Dispersed LMN	277
	1 Inverse Cubic Phase(s) Induce or Facilitate Membrane Fusion: The Stalk Mechanism 2 Inverse Bicontinuous Cubic Phases in Phosphoglyceride—	277
23.	Cholesterol Mixtures: Cholesterol as an Inducer of Biomembrane Fusion and Endocytosis Endocytosis of Dispersed LMN: Competing	279
	ceptor-Mediated Endocytosis of (Mixed-Lipid)	281
	spersed LMN 1 Characteristics of Scavenger Receptors (Versus LDL	287
	Receptor and LRP) 2 Biophysical Properties of a "Class B" Scavenger	287
24.	Receptor: SR-BI	292
	24.2.1 SR-BI Versus CD36	292
	24.2.2 SR-BI: Endocytosis and "Selective" Uptake?	294
24.	24.2.3 SR-BI, Membrane Domains, and Cholesterol 3 Endocytosis Mediated by SR-BI: Comparison of Human	297
	Tumor Cell Lines	299