

Handbook of LEGUNES of World Economic Importance

James A. Duke

United States Department of Agriculture Beltsville, Maryland

Plenum Press · New York and London

Library of Congress Cataloging in Publication Data

Main entry under title:

Handbook of legumes of world economic importance.

Includes index.

1. Legumes. I. Duke, James A., 1929-

SB317.L43H36 633.3

ISBN 0-306-40406-0

80-16421

© 1981 Plenum Press, New York A Division of Plenum Publishing Corporation 227 West 17th Street, New York, N.Y. 10011

All rights reserved

No part of this book may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, microfilming, recording, or otherwise, without written permission from the Publisher

Printed in the United States of America

Credits

- Acad. Repubc. Pop. Rom. 5, Bucharest, Rumania, 1957: Fig. 115.
- Ali, S. I., Flora of West Pakistan, Third Annual Report, Karachi, Pakistan: Fig. 26.
- Ali, S. I., Flora of West Pakistan 100, 1977: Figs. 51, 134, 136, 137.
- Ann. Jard. Bot. Buitenzorg 44, 1935: Fig. 29.
- Aublet, J. B. C. F., Histoire des Plantes de la Guiane Française, Vol. 4, Didot, London/Paris, 1775: Figs. 61, 108.
- Basu, B. D., Indian Medicinal Plants, Allahabad, India, 1918 (reprinted 1975): Figs. 24, 103, 142.
- Bentham, G., Handbook of the British Flora, Vol. 1, Reeve, London, 1865: Fig. 125.
- Berg. Char. Pflanzen Gattungen, Gaertner, Berlin, 1861: Fig. 41.
- Chuang, C.-C., and Huang, C., The Leguminosae of Taiwan for Pasture and Soil Improvement, Taipei, Taiwan, 1965: Figs. 70, 83, 139, 141.
- Constantin, P., Marvelles de la Nature. Les Plantes, Baillera, Paris, 1894-1896: Fig. 3.
- Corner, E. J. H., and Watambe, K., Illustrated Guide to Tropical Plants, Hirohawa, Tokyo, 1969: Fig. 19.
- Coste, H. J., Flora France, Vol. 1, 1901: Fig. 79.
- Curtis. Bot. Mag. 3, 1789: Fig. 105.
- Curtis. Bot. Mag. 139, 1913: Fig. 38.
- Dodson, C. H., and Gentry, A. H., Flora of the Rio Palenque Science Center, Los Rios, Ecuador, The Marie Selby Botanical Gardens, Sarasota, Florida, 1978: Figs. 40, 90.
- Duke, P. (tracings of xerocopies): Figs. 8, 13, 20, 34, 44, 54, 69, 88, 107, 111, 120.
- Engler, H. G. A., Pflanzenfam: Figs. 11, 15, 27, 52, 82.
- Engler, H.G.A., and Drude, C. G. O., Veget. der Erde, 1911: Fig. 17.
- Farmers Bull. 693, 1915: Fig. 73.
- Faul, K. (drawing): Fig. 85.
- Flora of Taiwan, Vol. 3, Epoch, Taiwan, 1977: Figs. 1, 9, 14, 16, 18, 25, 28, 30, 33, 35, 36, 43, 49, 50, 57, 58, 59, 60, 63, 67, 68, 75, 81, 89, 97, 99, 101, 102, 110, 122, 123, 145.
- Freeman, G. F., Bot. Gaz. 56, 1913: Fig. 92.
- Garcke, C. A. F., Illustrated Flora Deutschland, 17th ed., 1895; Fig. 10.
- Grieve, M., and Leyel, C. F., A Modern Herbal, Vol. 1, 1937: Fig. 86.

- Gunn, C. R., Classification of *Medicago sativa* L. using legume characters and flower colors, *U.S. Dep. Agric.*, *Tech. Bull.* **1574**, 1978; Fig. 74.
- Hegi, G., Illus. Flora Mittel. Europ., Munich, 1923-1924: Fig. 129.
- Hogg, P., and Johnson, C. W., Wild Flowers of Great Britain, Vol. 8, London, 1874: Fig. 80.
- Humboldt, A., Bonpland, A., and Kunth, C. S., Nova Genera et Species Plantarum, 1823: Fig. 109.
- Iowa Geol. Surv. Bull. 4, Weed flora of Iowa, Des Moines, 1913: Fig. 46.
- Irvine, F. R., Woody Plants of Ghana, Oxford University Press, London, 1961: Figs. 5, 12, 48, 91.
- J. Linn. Soc. London, Bot. 13, 1871-1873: Fig. 72.
- Karsten, H., Disch. Flora. Pharm. Med. Botanik, Spaeth, Berlin, 1880-1883: Fig. 6.
- Kew Bull., 1912: Fig. 71.
- Killip, E. E., and Smith, A. C., The identity of South American fish poisons, J. Wash. Acad. Sci. 20(5), 1930: Fig. 62.
- Kotschy, T., De plantis nilotico—Acthiopicis Knoblecheriana, 1864-1865: Fig. 32.
- Lindley, J., Med. Econom. Bot., 1849: Fig. 45.
- Maiden, J. H., The Forest Flora of New South Wales, Vol. 3, Government Printers, Sydney, Australia, 1907: Fig. 2.
- Maiden, J. H., Wattles and Wattles Barks, 2nd ed., Government Printers, Sydney, Australia, 1891: Fig. 4.
- Moris, G. G., Flora Sardoa Iconographic, 1837: Fig. 76.
- Notizblatt Bot. Gart. Mus. Berlin. Dahlem., 1909: Figs. 98, 100.
- Notizblatt Bot. Gart. Mus. Berlin. Dahlem., 1911: Fig. 106.
- Reed, C. F., Selected Weeds, 1970: Fig. 135.
- Reichenbach, H. G., and Beck, G., Icon. Fl. Germ. Helv. 22, 1903: Figs. 42, 47, 65, 87, 113, 116, 117, 127.
- Schlechtendahl, D. F. L. von, et al., Flora von Deutschland, 5th ed., Bd. 23, 1880; Fig. 121.
- Schlechtendahl, D. F. L. von, et al., Flora von Deutschland, Bd. 24, 1880: Figs. 130, 132, 133.
- Smartt, J., Tropical Pulses, Longman Group Ltd., London, 1976: Figs. 143, 146.
- Syme, J. T., et al., English Botany, Vol. 3, 3rd ed., Hardwicke, London, 1864: Fig. 64.
- Taylor, T. M. C., The pea family of British Columbia, B.C. Prov. Mus. Handb. 32, 1974: Fig. 126.

Credits

- Trans. Linn. Soc. N.Y. 29, 1873-1875: Fig. 112. U.S. Dep. Agric., Bull. 119, 1914: Fig. 138.
- Westphal, E., Pulses in Ethiopia. Their Taxonomy and Agricultural Significance, Centre for Agricultural Publishing and Documentation, Wageningen, 1974: Figs. 53, 55, 56, 66, 84, 93, 94, 95, 96, 128, 131, 140, 144.
- Wight, R., Icon. Pl. Ind. Orientalis 1, 1840: Figs. 23, 37.
- Wight, R., Icon. Pl. Ind. Orientalis 3, 1843-1845: Fig. 21.
- Wilbur, R. L., The Leguminous Plants of North Carolina, The North Carolina Agricultural Experiment Station, 1963: Figs. 78, 104, 118.
- Zohary, M., PL-480 Project A-10-CR-11, 1967: Figs. 114, 119, 124.

Contributors

Dr. J. C. Baudet Faculte des Sciences Agronomique de L'Etat 5800 Gembloux, Belgium

Dr. T. E. Boswell Texas A. & M. University Plant Disease Research Station Yoakum, Texas 77995

Dr. J. L. Brewbaker University of Hawaii Honolulu, Hawaii 96822

Dr. A. H. Bunting
Plant Science Laboratory, University of
Reading
White Knights
Reading, England
RG62AF

Dr. W. V. Campbell
Associate Professor of Insect Pathology
The School of Agriculture and Life
Sciences and Forest Resources
North Carolina State University
Raleigh, North Carolina 27607

Dr. S. Chandra Senior Plant Breeder Division of Genetics and Plant Physiology Central Soil Salinity Research Institute Karnal, 132001 India

Dr. C. S. Cooper Plant Science Research Division United States Department of Agriculture Bozeman, Montana 59715 Dr. W. A. Cope Research Leader USDA-SEA-FR-SR North Carolina State University P.O. Box 5155 Raleigh, North Carolina 27607

Dr. S. Dana
Bidhan Chandra Krishi
Vistva Vidyalaya
Faculty of Agriculture
Department of Genetics and Plant
Breeding
Kalyani 741235
West Bengal, India

Dr. T. E. Devine Room 218, Building 001, BARC-West Department of Agriculture Beltsville, Maryland 20705

Dr. Tricia Dodd

Plant Science Laboratory, University of
Reading
White Knights
Reading, England
RG62AF

Dr. J. A. Duke Chief Economic Botany Laboratory Building 265, BARC/East United States Department of Agriculture Beltsville, Maryland 20705

Dr. N. R. Farnsworth College of Pharmacy University of Illinois 833 S. Wood Street Chicago, Iuinois 60612

Dr. M. B. Forde
Plant Introduction Office
Grasslands Division
DSIR, Private Bag
Palmerson North, New Zealand

Dr. W. Foulds Science Department Claremont Teachers College West Australia

Mr. A. Fyson Department of Biological Sciences The University Dundee, United Kingdom

Dr. P. B. Gibson Agronomy Department Clemson University Clemson, South Carolina 29631

Dr. M. A. Golden Room 159, Building 011-A United States Department of Agriculture Beltsville, Maryland 20705

Dr. H. J. Gorz University of Nebraska Lincoln, Nebraska 68503

Dr. Peter H. Graham CIAT, Apartado Aereo 6713 Cali, Colombia

Dr. W. C. Gregory Professor of Genetics The School of Agriculture North Carolina State University Raleigh, North Carolina 27607

Dr. Ray O. Hammons Research Leader USDA-SEA-FR-SR Georgia Coastal Plains Experiment Station Tifton, Georgia 31794

Contributors

Dr. R. Hegnauer Laboratorium and Voor Experimenele Plantensystematiele Schelpenkade 14a 2131-2T Leiden Netherlands

Dr. J. M. Hopkinson Agricultural Branch Department of Primary Industries Brisbane Old Australia 4000

Dr. C. S. Hoveland Auburn University Auburn, Alabama 36830

Dr. E. M. Hutton
Chief of Tropical Crops and Pastures
Commonwealth Scientific and Industrial
Research Organization
St. Lucia, Queensland, Australia

Dr. T. Hymowitz University of Illinois at Urbana-Champaign Urbana, Illinois 60801

Dr. R. F. Keeler Utah State University UMC 48, Agricultural Science Building Logan, Utah 84322

Dr. T. N. Khan Department of Agriculture Jarrah Road South Perth, 6151 Western Australia

Dr. A. E. Kretschmer, Jr. University of Florida Institute of Food, Agricultural Sciences Agricultural Research Center Fort Pierce, Florida 33450

Dr. J. Langenheim University of California Botany Department Santa Cruz, California 95053

Dr. R. M. Lantican
University of the Philippines at
Los Banos
College of Agriculture
College, Laguna, 3720
Philippines

Dr. J. M. Lenne University of Florida Institute of Food, Agricultural Sciences Agricultural Research Center Fort Pierce, Florida 33450

Dr. R. L. Lynch

Dr. L. 't Mannetje CSIRO Mill Road St. Lucia QLD. Australia 4067

Dr. R. Marechal Faculte Des Sciences Agronomiques De L'Etat 5800 Gembloux Belgium

Dr. J. D. Miller Agronomy Department Georgia Coastal Plains Experiment Station Tifton, Georgia 31794

Dr. A. J. Norden
Institute of Food and Agriculture
Sciences
College of Agriculture
University of Florida
304 Newell Hall
Gainesville, Florida 32611

Dr. B. N. Okigbo
Director
International Institute for Tropical
Agriculture
Ibadan, Nigeria

Dr. B. P. Pandya Agricultural Station Pantnager, India

Ms. Hazel Pollard DSAD, National Agricultural Library Beltsville, Maryland 20705

Dr. K. O. Rachie CIAT, Apartado Aereo 6713 Cali, Valle, Colombia

Dr. C. F. Reed Reed Herbarium 10105 Harford Road Baltimore, Maryland 21234

Dr. Peter P. Rotar
Department of Agronomy and Soil
Science, CTA
University of Hawaii at Manoa
3190 Maile Way
Honelulu, Hawaii 96822

Dr. D. G. Roux Department of Chemistry University of The Orange Free State P.O. Box 339, Bloemfontein 9300 Republic of South Africa Dr. N. H. Shaw CSIRO Division of Tropical Crops and Pastures St. Lucia, Queensland Australia

Dr. C. E. Simpson
Texas A. & M. University
Agricultural Research and Extension
Center
Stephenville, Texas 76401

Dr. R. P. Sinha

Dr. A. E. Slinkard Senior Research Scientist Crop Science Department University of Saskatchewan Saskaton, Saskatchewan Canada S7N 0W0

Dr. J. Smartt Department of Biology University of Southampton 509 5NH United Kingdom

Dr. Donald H. Smith Associate Professor Texas A. & M. University Plant Disease Research Station Yoakum, Texas 77995

Dr. Olin D. Smith Associate Professor of Peanut Breeding Soil and Crop Science Texas A. & M. University College Station, Texas 77843

Dr. R. R. Smith USDA, ARS, North Central Region Department of Agronomy University of Wisconsin Madison, Wisconsin 53706

Dr. J. I. Sprent Department of Biological Sciences The University Dundee, United Kingdom

Dr. R. J. Summerfield Plant Environmental Laboratory Shinfield Grange Cutbush Lane, Shinfield Reading, Berks United Kingdom

Dr. N. L. Taylor University of Kentucky Agronomy Department College of Agriculture University of Kentucky Lexington, Kentucky 40506 Dr. C. E. Townsend Crops Research Laboratory Colorado State University Bay Road south of West Prospect Fort Collins, Colorado 80523

Dr. L. J. G. van der Maesen Pulse Germplasm Botanist 1-11-256 Begumpet Hyderabad, 500016 A. P. India

Dr. Jurgen K. P. Weder Institut FUR Lebensmittelchemie Technische Universitat Munchen 8 Munchen 2 Lothstrasse 17 Nese Ruf-NR. 2105-4246 West Germany Dr. E. B. Whitty
Associate Professor of Field Crop
Management
Agronomy Department, 304 Newell Hall
University of Florida
Gainesville, Florida 32611

Dr. E. C. Williams

Dr. M. C. Williams
University of Agriculture and Applied
Sciences
Logan, Utah 84322

Dr. J. C. Wynne Crop Science Department North Carolina State University Raleigh, North Carolina 27607

Dr. Clyde T. Young Food Science Department Georgia Agricultural Experiment Station Experiment, Georgia 30212

Dr. D. M. Zohary Department of Genetics Hebrew University Jerusalem, Israel

Contents

Credits vii
Contributors ix
Introduction 1
Format 1
Text Abbreviations 3
Collaborators 4
Legume Species
Appendix 311
TABLE 1. Legume "Toxins": Their Toxicity and Generic
Distribution 311
TABLE 2. Legume Genera and Their Toxins 314
TABLE 3. Ecosystematic Attributes of Legumes 317
TABLE 4. Economic Legumes: Their Tolerances, Yields, Centers of
Diversity, and Ecocenters 321
TABLE 5. Recommended Inoculants for Various Legumes 327
TABLE 6. Zero-Moisture Nutritional Analysis of Legumes 328
TABLE 7. Amino Acid Composition of Various Legumes
(g/16 g N) 334
General References

Introduction

In 1971, Dr. Quentin Jones, now of the National Program Staff, SEA, USDA, suggested that the Plant Taxonomy Laboratory devise a format for concise write-ups on 1,000 economic plants (Duke and Terrell, 1974; Duke et al., 1975). Dr. C. F. Reed was contracted to search the literature on these economic plants, which included 146 species of legumes. From 1971 through 1974, Dr. Reed prepared rough drafts of write-ups on the 1,000 species. It was my responsibility to establish the format and monitor the write-ups, to ensure that they would answer many questions on legumes directed to the USDA by our taxpaying public. Since then, a computerized system alerts me to new publications on legumes. I have ordered for our files copies of the more promising documents. With the evolution of the manual, many later references supplanted earlier ones. Some repetitive information was dropped by condensing the write-ups on secondary species within the genera.

Informally in 1975 and formally in 1976, Dr. Roger Polhill, Curator of Legumes, Royal Botanical Gardens, Kew, joined by Dr. A. S. Bunting, Professor, The University of Reading, and Dr. J. P. M. Brenan, Director, Royal Botanical Gardens, Kew, conceived the splendid idea of an International Legume Conference at Kew. We agreed that this would be a good forum for presentation of the Handbook of Legumes. Beginning in 1976, I circulated second drafts of the individual legume write-ups to a mailing list of contributors. Responses ranged from obloquy to praise; many contributors went over the drafts carefully.

I also benefited from attending a National Academy of Science Legume Conference in Maui.

Hawaii, where an international panel convened to discuss and assemble information on underexploited tropical legumes. Conversations at that meeting and subsequent correspondence with the participants also yielded new information on some of the tropical legumes.

Finally in 1978, 100 copies of the writeups were delivered to the International Legume Conference at Kew, July 24th-August 4, and all were given to potential cooperators before my lecture on the manual (July 31st). New information presented in lectures at that conference and personal communications behind the scenes have also been used to update and embellish the write-ups so that they are more than a bibliographic echo

On May 7, 1978, I was transferred from USDA's Plant Taxonomy Laboratory to the Medicinal Plant Resources Laboratory (now the Economic Botany Laboratory). Since then, I have included the Folk Medicine paragraph in the final format. My computerized ecosystematic program (Duke, 1978b) includes data not only on legumes, but on economic plants in general, many of which are intercropped with legumes to good advantage.

FORMAT

Except for beginning each write-up under the heading of scientific name, there was little unanimity about what should be included or excluded, and in what order the included paragraphs should be arranged. I have, without sacrificing clarity, adopted a terse style, often leaving out the verbs in descriptive sentences. Furthermore, I have gener-

ously sprinkled the text with abbreviations that are identified in the following section titled Text Abbreviations. It should be noted that these abbreviations may differ from those specifically defined in tables found in the Appendix. I finally elected to follow the format below, at least for the first species in each genus.

NOMENCLATURE: All agreed that the scientific name, authority, and recent significant synonyms be included. Because Anglicans consistently use the word "lucerne" for the plant Americans call "alfalfa," and because this manual was intended for international usage, common names in a few major languages are included. However, the large number of common names dictated that most of them be omitted.

Uses: Following the Nomenclature Section, important general uses are reported. Some of the uses that may seem trivial in developed countries are important in developing countries, and vice versa.

FOLK MEDICINE: Folk medicine is separated from general uses, even though many collaborators believe that the information is parascientific at best. Frequently, however, folk medical usage is related to chemical characteristics. Certainly the USDA does not recommend folk medicine, and discourages self-medication. Still, I quote Dr. R. J. Mc-Cracken from Perdue and Hartwell (1975). "We have heard much lately about the use of plants and the products derived from them, in meeting world food problems. But of equal concern and importance is human health and in this respect plants have much to contribute that is of therapeutic value." In the cancer-screening program of the National Institutes of Health, several compounds found in legumes have proved active against experimental cancers. Cassia obtusa has been reported to give the active colubrinol: Cercidium microphyllum has given an active; Coronilla varia gave hyrcanoside: Crotalaria spectabilis, monocrotaline; Derris trifoliata, rotenone; Entada phaseoloides, an active; Lonchocarpus urucu, rotenone; and Piscidia erythrina, rotenone (Perdue and Hartwell, 1975). Folklore on medicinal plants is a good guide in our quest for new medicinal chemicals.

CHEMISTRY: I did not include detailed analytical data on the chemistry of legumes but included nutritional data, where available. Often approxi-

mate nutritional composition varied widely among sources. In the individual chapters I have reported the nutritional data from the various sources listed in the references. Many of these data were recently published (Duke 1977a). To facilitate comparison, I tabulated nutrients (zero-moisture basis) in Table 6 and amino acid compositions in Table 7 of the Appendix. I also list chemurgic chemicals and/or those cited as "toxic" (NIOSH, 1975). There has been discussion in the United States urging that plants containing toxins be so labeled. Consequently they were included in Appendix Tables 1 and 2, derived from my Phytotoxin Tables (Duke, 1977b). Many correspondents questioned the merit of including mere generic lists. I have initiated a computer program to accommodate the quantitative distribution of chemicals within the species and the location in the plant, rather than in the genera. That should prove even more useful than the Phytotoxin Tables.

DESCRIPTION: The manual was designed more specifically for agronomists than for taxonomists, although I hope both groups will find it useful. Simple terms were sought for the descriptions. Line drawings have been borrowed from the open literature where possible.

GERMPLASM: Space limited the discussion of germplasm to the more important cultivars, comments on the centers of origin, breeding mechanisms, tolerances, and chromosome counts.

DISTRIBUTION: Modern distribution of many economically important legumes bears little relation to their historic distribution. In this section I tried to summarize facts about current and historic distribution.

ECOLOGY: For more than 5 years, I have solicited local floral checklists associated with ecological data. The resultant data complement other published data. Ecological data from my computerized ecosystematic data base (Duke, 1978a,b,c) on legumes are tabulated in the Appendix (Table 3). If Table 3 represented the entire world rather than a small biased sample, the climate and pH of a remote area might be estimated by a study of its native legumes. The concluding sentence of the Ecology Section with data on life zones, annual precipitation, annual temperature, and pH is derived from my ecosystematic program; the data were con-

firmed by my computer program. Some data may differ from those published but each datum was based on a case history that is retrievable by computer.

CULTIVATION: Here I included information for developed "mechanized" societies, for developing "manual" societies, or for both, on land preparation, planting, fertilization, intercropping, cultivation and maintenance of the legume until harvest. Some data on field spacing and seed rates were often included, but these vary so much from system to system, that I reported only a few.

HARVESTING: Methods for harvesting were reported, often with notes on processing the harvested plant. Data on optimum stage of growth and conditions for harvest also were included, where available.

YIELDS AND ECONOMICS: Conventional data for yields were reported. Data for biomass, nitrogen fixation, and any other available nonconventional estimates of yield also were included. Dollar figures, where possible, are tied to dates so that this manual would not immediately lose its comparative value.

BIOTIC FACTORS: Pests, at least some of the more important ones, were listed. I consulted the historic file maintained in the U.S. National Fungus Collection at Beltsville, and was assisted editorially by many of my colleagues in other disciplines at Beltsville. However they can be held responsible only for the correct names. I found no one with time enough to check out all the names recorded in the National Fungus Collection, hence many synonyms have crept in. I would rather have a species listed under an outdated name than not listed at all. For conservation of space, the authors of the species names of the various pests have been deleted. Occasionally, I have listed pesticides in this section. This section is strictly bibliographic, and the lists do not constitute a recommendation or endorsement of these pesticides by the USDA. As a matter of fact, some of them are prohibited in the US and many are dangerous, unless used strictly according to the instructions.

SPECIFIC REFERENCES: Many correspondents have pointed out several recent references, too many for inclusion. General familial or generic references

consulted in early drafts or in final revisions have been placed in the reference citations of the section titled General References. Even though these works were consulted in preparing individual write-ups, they are listed only in the final general collection of references. For inclusion with each write-up, preference has been given to specific references or those dated 1975 or later, especially review papers. General references were listed only at the end of the manual; a few specific references were listed at the end of most chapters.

were listed at the end of most chapters.						
	TEXT ABBREVIATIONS					
	ann	annum, annual				
	Apr.	April				
	Aug.	August				
	avg.	average				
	AVRDC	The Asian Vegetable Research				
		and Development Center				
	В	boron				
	B.C.	before Christ				
	BP	before present				
	Br	bromine				
	bu	bushel				
	BYMV	bean yellow mosaic virus				
	°C	degrees centigrade (Celsius)				
	Ca	calcium				
	ca.	about, nearly				
	cf.	compare with				
	CIAT	Centro Internacional de				
	CI.	Agricultura Tropical				
	Cl	chlorine				
	cm	centimeter				
	Cu	copper				
	cu	cubic				
	cv	cultivar				
	cvgr	cultigroups				
	cwt	hundredweight				
	Dec.	December				
	DM	dry matter				
	dm EC	decimeters				
		electrical conductivity				
	e.g. et al.	for example and others				
	etc.	and so forth				
	FAO	Food and Agricultural Organization				
	Ino	of the United Nations				
	FDA	Federal Drug Administration				
	Fe	iron				
	Feb.	February				

Fig.

figure

Introduction

Introduction						
Fl.	flower	prec	precipitation			
Fr.	fruit	. R	reproductive			
g	gram	S	sulfur			
ha	hectare	SA	South America			
HCN	hydrogen cyanide	Sep.	September			
hr	hour	sp.	species (singular)			
I	iodine	spp.	species (plural)			
i.e.	that is	sq	square			
IITA			subspecies (singular)			
	Agriculture	ssp. sspp.	subspecies (plural)			
IU	International Unit	temp.	temperature			
Jan.	January	μ g	microgram			
K	potassium	U.K.	United Kingdom			
kg	kilogram	USDA	United States Department of			
km	kilometer		Agriculture			
lb	pounds	USSR	Russia			
LAI	Leaf Area Index	V	vegetative			
m	meter	var	variety			
MD	Maryland	WARF	Wisconsin Alumni Research			
Mar.	March		Foundation			
max.	maximum	WI .	West Indies			
Mg	magnesium	WM	wet matter (as opposed to DM)			
mg	milligram	WOI	Wealth of India			
ml	milliliter	wt.	weight			
mm	millimeter	yr	year			
min	minute					
min.	minimum	Collaborators: Perhaps the most important item in				
Mn	manganese	each chapter is the list of contributors. Many spe-				
Mo			cialists have contributed and some lists may be			
mos.	months	incomplete. I wish, however, to thank R. F.				
MT	metric ton	Barnes, L. R. Batra, T. E. Boswell, J. L. Brew-				
N	nitrogen	baker, K. R. Bromfield, A. H. Bunting, J. C.				
n as			Burton, W. V. Campbell, S. Chandra, W. A. Cope,			
in 2			T. C. Davidson, T. E. Devine, D. Elling-			
Na	sodium		ton, N. R. Farnsworth, M. B. Forde, A. Fyson, M.			
NC	cultigroups	Golden, H. J. Gotz, P. H. Graham, W. C. Gregory,				
Ni	nickel	C. R. Gunn, R. O. Hammons, R. Hegnauer, C. C.				
NIOSI	•	Heyn, E. M. Hoover, C. S. Hoveland, E. M.				
NT TT	Safety and Health	Hutton, T. Hymowitz, T. J. Kahn, R. F. Keeler, L.				
N.H.	Northern Hemisphere	Knutson, A. E. Kretschmer, Jr., J. Langenheim,				
No.			R. M. Lantican, R. L. Lynch, R. Marechal, J. D.			
Nov.			Miller, A. J. Norden, B. N. Okigbo, B. P. Pandya,			
NPK	nitrogen: phosphorus: potassium	R. M. F	Polhill, H. Pollard, K. O. Rachie, C. F.			

Reed, P. P. Rotar, D. G. Roux, C. E. Simpson, A.

E. Slinkard, J. Smartt, D. H. Smith, O. D. Smith,

J. I. Sprent, R. J. Summerfield, R. A. Taber, N. L.

Taylor, E. E. Terrell, C. E. Townsend, L. J. G.

van der Maesen, J. K. P. Weder, E. B. Whitty, E.

C. Williams, M. C. Williams, J. C. Wynne, C. T

Young, and D. Zohary.

Oct.

p.

pp. P

PA

PI

ppm

October

phosphorus

Pennsylvania

plant introduction

parts per million

page

pages

Legume Species

Acacia farnesiana (L.) Willd.

FAMILY: Mimosaceae

COMMON NAMES: Cassie, Huisache Synonym: Mimosa farnesiana L.

Uses: Cassie perfume distilled from the flowers. Cassie absolute used in preparation of violet bouquets, extensively used in European perfumery. Cassie pomades manufactured in Uttar Pradesh and

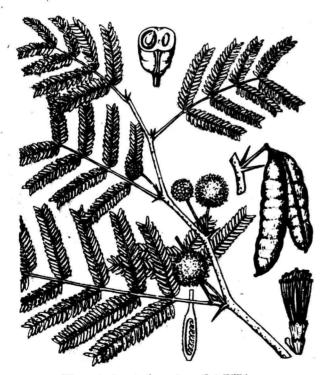


Figure 1. Acacia farnesiana (L.) Willd.

the Punjab. Pods contain 23% tannin, a glucoside of ellagic acid, and are used for tanning leather. Bark also used for tanning and dyeing leather in combination with iron ores and salts. In Bengal and West Indies pods used for a black leather dye. Gummy substance from pods used in Java as cement for broken crockery. Gum exuding from trunk considered superior to gum arabic in arts. In Ivory Coast trees used as ingredient in arrow poison; elsewhere used as fences and to check erosion. Wood is hard and durable underground, used for wooden plows and for pegs. Often planted as an ornamental.

FOLK MEDICINE: Bark is astringent and demulcent, and with leaves and roots is used for medicinal purposes. Woody branches used in India as tooth-brushes; gummy roots chewed for sore throat. Said to be used as alterative, antispasmodic, aphrodisiac, astringent, demulcent, antidiarrhetic, febrifuge, antirheumatic, and stimulant.

CHEMISTRY: Dried seeds of one Acacia sp. reported to contain per 100 g: 377 calories, 7.0% moisture, 12.6 g protein, 4.6 g fat, 72.4 g carbohydrate, 9.5 g fiber, and 3.4 g ash. Raw leaves of Acacia contain per 100 g: 57 calories, 81.4% moisture, 8.0 g protein, 0.6 g fat, 9.0 g carbohydrate, 5.7 g fiber, 1.0 g ash, 93 mg Ca, 84 mg P, 3.7 mg Fe, 12,255 μ g β -carotene equivalent, 0.20 mg thiamine, 0.17 mg riboflavin, 8.5 mg niacin, and 49 mg ascorbic acid. Reporting 55% protein on a dryweight basis, Van Etten et al. (1963) break down the amino acids as follows: lysine, 4.7 (g/16 g N); methionine, 0.9;

arginine, 9.2; glycine, 3.4; histidine, 2.3; isoleucine, 3.5; leucine, 7.5; phenylalanine, 3.5; tyrosine, 2.8; threonine, 2.5; valine, 3.9; alanine, 4.3; aspartic acid, 8.8; glutamic acid, 12.6; hydroxyproline, 0.0; proline, 5.1; serine, 4.1; with 76% of the total nitrogen as amino acids. Cassie has been reported to contain anisaldehyde, benzoic acid, benzyl alcohol, butyric acid, coumarin, cresol, cuminaldehyde, decyl aldehyde, eicosane, eugenol, farnesol, geraniol, hydroxyacetophenone, methyleugenol, methyl salicylate, nerolidol, palmitic acid, salicylic acid, and terpineol. The genus Acacia is reported to contain: anhaline, anisaldehyde, anthraquinone, benzaldehyde, benzyl alcohol, butyraldehyde, cresol, cuminic aldehyde, decanal, dimethyl tryptamine, eugenol methyl ether, gallic acid, heptanoic acid, hydrocyanic acid, indole, isobutyraldehyde, linalool, methyl salicylate, nicotine, palmitic acid, phenethylamine, quercitin, rutin, saponin, tannic acid, terpineol, trigonelline, tyramine (for relative toxicities, see Appendix Table 2). Seeds of the genus Acacia reported to contain trypsin inhibitors and chymotrypsin inhibitors.

DESCRIPTION: Thorny bush or small tree, 1.5 m tall; bark, light brown, rough; branches glabrous or nearly, purplish to gray, with very small glands; stipules spinescent, usually short, up to 1.8 cm long, rarely longer, never inflated; leaves twicepinnate, with a small gland on petiole and sometimes one on the rachis near top of pinnae; pinnae 2-8 pairs, leaflets 10-12 pairs, minute, 2-7 mm long, 0.75-1.75 mm wide, glabrous, leathery; flowers in axillary pedunculate heads, calvx and corolla glabrous, scented; pod indehiscent, straight or curved, 4-7.5 cm long, about 1.5 cm wide, subterete and turgid, dark brown to blackish, glabrous, finely longitudinally striate, pointed at both ends; seeds chestnut brown, in 2 rows, embedded in a dry spongy tissue, 7-8 mm long, about 5.5 mm broad, smooth, elliptic, thick, only slightly compressed; areole 6.5-7 mm long, 4 mm wide.

GERMPLASM: Both A. farnesiana and its var cavenia are extensively cultivated in and around Cannes, southern France, the center for production of the perfume; cavenia seems more resistant to drought and frost. Assigned to the South American Center of Diversity, cassie or cvs thereof is reported to exhibit tolerance to drought, high pH, heat,

low pH, salt, sand, slope, and savanna. (2n = 52, 104).

DISTRIBUTION: Probably native to tropical America, but widely naturalized and cultivated—Africa (Rhodesia, Mozambique) and Australia. Planted in coastal areas of Ghana and elsewhere in tropical Africa. Grown throughout India, often in gardens.

ECOLOGY: Thrives in dry localities and on loamy or sandy soils; may serve as a sand binder. Grows on loose, sandy soil of river beds, on pure sand in plains of Punjab. Requires a dry tropical climate. Ranging from Warm Temperate Dry through Tropical Desert to Moist Forest Life Zones, cassie is reported to tolerate annual precipitation of 6.4–40.3 dm (mean of 20 cases = 14.0 dm), annual mean temperature of 14.7–27.8°C (mean of 20 cases = 24.1°C), and pH of 5.0–8.0 (mean of 15 cases = 6.8).

CULTIVATION: Propagated mainly from seed and cuttings. Seeds germinate readily and plants grow rapidly. Plants do not require much cultivation, watering, or care.

HARVESTING: Trees begin to flower from the third year, mainly November-March. Perfume is extracted from the flowers in form of concrete or pomade. Macerated flowers held several hours in melted purified natural fat, then repeatedly replaced by fresh flowers until the fat is saturated with perfume. Fat melted, strained, and cooled to yield pomade. Odor is that of violets but more intense. For absolute, pomade is mixed with alcohol (2-3 kg to ca. 4 liters) and held for 3-4 weeks at ca. -5°C. The alcohol is separated by distillation. The extract is an olive-green liquid with strong odor of cassie flowers.

YIELDS AND ECONOMICS: Mature trees yield up to 1 kg of flowers per season. Southern France (Cannes and Grasse) is main production center for cassie flower perfume. India and other Eastern countries produce much for local use.

BIOTIC FACTORS: Fungi reported on this plant include: Camptomeris albizziae, Clitocybe tabescens, Hypocrea borneensis, Lenzites palisoti, L. repanda, Phyllachora acaciae, Phymatotrichum

omnivorum, Polystictus flavus, Ravenelia austris, R. hieronymi, R. siliquae, R. spegazziniana, Schizophyllum commune, Systingophora hieronymi, Tryblidiella rufula, and Uromycladium notabile. It may also be parasitized by the flowering plants Dendrophthoe falcata and Santalum album.

CONTRIBUTORS: J. A. Duke, C. F. Reed, J. K. P. Weder.

Acacia mearnsii de Wild.

FAMILY: Mimosaceae

COMMON NAMES: Black wattle, Acacia negra,

Acacia noir, Schwarze akazie,

Gomboom

SYNONYMS: Acacia mollissima auct., not Willd.

Acacia decurrens var mollis Lindl.

USES: Tree of economic importance in South and East Africa, Rhodesia, India, and Rio Grande do Sul area of South America and environs for fuel and for tanning of soft leather. Dried bark contains 30–54% tannin. Trees furnish fuel and lumber in some areas and add nitrogen and organic material to the soil. Bark is used for wood adhesives and flotation agents.

FOLK MEDICINE: Products are often used in folk medicine as styptics or astringents.

CHEMISTRY: Black wattle bark contains (-)-robinetinidol and (+)-catechin; the biflavonoids (-)-fisetinidol-(+)-catechin (2 diastereoisomers), (-)-robinetinidol-(+)-catechin and (-)-robinetinidol-(+)-gallocatechin; triflavonoids and condensed tannins. The heartwood is rich in (+)-leucofisetinidin (mollisacacidin) together with (-)-fisetinidol, (+)-fustin, butin, fisetin, butein, and biflavonoid condensates (tannins).

DESCRIPTION: Tree 6-20 m tall, 10-60 cm in diameter; crown conical or rounded; all parts except flowers usually pubescent or puberulous; stems without spines or prickles; leaves bipinnate, on petioles 1.5-2.5 cm long, with a gland above; rachis 4-12 cm long with numerous raised glands all along its upper side; pinnae in 8-30 pairs, pinnules in 16-70 pairs, linear-oblong, 1.5-4 mm long,

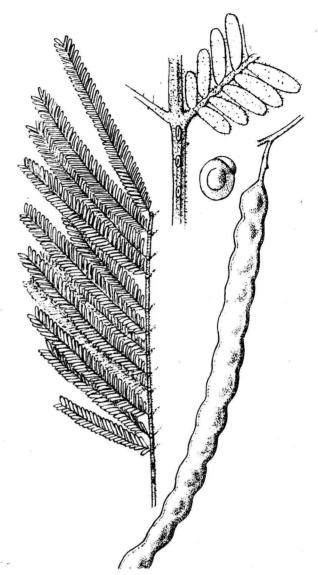


Figure 2. Acacia mearnsii de Wild.

0.5-0.75 mm wide; flowers in globose heads 5-8 mm in diameter, borne in panicles or racemes, on peduncles 2-6 mm long; pale yellow and fragrant; pods gray-puberulous, or sometimes glabrous, almost moniliform, dehiscing, usually 3-10 cm long, 0.5-0.8 cm wide, with 3-14 joints; seeds black, smooth, elliptic or compressed ovoid, 3-5 mm long, 2-3.5 mm wide; caruncle chaspicuous; areole 3.5 mm long, 2 mm wide. Seeds 66,000-110,000/kg.

GERMPLASM: Can be erossed with Acacia decurrens; hybrids show more sterility than parents.

Meiosis is regular, with no gross cytological abnormalities, and sterility may be due to gene differentiation between species. There is little geographic overlap in the native Australian ranges of the species, and there are differences in phenology (flowering; seedset). Most of the characters that vary among species are quantitative. The development of black wattle strains or of hybrids with enhanced vigor, better quality bark, outstanding stem form, or resistance to insect pests and disease would benefit the wattle industry. Assigned to the Australian Center of Diversity, black wattle or cvs thereof is reported to exhibit tolerance to drought, laterite, and poor soil. (2n = 26.1)

DISTRIBUTION: Native to Southeast Australia (Victoria to New South Wales and southern Queensland) and Tasmania. Introduced and cultivated widely for afforestations. See Sherry (1971) for details.

ECOLOGY: In Kenya grows on or near Equator at altitudes of 2,000-2,800 m; is well adjusted to the climate of East Africa. Grows well at 30°S Lat. in South America on rolling terrain at altitudes of 50-70 m. Thrives on poor, dry soils but favors deep, moist, fertile soils. In Australia, black wattle may occur on soils derived from shales, mudstones, sandstones, conglomerates, and alluvial deposits; in Kenya on podsols, krasnozems, sandy hills, lava flows or on mixtures of lava and contemporaneous volcanic tuffs and breccias. In South America. grows on red clay or sandy soils that have suffered from severe erosion and soil depletion (ferruginous clay loams with little or no free silica). In East Africa grows where annual rainfall is 1,041–1,321 mm, (about 75% between April and September). On the equator where black wattle is grown in South America, the rain pattern is nearly opposite. mean annual temperature range is 17-23°C; there is little seasonal variation, but considerable diurnal variation. At higher altitudes in South America, frost is a risk and heavy snows may break tree limbs. Tannin content varies inversely with precipitation. Ranging from Warm Temperate Dry through Tropical Thorn to Tropical Moist Forest Life Zones, black wattle is reported to tolerate annual precipitation of 6.6-22.8 dm (mean of 6 cases = 12.6), annual mean temperature of $14.7-27.8^{\circ}$ C (mean of 6 cases = 22.6° C), and pH of 5.0-7.2 (mean of 5 cases = 6.5).

CULTIVATION: Propagation by seed is easy. Seeds retain their viability for several years. For germination seeds are covered with boiling water and allowed to stand until cool. This cracks the hard outer coat and facilitates germination. Seeds may be broadcast or sown in rows on any barren site. Usually they are sown about 5 cm apart in seedbeds, and are transplanted after 3-6 months. In South America, fields are usually plowed and harrowed in April or May. Seedlings are set out May-November, but usually in winter, June-August, after a rain. Plants are spaced 2 m each way, at rate of 2,500/ha. Propagation by cuttings is almost impossible without mist. Air layering is more promising. Two types of farmers grow acacia: the tanner or business person plants 200 ha or so entirely to black wattle, usually one section at a time so that seeds can be planted and harvested within the same year and continue year after year: the farmer plants half or less of the land to black wattle and the rest to crops such as corn, beans, manioc, sugarcane, other vegetables, or pasture. The farmer plants 2-6 ha of acacia each year and thus evenly distributes work and production. Oxen may be useful for plowing, but most work is manual, using only plows and hoes. Intercrops may be grown the first year during which trees grow about 4-5 m in height, and about 2.5 cm in diameter.

HARVESTING: Trees provide bark 5-10 years after seeding (ave. 7). Bark is stripped from lower part of tree, then tree is felled, the remaining bark removed, and tree and bark are cut into 1-m lengths. Thoroughly dried bark is arranged in bales of 75–80 kg when ready for transportation. Tanning power improves by 10-15% in bark carefully stored for a season. Percent tannin does not differ between barks harvested in dry and wet seasons. However, the amount of bark on trees may be less on poor than on rich soils. Tannin runs about 25-35% of dried bark, on either poor or rich soil. Acacia bark may be sold as baled bark or bark powder. Dried bark may go first to commercial bark processors where it is ground or shredded in a hammermill, then sold in 40-kg sacks. Bark powder is sold in 60kg sacks. Liquid extract is sold in 300-kg wooden barrels. In Rio Grande do Sul an estimated 5.000 MT of liquid extract is produced annually.

YIELDS AND ECONOMICS: Like some mangrove species, black wattle in pure stand produces more