THE
ORGANIZATION
OF CELLS
AND OTHER
ORGANISMS
L. E. R. PICKEN

E702

PI

8590291

外文书库

THE

ORGANIZATION OF CELLS

AND OTHER ORGANISMS

BY
LAURENCE PICKEN

原中山医 院生物系主任 黄绮 教授惠赠

OXFORD
AT THE CLARENDON PRESS

© Oxford University Press 1960

PRINTED IN GREAT BRITAIN

SPOMINU FRANCA FORTUNATA ČLOVEKA IN PRIJATELJA

PREFACE

It is addressed to postgraduate students and research workers. Though its scope is by no means limited to a consideration of recent developments, its writing has largely been made possible by the modern practice of publishing specialized reviews of recent work in the separate fields here brought into relation. In some circles, this type of relatively ephemeral review is criticized for its lack of integration. It performs none the less a valuable service, and will continue to do so until the practical need for a means of access to past work—a means more direct than the voluminous printed word—forces the sciences to abandon current conceptions, tacit and overt, of the aims and purposes of scientific publication. A scientific paper is many things other than a mere instrument serving to communicate to others the nature and results of a limited inquiry. The time may come, however, when the bulk of scientific literature is seen to act as a hindrance to accurate knowledge of the past, and with its coming habits may change.

In the course of this book, I have sought to make plain my indebtedness to reviewers in particular, by drawing attention in a footnote at the beginning of each chapter to the principal books and articles by which I obtained access to the literature of a particular field. Every effort has been made to limit the bibliography rather than to extend it, and the number of entries has been reduced whenever possible during revision. The aim has been to leave a sufficiency of references for credit to be given where it is due, and for the reader to be able to find his way into the relevant literature;

the bibliography has no wider aim of completeness.

A major part of my intention has been to illustrate the change and development of concepts, since it is still not widely appreciated that the working biologist is at least as much concerned with concepts as he is with things. In some instances the narrative has dipped briefly into the earlier literature in the hope, perhaps vain, of mitigating belief in the present as an epoch of superior intelligences by demonstrating the considerable age of many current conceptions. But again, I have not sought to be exhaustive in historical bibliography, or even consistent in the use of the historical approach.

I am aware that my coverage of recent publications in English is more complete than that of publications in other European languages, and that my treatment of Russian, Chinese, and Japanese publications, for example, is casual rather than systematic. Although this implies that the bibliography has a strongly Anglo-American bias, I do not think the book is a different book from what it would have been if all languages were equally covered. Indeed, I believe it might have been written in the language of any

one of the major nineteenth-century powers, and based on an almost exclusively national bibliography, without loss in range or restriction in outlook.

Many authors will find that I have but slightly paraphrased their words, or have used their own words in tacit quotation, in writing of their work or reporting their views. In covering a field so large that most topics lie outside the range of first-hand experience, it appeared essential to remain as close as possible to the author's own formulation, rather than to accept the risk of a deforming periphrasis. If I have paraphrased without acknowledging indebtedness, I here and now express that indebtedness, and also a more general indebtedness to all authors who have led me to new aspects of the literature, or who have provided references that might otherwise have been missed.

It is to be expected that such a compilation will contain errors of emphasis, interpretation, or fact. I have tried to reduce their number by seeking criticism from specialists in the various fields of the chapters. From their distinguished names the reader will gain assurance that the number of errors is less than it would have been but for their help. I trust that those readers who find their work misrepresented will kindly inform me of my inadequacies of presentation.

LAURENCE PICKEN

Jesus College **Cambridge**

ACKNOWLEDGEMENTS

My very grateful thanks are due to the following friends and colleagues who read and commented on single chapters or portions of chapters and in some instances on several chapters: Professor M. Abercrombie, F.R.S., Professor H. G. Callan, Professor K. W. Cleland, Dr. Ilham Dilman, Dr. Jean Hanson, Professor J. E. Harris, F.R.S., Dr. M. L. Johnson, Dr. P. Mitchell, Dr. J. M. Mitchison, Professor R. D. Preston, F.R.S., Professor J. T. Randall, F.R.S., Dr. K. M. Rudall, Dr. B. M. Shaffer, Dr. A. P. Sims, Dr. J. D. Smith, Dr. K. M. Smith, F.R.S., Professor M. M. Swann, Mr. Bernard Towers.

To Mr. R. G. Pottage, Dr. A. V. Grimstone, Mr. and Mrs. S. A. Wainwright, and Mr. and Mrs. M. J. Wells I owe a particular debt for the careful comparison of successive typescripts with each other and with the first proofs. My debt to Mrs. I. Marcks-Franke, who typed a fair copy of the bibliography from my cardindex, and prepared the subject-index, will only be fully appreciated by those who have themselves experienced these less-agreeable aspects of the author's

task.

Except where described as 'original', the text-figures have been re-drawn by Mr. M. J. Wells, who spared no pains in reproducing original shading and texture,

and in translating from lithograph or half-tone to line.

My thanks are also due to all those who so generously provided the photographs from which the plates were compiled: Dr. T. F. Anderson (Plate 2, Fig. 4); Professor W. T. Astbury, F.R.S. (Plate 8, Figs. 1a, b, 2); Dr. Elizabeth J. Batham (Plate 29, Fig. 5); Dr. W. Beermann (Plate 26, Fig. 3); Professor J. T. Bonner (Plate 8, Fig. 4; Plate 34); Dr. J. R. G. Bradfield (Plate 9, Figs. 2a-c); Professor H. G. Callan (Text-figs. 20, 21; Plate 10, Figs. 1a, b; Plate 12, Figs. 1, 3, 4; Plate 13, Figs. 1a, 2); Dr. G. B. Chapman (Plate 9, Fig. 1); Professor M. Chevremont (Plate 16, Figs. 1a-c); Professor L. R. Cleveland (Plate 19, Figs. 1a, b); Professor E. Fauré-Fremiet (Plate 20, Fig. 3); Dr. R. J. Goldacre (Plate 10, Figs. 2a, b; Plate 24, Figs. 1a, b); Dr. C. B. Goodhart (Plate 15, Fig. 3); Professor K. G. Grell (Plate 10, Fig. 4); Dr. A. V. Grimstone (Plate 13, Fig. 16; Plate 16, Figs. 3a-e; Plate 17, Fig. 2; Plate 20, Fig. 1); Dr. T. Gustafson (Plate 33, Fig. 3); Dr. Jean Hanson (Plate 22, Fig. 4); Professor J. E. Harris (Plate 11, Figs. 1a-c); Dr. A. J. Hodge (Plate 18, Fig. 2); Dr. A. L. Houwink (Plate 26, Figs. 2a, b); Dr. H. Hurst (Plate 5, Figs. 4a, b); Dr. H. E. Huxley (Plate 23, Figs. 2a, b); Professor N. Kamiya (Plate 14, Figs. 1a-c, Figs. 2a-c); Dr. E. Kellenberger (Plate 6, Fig. 3); Dr. J. A. Kitching (Plate 15, Figs. 1a, b); Dr. A. Klug (Plate 3, Fig. 3); Dr. L. W. Labaw (Plate 7, Fig. 1; Plate 8, Fig. 3); Professor F. E. Lehmann (Plate 33, Fig. 1); Dr. H. Leyon (Plate 18, Figs. 1, 3); Dr. E. H. Mercer (Plate 19, Fig. 3; Plate 21, Figs. 2-4; Plate 31, Fig. 4; Plate 32, Figs. 1, 2); Dr. J. M. Mitchison (Plate 14, Fig. 3; Plate 24, Fig. 2); Dr. C. Morgan (Plate 2, Fig. 1); Dr. A. Pijper (Plate 6, Figs. 1a-d); Professor R. D. Preston, F.R.S. (Plate 26, Fig. 1); Professor J. T. Randall, F.R.S. (Plate 16, Figs. 2a, b; Plate 20, Fig. 2; Plate 24, Fig. 3; Plate 27, Figs. 1-4; Plate 28, Figs. 1-6; Plate 30, Fig. 2); Dr. J. D. Robertson (Plate 25, Fig. 4); Dr. C. Robinow (Plate 5, Figs. 1-3); Lord Rothschild, F.R.S. (Plate 25, Fig. 1); Dr. K. M. Rudall

(Plate 29, Figs. 1-4; Plate 31, Figs. 2, 3); Dr. G. G. Selman (Plate 33, Fig. 2); Dr. M. J. R. Salton (Plate 7, Figs. 2, 3); Professor W. J. Schmidt (Plate 15, Figs. 2a, b); Dr. B. M. Shaffer (Plate 33, Figs. 4a-d); Professor F. S. Sjöstrand (Plate 17, Fig. 1; Plate 25, Fig. 2); Dr. D. S. Smith (Plate 23, Figs. 1a-c); Dr. K. M. Smith, F.R.S. (Plate 2, Figs. 2a, b, Fig. 3; Plate 4, Figs. 1, 2, 6); Dr. W. Straus (Plate 18, Fig. 4); Professor M. M. Swann (Plate 19, Figs. 2a-h); The Textile Physics Laboratory of the University of Leeds (Plate 21, Fig. 1; Plate 22, Figs. 1-3); Dr. W. Thornburg (Plate 22, Fig. 5); Professor J. Tomcsik (Plate 6, Figs. 2a, b); Professor A. Tyler (Plate 25, Fig. 3); Dr. C. Weibull (Plate 5, Fig. 5); Dr. P. Weiss (Plate 30, Figs. 1a, b); Dr. M. H. F. Wilkins (Plate 1; Plate 3, Fig. 1; Plate 10, Figs. 3, 5; Plate 11, Figs 2a-d; Plate 12, Fig. 2);

Professor R. C. Williams (Plate 3, Figs. 2a-c; Plate 4, Figs. 3-5, 7).

The following journals, publishers, and societies have kindly granted permission for the reproduction of illustrations: Academic Press Inc. (Text-figs. 28, 34, 38, 39, 56, 57; Plate 2, Fig. 1; Plate 3, Fig. 2; Plate 4, Figs. 3-5, 7; Plate 14, Fig. 2; Plate 17, Fig. 1; Plate 18, Fig. 3; Plate 20, Fig. 3; Plate 26, Fig. 3; Plate 33, Fig. 3); Acta Botanica Neerlandica (Plate 26, Fig. 2); Akademische Verlagsgesellschaft (Text-figs. 53, 54, 93); George Allen & Unwin Ltd. and Macmillan Co. (New York) (Text-fig. 110 from Darlington and Mather, The Elements of Genetics); American Journal of Botany (Plate 34); The American Naturalist (Text-figs. 107, 108; Plate 2, Fig. 4); American Philosophical Society (Text-fig. 22 from Trans. Amer. Phil. Soc.; Text-fig. 81 from Proc. Amer. Phil. Soc.); Biochemical Journal (Plate 28, Fig. 6); Biological Bulletin (Text-fig. 49; Plate 25, Fig. 3); Blackwell Scientific Publications Ltd. (Text-fig. 14); Gebrüder Borntraeger (Text-figs. 69, 71); Butterworths (Text-fig. 13; Plate 27, Figs. 1, 4; Plate 28, Figs. 1-3); Cambridge University Press and the Council of the Marine Biological Association (Text-fig. 31); Cambridge University Press (Text-fig. 67 from Beale, The Genetics of Paramecium aurelia; Text-fig. 87 from D'Arcy Thompson, Growth and Form; Text-fig. 94 from Gray, Proc. Camb. phil. Soc. biol. Sci. and A Textbook of Experimental Cytology; Plate 2, Figs. 2, 3 from Parasitology); Chapman & Hall Ltd. (Text-figs. 72, 74; Plate 26, Fig. 1); J. & A. Churchill Ltd. (Text-fig. 5; Plate 3, Fig. 3); Clarendon Press (Text-fig. 61); Colston Research Society and Butterworths (Text-fig. 64); the Company of Biologists Ltd. (Text-figs. 15, 57, 42, 43, 55, 63, 65, 66, 68, 73, 86, 94, 113; Plate 5, Fig. 4; Plate 8, Figs. 1, 2; Plate 10, Fig. 1; Plate 15, Fig. 1; Plate 19, Fig. 2; Plate 24, Figs. 1, 2; Plate 25, Fig. 1; Plate 33, Figs. 2, 4); Elsevier Publishing Co., Amsterdam (Text-fig. 36 from Kruyt, Colloid Science, 2, xi, 439 (1949), Fig. 6; Plate 6, Fig. 3 from Biochim. biophys. Acta, 17, 1 (1955), fig. 4; Plate 7, Fig. 1 and Plate 8, Fig. 3 from Biochim. biophys. Acta, 15, 325 (1954), figs. 2 and 6; Plate 19, Fig. 3 from Biochim. biophys. Acta, 3, 161 (1949); Plate 22, Fig. 4 from Biochim. biophys. Acta, 23, 229 (1957), fig. 8); the Faraday Society (Text-fig. 48); Genetics (Text-figs. 6, 7, 11); Gustav Fischer Verlag, Stuttgart (Text-figs. 51, 62); Long Island Biological Association (Text-fig. 12); Macmillan & Co. Ltd. (Text-figs. 8, 17, 27, 32, 44; Plate 4, Fig. 6; Plate 10, Fig. 2; Plate 21, Fig. 4 from Nature); Masson & Cie Editeurs (Text-figs. 35, 47, 52, 112; Plate 16, Fig. 1); New York Academy of Sciences (Plate 11, Figs. 2a-c); Erven P. Noordhoff Ltd. (Text-figs. 21, 70); Oliver & Boyd Ltd. (Text-fig. 19); Faculty of Sciences, Osaka University (Text-fig. 33); Pergamon Press

Ltd. (Text-figs. 2, 3, 4 from Markham, Progress in Biophysics (1953)); Presses Universitaires de France (Text-fig. 111); Princeton University Press (Plate 8 Fig. 4); Revue Suisse de Zoologie (Text-figs. 88-90); Rockefeller Institute Press (Text-fig. 29 from Journal of General Physiology, Text-figs. 41, 46; Plate 16, Fig. 2; Plate 18, Fig. 2; Plate 20, Fig. 2; Plate 21, Fig. 2; Plate 22, Fig. 5; Plate 23, Fig. 2a; Plate 25, Fig. 4; Plate 30, Fig. 1 from Journal of Biophysical and Biochemical Cytology); the Royal Microscopical Society (Plate 12, Fig. 2); the Royal Society (Text-figs. 18, 23, 79; Plate 9, Fig. 3; Plate 13, Fig. 1a; Plate 24, Figs. 4, 5); the Royal Society of Queensland (Text-figs. 82, 83); E. Schweizerbart'sche Verlagsbuchhandlung (Text-figs. 91, 92); Science (Plate 18, Fig. 4); Society for Applied Bacteriology and the Editor of Journal of Applied Bacteriology (Plate 5, Fig. 1); Society for General Microbiology (Text-fig. 16); Society of Leather Trades Chemists (Text-fig. 58); Springer-Verlag, Heidelberg (Textfigs. 75-78; Plate 15, Fig. 2; Plate 33, Fig. 1); Springer-Verlag, Vienna (Textfig. 40); Swedish Academy of Sciences (Plate 5, Fig. 5); University of California Press (Text-fig. 50 from University of California Publications, Zoology); University of Chicago Press (Text-figs. 9, 10, 24, 59, 60, 80 from Proceedings of the National Academy of Sciences); Urban & Schwarzenberg (Text-fig. 85); VEB Gustav Fischer Verlag (Text-fig. 25); The Williams & Wilkins Co. (Text-fig. 1: Plate 9, Fig. 1); Wistar Institute of Anatomy and Biology (Text-fig. 45 from American Journal of Anatomy; Text-figs. 97-106 from Journal of Experimental Zoology; Text-fig. 100 from Journal of Morphology; Plate 25, Fig. 2 from Journal of Cellular and Comparative Physiology); Zeitschrift für Naturforschung (Plate 18. Fig. 1).

It is a pleasure to recall the facilities afforded me in the University of Washington, Seattle, during my tenure of a Walker-Ames Visiting Professorship in Zoology in the Spring Quarter of 1959, whereby the burden of attending single-

handed to matters of copyright was greatly lightened.

I wish finally to record that this book was completed and the bibliography checked in the incomparable library of the Stazione Zoologica di Napoli.

L.E.R.P.

LIST OF PLATES

- Model of desoxyribonucleic acid B, based on data published by Langridge et al. (1957). Unpublished photograph provided by Dr. M. H. F. Wilkins. See pp. 32, 108.
- I. Section through the cytoplasm of a cell from the chorio-allantoic membrane of a chick infected with herpes simplex virus. The sectioned virus bodies reveal a hollow or solid core surrounded by a single or double membrane. Photograph by C. Morgan, S. A. Ellison, H. M. Rose, and D. H. Moore from Williams (1954). Photograph provided by Dr. C. Morgan. See p. 25.
 - 2a. Shadowed granules from nuclei of hypodermis or fat-body cells of granulosis-infected *Natada nararia*, the nettle grub (Smith and Xeros. 1954). See p. 25.
 - 2b. Granules shadowed after treatment with a weak aqueous solution of sodium carbonate. The outer capsule is now represented by an expanded residue covering the inner capsule. Dissolution of the latter reveals a thin virus rod, itself enclosed in a membrane (Smith and Xeros, 1954). Photographs provided by Dr. K. M. Smith. See p. 25.
 - 3. Section through a mass of vesicles of polyhedral virus in the nucleus of a blood cell of *Tipula paludosa*. Each vesicle contains a single virus rod. Near the nuclear membrane, the vesicles appear to be compressed, as crystallization and the formation of polyhedra progresses. (Compare with the cytoplasmic virus shown in Plate 4, Fig. 6.) Photograph by S. Vernon-Smith from Smith (1955b); provided by Dr. K. M. Smith. See p. 25.
 - 4. Stereo electron micrographs of T2 'phage adsorbed on 'ghosts' of Escherichia coli B, dried from the frozen state; unshadowed (Anderson, 1951, 1952). Photograph provided by Dr. T. F. Anderson. See p. 25.
- 3. I. A crystal of tobacco mosaic virus in a cell of a hair from an infected plant, photographed between crossed Nicols with compensation. When the cell is injured such crystals break down into paracrystalline needles. Unlike the intracellular crystals, all precipitates of the virus formed in vitro are paracrystals (Wilkins et al., 1950). Unpublished photograph provided by Dr. M. H. F. Wilkins. See p. 30.
 - 2a. Unshadowed electron micrograph of tobacco mosaic virus particles.
 - 2b. Electron micrograph of shadowed TMV particles.
 - 2c. Electron micrograph of shadowed X-protein. This is an abnormal protein from infected plants which aggregates as rods, morphologically indistinguishable from native TMV, but not infective and of variable length (Williams, 1954). Photographs provided by Dr. R. C. Williams. See p. 29.
 - 3. X-ray diffraction diagram obtained from an oriented gel of a normal strain of tobacco mosaic virus. The virus particles are oriented with their long axes parallel to the white streak. The equatorial reflections out to 5 A indicate hexagonal packing of the particles. Unlike the equatorial reflections, the non-equatorial reflections are not affected by the water content and yield information about the internal structure of the particles. A pronounced helical feature of the diagram is the tendency of the meridional reflections to split. The horizontal stratification corresponds to the axial period of 69 A (Franklin et al., 1957). Photograph provided by Dr. A. Klug. See p. 32.
- 4. I. Rhombic dodecahedral crystals of tomato bushy stunt virus. These crystals belong to the cubic system and are isotropic. Photograph provided by Dr. K. M. Smith. See p. 30.
 - 2. Electron micrograph of a replica of a tobacco necrosis virus crystal. The virus particles are packed in approximately cubic packing, but such crystals are birefringent (Markham et al., 1948). Photograph provided by Dr. K. M. Smith. See p. 30.

- 3. Close-packed array of non-spherical, polyhedral particles of tomato bushy stunt virus. The top row of particles is slightly distorted by shadowing (Williams, 1954). Photograph provided by Dr. R. C. Williams. See p. 30.
 - 4. Ordered arrays of particles of purified poliomyelitis virus (MEF-1, Type II) grown in monkey kidney tissue. Single particles appear larger because of flattening on drying, and deposition of the shadowing film (Williams, 1954). Photograph provided by Dr. R. C. Williams. See p. 26.
 - 5. Frozen-dried and double-shadowed spherical particles of influenza virus (P.R.S.). Double-shadowing defines the complete contour. Chick-embryo infections also contain filamentous forms of the virus (Williams, 1954). Photograph provided by Dr. R. C. Williams. See p. 25.
 - 6. Low-power electron micrograph of part of an ultra-thin section through a methacrylate-embedded pellet of purified *Tipula paludosa* virus. The centre-to-centre distance between the polyhedral virus particles in the closest packed arrays is 1,300 A. In contrast to the polyhedrosis of *T. paludosa* (Plate 1, Fig. 3) this is a cytoplasmic virus. It is also the largest virus of which the particles are completely uniform in size and shape (Williams and Smith, 1957). Photograph provided by Dr. K. M. Smith.
 - 7. T4 'phage shocked with ammonium acetate, showing tail fibres, tail, and faceted heads (Williams and Fraser, 1956). Photograph provided by Dr. R. C. Williams. See p. 25.
- 5. Ia. Section of a resting spore of B. megaterium fixed with osmium tetroxide. The cortex (electron-transparent) surrounds the dormant bacillus (the core). Within the latter, electron-transparent regions may be the chromatinic elements. See p. 58.
 - Ib. Section of an osmium-fixed, acid-treated, resting spore of B. megaterium, stained with 1 per cent. lanthanum nitrate, and showing the fine structure of the cortex. The surface of the core is dense and thrown into folds, probably as a result of shrinkage. Photographs by Mayall and Robinow (1957), provided by Dr. C. F. Robinow. See p. 58.
 - 2a. Chromatinic bodies in cells from 1-1½-hr.-old cultures of B. megaterium at 37° C, fixed with osmium tetroxide vapour, hydrolysed with HCl and stained with Azure-A. First- and second-division stages are shown. There is an empty spore coat in the top left-hand corner. See p. 58.
 - 2b. Irregular chromatinic patterns in cells from a somewhat older culture of B. megaterium in which lipid droplets are abundant. See p. 59.
 - 2c. Accessory granules in B. megaterium. These are shown in germinating spores developing into the first generation of vegetative bacilli. Unpublished photographs provided by Dr. C. F. Robinow. See p. 60.
 - 3a. Chromatinic bodies in *Tetracoccus* showing characteristic configurations. This again is an Azure-A preparation.
 - 3b. A preparation of *Tetracoccus* showing the arrangement of cell walls in the groups of cocci. This may be compared with the distribution of chromatinic bodies shown in 3a. The cells are stained by Hale's method. Unpublished photographs provided by Dr. C. F. Robinow. See p. 58.
 - 4a. Electron diffraction pattern obtained from Saccharomyces cerevisiae cells after contact with chloroform. The 4·12 A ring is intensified and shows two rectangular grids of spots. On the fainter, outer, 3·72 A ring, a pair of spots is associated with a pair on the inner ring, as indicated by the dotted lines. The electron beam presumably covered two cells only, and the a- or b-axes of the oriented, bound lipid are distributed on each cell as in a single mosaic crystal.
 - 4b. Electron diffraction pattern obtained from Escherichia coli cells after contact with chloroform. A continuous ring pattern such as this would be produced by the many bacteria in the field in random orientation, even if the crystals on the individual bacteria were aligned as in the yeast cell.

- 5. The presence of the 4·12 and 3·72 rings in these photographs implies that the hydrocarbon chains of the lipids are normal to the surface of the cell wall (Hurst, 1952). Photographs provided by Dr. H. Hurst. See p. 64.
 - 5. Spiral aggregate of flagella obtained from *Proteus vulgaris* by mechanical separation (Weibull, 1950). The photograph was taken at high focus and with phase contrast. Photograph provided by Dr. C. Weibull. See p. 79.
- 6. Ia. Salmonella typhi (dark ground), showing the type of random contact observed between individuals at rest.
 - 1b. Salmonella typhi (dark ground), showing polar contacts established in the presence of O-antibody. See p. 69.
 - Ic. Salmonella typhi (dark ground). The agglutinated individuals are linked through their flagella. Both flagella and body are enlarged by adsorption of H-antibody. See p. 69.
 - Id. Salmonella typhi (dark ground). In the presence of Vi-antibody, aggregates are built up in which contact between individuals is mainly lateral. Unpublished photographs provided by Dr. A. Pijper. See p. 71.
 - 2a. Structure of the capsule in Bacillus M (phase contrast). After inoculation from an agar culture into Sauton's medium, with aeration for 3 hours at 32° C, the centrifuged sediment of young bacilli was maintained at 20–28° C for 3 days. A suspension of the sediment was then treated with polypeptide antibody (Tomcsik and Guex-Holzer, 1954).
 - 2b. A chain of bacilli from the same suspension after treatment with polysaccharide antibody, showing unusually well-developed transverse septa (Tomcsik and Guex-Holzer, 1954). Photographs provided by Professor J. Tomcsik. See pp. 83, 84.
 - 3. The receptor for 'phage-T5, isolated from *E. coli* B, has combined with 'phage and appears in the photograph as a small, spherical particle (Weidel and Kellenberger, 1955). Electron micrograph supplied by Dr. E. Kellenberger. See p. 76.
 - I. Electron micrograph of a portion of the wall of an unidentified, Gram-positive rod, showing rectangular periodic structure on the inside of the wall (Labaw and Mosley, 1954). The scale is 0·5 μ. Photograph provided by Dr. L. W. Labaw. See pp. 75, 77.
 - 2. Cell wall of *Rhodospirillum rubrum* prepared for electron microscopy by freezedrying. The spherical macromolecules are about 100 A in diameter (Salton and Williams, 1954). Photograph provided by Dr. M. R. J. Salton. See pp. 76, 77.
 - 3. Cell wall of *B. megaterium*. At this resolution the structure appears homogeneous (Salton and Williams, 1954). Photograph provided by Dr. M. R. J. S. Iton. See p. 76.
 - Ia. X-ray diffraction pattern obtained from a film of Proteus flagella, taken at room humidity and with the beam parallel to the plane of the film (Astbury et al., 1955).
 - 1b. X-ray β -fibre diagram obtained after stretching a film of *Proteus* flagella by c. 200 per cent. in a saturated aqueous solution of ammonium sulphate at 90–95° C (Astbury et al., 1955). Photographs provided by Professor W. T. Astbury. See p. 79.
 - 2. Electron micrograph of a 'lock' of *Proteus* flagella (Astbury et al., 1955). Photograph provided by Professor W. T. Astbury. See pp. 79, 81.
 - 3. Electron micrographs of flagella from an unidentified organism, showing a counterclockwise double helix (Labaw and Mosley, 1954). The scale is 0.5 μ . Photographs provided by Dr. L. W. Labaw. See p. 80.
 - 4. Development of the fruiting body of the myxobacterium, *Chondromyces crocatus*, photographed at hourly intervals by Dr. E. A. Wheaton (Bonner, 1952a). Photographs provided by Dr. J. T. Bonner. See p. 85.
 - 9. I. Ultra-thin sections of cells of a Rough strain of Bacillus cereus. The individual cut in longitudinal section shows the developing new wall and the electron-transparent 'peripheral bodies' embracing the edge of the annular disk. To right and left of the

- developing septum are electron-transparent regions marking the site of the chromatinic bodies (Chapman and Hillier, 1953). Photograph provided by Dr. G. B. Chapman. See p. 82.
- 9. 2. Photomicrographs of Cristispira balbiani from the crystalline style of the oyster photographed in ultraviolet light (wavelength 2,570 A) at three different levels of focus. Parts of the undulating membrane can be seen in focus in (a), (b), and (c). Blocks of ultraviolet-absorbing material in linear sequence are visible in all three photographs but are shown clearly in (b). Unpublished photographs provided by Dr. J. R. G. Bradfield. See p. 92.
 - 3. The movement of a green helical organism. Shots from a ciné film taken at the rate of eight pictures a second. From left to right and from above down their serial numbers are: 2, 5, 9, 16, 20, 22, 24, 26. The set of six photographs covers a time interval of about 3 seconds. The helix appears to be left-handed because the lower parts of the coils are in focus. The granule marked with an arrow moves from the posterior end forwards until it is about a third of the way along the body. In Figs. (e) to (h) a stationary granule, X, is visible on the posterior edge of the helix (Picken, 1940a). See p. 94.
- ia. Optical section of an oocyte nucleus of Xenopus laevis isolated in potassium phosphate solution at pH 6.4. The structural colloid is unswollen and of normal translucency.
 - 1b. Optical section of an oocyte nucleus of *Triturus cristatus carnifex* isolated in potassium phosphate solution at pH 7. The structural colloid has dispersed, and the nuclei and chromosomes lie on the floor of the nucleus. Photographs from Callan (1952) provided by Professor H. G. Callan. See p. 105.
 - 2a. Three-dimensional bacterial array of an unidentified organism in a very dilute balanced salt solution (Chalkley's medium) containing 1.5-3 per cent. of glycerol. End-on view.
 - 2b. Side-view of another aggregate. Photographs from Goldacre (1954) provided by Dr. R. J. Goldacre. See p. 136.
 - 3. Sperm heads of Sepia oriented by surface forces at the edge of a drop, photographed in polarized light. Unpublished photograph by Wilkins and Battaglia provided by Dr. M. H. F. Wilkins. See p. 109.
 - 4. Tokophrya sp. (Suctoria) photographed in life to show the appearance of strings of beads in the macronucleus, due to optical section of successive turns of spiralized chromosomes. Unpublished photograph provided by Professor K. G. Grell. See p. 102.
 - 5. X-ray diffraction pattern of intact T2 'phage at 75 per cent. relative humidity. The pattern is largely due to the DNA component. Unpublished photograph provided by Dr. M. H. F. Wilkins, See p. 108,
- 11. I a, b, c. Shots from a ciné film of the fall of the nucleolus under gravity through the nucleus of an oocyte of Echinus esculentus. The time interval between (a) and (c) is about 3 minutes. The diameter of the nucleus is 70 \(\mu\). Unpublished photographs from the negative provided by Professor J. E. Harris. See p. 104.
 - 2a. X-ray diffraction photograph of calf thymus DNA (sodium salt) in the A configuration, at 75 per cent. relative humidity.
 - 2b. X-ray diffraction photograph of DNA from avian tubercle bacilli, at 75 per cent. relative humidity.
 - 2c. Comparison of the X-ray diffraction photographs of (below) unfixed Loligo sperm in the intact spermatophore; (above) fibres (prepared from sperm removed from the spermatophore) washed in water, dried and rehumidified at 98 per cent. relative humidity.
 - 2d. X-ray diffraction photograph of wet trout sperm heads. The first intense ring out from the central blackened area corresponds to the first equatorial reflection in (a), (b), and (c). Unpublished photograph by M. H. F. Wilkins and H. R. Wilson (1955).

- No.
 - Photographs (a), (b), and (c) from Wilkins (1957) where a list of collaborators concerned in the preparation of the various specimens will be found (op. cit., p. 184). Photographs provided by Dr. M. H. F. Wilkins, See pp. 108, 109.
- 12. I. Chromosome XII entire from *Triturus cristatus carnifex* -/+ heterozygote at the giant loops locus (marked by an arrow.) The opposed arrows mark the centromeres. Unpublished photograph provided by Professor H. G. Callan. See p. 114.
 - 2. A bundle of air-dried locust sperm in glycerol, photographed in polarized ultraviolet light (2,652 A) with a reflecting objective. The width of a single sperm head is 0.7 μ . The electric vector is horizontal, and in two quadrants absorption (due to oriented DNA) is strong. Photograph by Wilkins (1953) provided by Dr. M. H. F. Wilkins. See p. 109.
 - 3. Late zygotene or pachytene in *Chortippus parallelus*—an aceto-orcein testis squash. The lack of definition is characteristic of these chromosomes in zygotene-pachytene-diplotene. Unpublished photograph provided by Professor H. G. Callan. See p. 136.
 - 4. Birefringence of the nuclear membrane in an isolated nucleus of *Triturus cristatus carnifex*. Unpublished photograph provided by Professor H. G. Callan. See p. 111.
- 13. Ia. Transmission electron micrograph of the nuclear membrane of *Xenopus laevis* isolated in distilled water, fixed for a minutes in our per cent. phosphotungstic acid. Photograph from Callan and Tomlin (1950) provided by Professor H. G. Callan. See Towlis.
 - Ib. Ultra-thin section of the nuclear membrane of *Trichonympha* (Zoomastigina) showing the appearance of a vesicular, perinuclear cisterna. In some instances the 'pores' seem to be patent; in others closed by a thin membrane. The nuclear contents appear to be finely fibrillar. Unpublished photograph by Dr. A. V. Grimstone. See p. 112.
 - 2. Prometaphase of mitosis in *Triturus cristatus carnifex* (aceto-orcein tail-tip squash) showing centromeres. Unpublished photograph provided by Professor H. G. Callan. See p. 133.
- 14. I a, b, c. Three stills from a ciné film taken by the late Professor W. Seifriz and projected during a lecture in the Stazione Zoologica at Naples in 1952, showing streaming in the mycetozoan, Reticulomyxa filosa. The stills are at intervals of approximately 3 sec. Arrows mark vacuoles moving in adjacent cytoplasmic streams of opposite direction. Unpublished photograph provided by Dr. N. Kamiya from his copy of Professor Seifriz's film. See p. 172.
 - 2 a, b, c. Protoplasmic thread between two masses of the mycetozoan, *Physarum polycephalum*, forming loops as a result of intrinsic torsion. The photographs were taken at intervals of from 5 to 10 min. Photographs from Kamiya and Seifriz (1954) provided by Dr. N. Kamiya. See p. 175.
 - 3. Pseudopodium of Amoeba proteus photographed between crossed Nicols (with compensation) to show birefringence of the cortex. Unpublished photograph provided by Dr. J. M. Mitchison. See p. 165.
- 15. 1a. Discophrya piriformis before treatment;
 - 1b. after exposure to a pressure of 10,000 lb. in. for 31 min. The cytoplasm is rounded up within the expanded pellicle, and some of the tentacles are now curved. Photographs from Kitching (1954a) provided by Dr. J. A. Kitching, See p. 167.
 - aa. Transverse section of the gut of a frog tadpole, fixed with sublimate and acetic, stained with haematoxylin and orange-G, photographed in the light microscope.
 - 2b. The same between crossed Nicols with compensation (1/16th λ mica-plate). Positive birefringence of the cytoplasm with respect to the long axis of the cell. Photographs from W. J. Schmidt (1943) provided by Professor Schmidt, See p. 176.

- 15. 3. Cytoplasmic nucleic acid or nucleotides not precipitable by centrifugation. Photograph of a paper chromatogram made in ultraviolet light (2,650 A). Regions absorbing light of this wavelength appear black. The experimental spot in the middle of the base-line is the clear intermediate cytoplasmic layer from an axolotl neurula centrifuged at 1,000 g, and the spot to the right of this is from the same cytoplasmic layer recentrifuged at 50,000 g. Both show strong absorption by nucleotides or nucleic acids remaining on the base-line; and from both, spots have moved from the base-line with an R_F of 0·18, calculated from the movement of the adenine control—the large spot on the left near the bottom of the photograph. The strong ultraviolet absorption of the spots from the intermediate cytoplasmic layer confirms Brachet's conclusion that early embryos contain a considerable amount of nucleic acid or nucleotide not precipitable by centrifugation. Unpublished photograph from Goodhart (1950) provided by Dr. C. B. Goodhart. See p. 183.
- 16. I a, b, c. Photomicrographs from a ciné film of a fibroblast from a chick embryo (phase contrast) showing the plasticity of the mitochondria. The time interval between (a) and (b) is 1 min.; between (b) and (c) 1 min. Part of the nucleus and nucleolus is visible on the left-hand margin. Note local and transitory expansions and attenuations, rupture, bending, branching, alteration in refractive index, and displacement relative to the nucleus. The black granules are lipidic droplets. Photographs by J. Frederic and M. Chèvremont (1952) provided by Professor M. Chèvremont. See p. 196.
 - 2a. Transverse section of part of the cortical zone of Stentor polymorphus near the adoral region showing mitochondria with tubular internal structure. Note also the complexity of the pellicle, composed of two paired membranes, the outer pair being well separated from the body cytoplasm between the ciliary rows.
 - 2b. These mitochondria also exist in an 'empty' condition, expanded in volume, and with the tubular sub-elements reduced to a peripheral layer. Both conditions, 'normal' and 'empty', are shown in this photograph. Photographs from Randall and S. F. Jackson (1958) provided by Professor J. T. Randall. See p. 201.
 - 3 a, b, c, d, e. Transformation of thread-like mitochondria to spherical vesicles in a preparation of mitochondria from rat liver isolated in 0.44 M sucrose and progressively diluted with distilled water. Mitochondria prepared by Dr. J. B. Chappell and photographed by Dr. A. V. Grimstone. The stages visible may be compared with the diagram by Cleland and Slater (1953). See p. 197.
- 17. I. Ultra-thin section of the Golgi region of an exocrine cell from the pancreas of a mouse. The upper half of the photograph is occupied by a single mitochondrion surrounded by α-cytoplasmic membranes. A stack of three or four pairs of γ-cytoplasmic membranes (Golgi membranes) partly bounding vacuolar spaces, in the lower half of the photograph, is the characteristic element of the Golgi region. Below and to the right is a uniformly dense zymogen granule, and to the left a Golgi granule of lower opacity and coarser texture than the zymogen granule, and with a dense surface layer. Photograph from Sjöstrand and Hanzon (1954) provided by Professor F. S. Sjöstrand. See pp. 200, 230, 243, and 244.
 - 2. Ultra-thin transverse section of a single parabasal of Trichonympha (Zoomastigina). The supporting filament occupies the cup-shaped cavity in the upper side of the group of saccules. Note the gradual increase in size of the saccules towards the side away from the filament. In the lower right-hand corner is part of the saccules of a second parabasal. Between the two are a few saccules of the system of α -cytomembranes or endoplasmic reticulum. Unpublished photograph provided by Dr. A. V. Grimstone. See p. 238.
- 18. I. Single chromatophore from the egg of *Fucus vesiculosus* seen in ultra-thin section. In this type of chloroplast grana are absent, and the groups of chlorophyll-bearing lamellae extend throughout the substance of the plastid. This structure also occurs in the chloroplasts of the parenchyma sheath in the leaf of *Zea mays*. Photograph from Leyon and von Wettstein (1954) provided by Dr. H. Leyon. See p. 213.

- 18. 2. Part of a mesophyll chloroplast from a 3 to 4 weeks old leaf of Zea mays seen in ultra-thin section. The denser regions with accurately parallel lamellae are the grana. Note the connexion of these lamellae with the extra-granular lamellae embedded in the stroma. The nature of the dense spherical particles has not as yet been established. Photograph from Hodge, McClean, and Mercer (1955) provided by Dr. A. J. Hodge. See p. 213.
 - 3. Ultra-thin section of a young chloroplast of Aspidistra elatior showing a central crystalline body composed of scarcely resolved granules arranged in layers. Note the continuity between the crystal layers and the lamellae of the chloroplast. Photograph from Leyon (1954a) provided by Dr. H. Leyon. See p. 216.
 - 4. Stages in the spontaneous disintegration of chromoplasts of the carrot in aqueous suspension: (a) shows splitting from the edge; (b) suggests the presence of parallel fibrils or strip-like lamellae. Photographs from Straus (1950) provided by Dr. W. Straus. See p. 220.
- 19. Ia. Phase-contrast photograph (light contrast) of the spindle in a living Barbulanympha (Zoomastigina). The spindle has formed between voluminous asters arising at the ends of ribbon-like centrioles. Extra-spindle fibres extend towards the nuclear membrane. Unpublished photograph provided by Professor L. R. Cleveland. See pp. 260, 265.
 - Ib. Phase contrast photograph (dark contrast) of chromosomes in *Barbulanympha* attached to the nuclear membrane by their telomeres. Unpublished photograph provided by Professor L. R. Cleveland. See p. 261.
 - 2. Mitotic figures of *Psammechinus miliaris* in polarized light with compensation showing the development of asters and spindle during first cleavage. The first photograph (a) was taken at 50 min after fertilization; the rest succeed as follows: (b) 52 min; (c) 54 min; (d) 55 min; (e) 56 min 20 sec; (f) 57 min; (g) 58 min; (h) 59 min. Photographs from Swann (1951a, b) provided by Professor M. M. Swann. See pp. 264, 265.
 - 3. Micro-X-ray photographs obtained from the several levels of a single human hair root with the X-ray beam at right angles to the axis of the hair and at a specimen to film distance of 1 cm: (a) from the bulb; no fibrillar orientation; (b) immediately above the bulb; fully developed a-keratin pattern, although this region is unconsolidated; (c) the same region after disorientation by heating; the pattern has changed to the disoriented β -keratin type; (d) the a-keratin pattern of the fully hardened, S-S-bonded hair above (c). Photographs from Mercer (1949a) provided by Dr. E. H. Mercer. See p. 277.
- 20. I. Transverse section through the rostral region of Trichonympha showing sections of the flagella lying between two cytoplasmic plates. In these flagella the outer ring of nine fibrils is tangentially double. The nature of the cytoplasmic inclusions has not been established. Unpublished photograph provided by Dr. A. V. Grimstone (1956). See p. 271.
 - 2. Slightly oblique transverse section through the triple row of kinetosomes (basal granules or bodies) associated with the cilia of a single membranelle from the adoral zone of Stentor polymorphus (Ciliata, Spirotricha, Heterotricha). Sections of the basal bodies in the upper part of the photograph are more distal, those below more proximal, in level. A distinct central particle (p) is visible in some of the former; on this the central filaments of the cilium terminate. The particle rests on a septum (s), visible in some of the sections. Below this, the kinetosome appears to consist of the outer filaments of the cilium. Photograph from Randall and S. F. Jackson (1958) provided by Professor J. T. Randall. See p. 273.
 - 3. Section parallel to the plane of the scopula in Campanella umbellaria (Ciliata, Peritricha), passing through the array of kinetosomes (basal bodies), of which the peripheral members are arranged in regular concentric rows, while the central members are distributed at random. Each of the regularly arranged basal bodies gives rise to a cilium, c. 10 μ long, which appears to be surrounded by a fine tubule, the walls of which consist of filaments, 50 to 100 A in diameter. Each tubule seems to be a

- prolongation of the surface membrane of a cilium. Photograph from Rouiller, Fauré-Fremiet, and Gauchery (1956) provided by Professor E. Fauré-Fremiet. See p. 273.
- I. Isolated fusiform cortical cells of wool. Photograph provided by the Textile Physics Laboratory of the University of Leeds. See p. 278.
 - 2. Transverse section of portions of two cells from the cortex of the upper bulb region of a human hair, showing early filaments of keratin. Photograph from Birbeck and Mercer (1957a) provided by Dr. E. H. Mercer. See p. 278.
 - 3. Transverse section of part of a cell from the upper follicle cortex of a human hair, showing late bundles of keratin filaments. Unpublished photograph provided by Dr. E. H. Mercer. See p. 278.
 - 4. Gold-shadowed protofibrils of keratin obtained by enzymic disintegration of wool fibres, showing the protofibrils as strings of corpuscles, each corpuscle rather more than 100 A wide. Photograph from Farrant, Rees, and Mercer (1947) provided by Dr. E. H. Mercer. See p. 278.
- I. X-ray diffraction pattern of α-keratin, fibre axis parallel to short edge of plate. See p. 283.
 - 2. X-ray diffraction pattern of β -keratin, orientation as before. See p. 283.
 - 3. So-called 'cross- β ' X-ray diffraction pattern of keratin, orientation of specimen as before, but the 'backbone' spacing appears on the meridian. See p. 280.

Photographs 1, 2, and 3 provided by the Textile Physics Laboratory of the University of Leeds.

- 4. A single myofibril from a glycerol-extracted rabbit's psoas muscle, photographed under 'even-field' illumination in a Cooke-Dyson interference microscope, (left) before, (right) after extraction of myosin; together with densitometer tracings. Photographs from H. E. Huxley and Hanson (1957) provided by Dr. Jean Hanson. See p. 299.
- 5. Birefringence of the fresh axon of a fibre from a frog's sciatic nerve (R. pipiens or R. catesbiana). The bare axon protrudes from the brilliantly luminous sheath, visible on the left. Photograph from Thornburg and de Robertis (1956) provided by Dr. W. Thornburg. See p. 286.
- 23. Ia. Longitudinal section of developing tergosternal muscle from a young pupa of Tenebrio molitor. The myofilaments occur in strands consisting of a few filaments only, relatively widely spaced, not accurately parallel, and without transverse striation.
 - 1b. Longitudinal section of the same muscle from a somewhat older pupa. The strands now contain many more filaments in close parallel array but still without transverse striction.
 - 1c. Longitudinal section of material from a late pupa showing striated myofibrils with strongly opaque Z-lines delimiting the sarcomeres.
 - (a), (b), (c), unpublished photographs provided by Dr. D. S. Smith. See p. 294.
 - 2a. Section through a single sarcomere of glycerinated psoas muscle from a 'rabbit parallel to the 1120 plane, showing primary filaments with large interfilament spacing (c. 500 A) and pairs of secondary filaments between. Note the interruption of the secondary filaments at the H-zone, the cross bridges between primary and secondary filaments, the tapering of the primary filaments at the ends of the A-bands, and the thickening of the primary filaments in the H-zone. Photograph from H. E. Huxley (1957) provided by Dr. H. E. Huxley. See pp. 302, 304.
 - 2b. Cross-section through myofibrils of the same material. In the sections to the left and above, the plane of section passes through the H-zone; in those to the right and below, through the A-region. In the latter the interpenetrating hexagonal arrays of primary and secondary filaments can be seen. Unpublished photograph provided by Dr. H. E. Huxley. See p. 299.