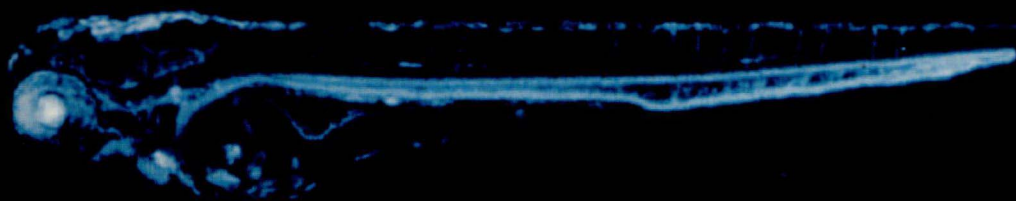


# Optical Nanoscopy and Novel Microscopy Techniques

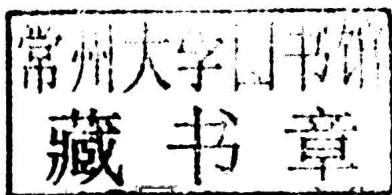


Edited by Peng Xi



CRC Press  
Taylor & Francis Group

# Optical Nanoscopy and Novel Microscopy Techniques



Edited by Feng Xi



**CRC Press**

Taylor & Francis Group  
Boca Raton London New York

---

CRC Press is an imprint of the  
Taylor & Francis Group, an **Informa** business

CRC Press  
Taylor & Francis Group  
6000 Broken Sound Parkway NW, Suite 300  
Boca Raton, FL 33487-2742

© 2015 by Taylor & Francis Group, LLC  
CRC Press is an imprint of Taylor & Francis Group, an Informa business

No claim to original U.S. Government works

Printed on acid-free paper  
Version Date: 20140717

International Standard Book Number-13: 978-1-4665-8629-1 (Hardback)

This book contains information obtained from authentic and highly regarded sources. Reasonable efforts have been made to publish reliable data and information, but the author and publisher cannot assume responsibility for the validity of all materials or the consequences of their use. The authors and publishers have attempted to trace the copyright holders of all material reproduced in this publication and apologize to copyright holders if permission to publish in this form has not been obtained. If any copyright material has not been acknowledged please write and let us know so we may rectify in any future reprint.

Except as permitted under U.S. Copyright Law, no part of this book may be reprinted, reproduced, transmitted, or utilized in any form by any electronic, mechanical, or other means, now known or hereafter invented, including photocopying, microfilming, and recording, or in any information storage or retrieval system, without written permission from the publishers.

For permission to photocopy or use material electronically from this work, please access [www.copyright.com](http://www.copyright.com) (<http://www.copyright.com/>) or contact the Copyright Clearance Center, Inc. (CCC), 222 Rosewood Drive, Danvers, MA 01923, 978-750-8400. CCC is a not-for-profit organization that provides licenses and registration for a variety of users. For organizations that have been granted a photocopy license by the CCC, a separate system of payment has been arranged.

**Trademark Notice:** Product or corporate names may be trademarks or registered trademarks, and are used only for identification and explanation without intent to infringe.

**Visit the Taylor & Francis Web site at**  
**<http://www.taylorandfrancis.com>**

**and the CRC Press Web site at**  
**<http://www.crcpress.com>**

# Optical Nanoscopy and Novel Microscopy Techniques



---

# Preface

Microscopy is at the forefront of multidisciplinary research. It was developed by physicists, made specific by chemists, and applied by biologists and doctors to better understand the function of the subunits of the human body. For this very reason, the field has been revolutionized constantly in past decades.

The objective of this book is to choose some of those revolutionary ideas and help a general audience from broad disciplines to achieve a fundamental understanding of these technologies and to better apply them in their daily research. It is organized as follows: Chapters 1–3 are on super-resolution optical microscopy, in which the targeted modulation, such as STED and SIM, or the localization methods, such as PALM, are discussed; Chapters 4–7 are on the novel development of fluorescent probes, such as organic small-molecule probes, fluorescent proteins, and inorganic labels such as quantum dots; Chapters 8–10 are about advanced optical microscopy, such as fluorescence lifetime imaging, fiber optic microscopy, scanning ion conductance microscopy, and the joining of optics and acoustics—photo-acoustic microscopy. Following each chapter is a detailed list of references so that the readers can use this book as a review of the relevant fields. In addition, to confirm understanding of the materials presented, we provide several problems after each chapter.

I express my deepest appreciation to the contributors of each chapter. Usually readers only remember the name of the editor, but the contributors are the real teachers and where the credit is due. I thank Li-Ming Leoung for her invitation to edit the book and for continuous support during the manuscript's preparation. I thank Amit Lal for proofreading the manuscript. I would like to thank all the CRC editorial team for their professional service, which is essential for the book to reach readers. Also, my sincere gratitude to my wife, Guanqin Chang, and our daughter, Yueming Xi, for their understanding and for sacrificing their quality time with me.

And thank you, my dear reader, for choosing this book. I guarantee that it will be a delightful and resourceful journey.

**Peng Xi**

*Zhong Guan Yuan, Peking University*



---

# Contributors

**Hao Chang**

Institute of Biophysics  
Chinese Academy of Sciences  
Beijing, China

**Peng R. Chen**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Dan Dan**

Xi'an Institute of Optics and  
Precision Mechanics  
Chinese Academy of Sciences  
Xi'an, China

**Yichen Ding**

Peking University  
Peking, China

**Zhicong Fei**

IMM  
Peking University  
Peking, China

**Jianhong Ge**

State Key Laboratory of Modern  
Optical Instrumentation  
Zhejiang University  
Hangzhou, China

**Yuchun Gu**

IMM  
Peking University  
Peking, China

**Jin U. Kang**

Department of Electrical and  
Computer Engineering  
Johns Hopkins University  
Baltimore, Maryland

**Fu-Jen Kao**

Institute of Biophotonics  
National Yang-Ming University  
Taipei, Taiwan

**Cuifang Kuang**

State Key Laboratory of Modern  
Optical Instrumentation  
Zhejiang University  
Zhejiang, China

**Ming Lei**

Xi'an Institute of Optics and  
Precision Mechanics  
Chinese Academy of Sciences  
Xi'an, China

**Changhui Li**

Department of Biomedical  
Engineering  
Peking University  
Peking, China



**Jie Li**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Po-Yen Lin**

Institute of Physics  
Academia Sinica  
Taipei, Taiwan

**Xuan Liu**

Biomedical Engineering  
Michigan Technological University  
Houghton, Michigan

**Yujia Liu**

Faculty of Science  
Macquarie University  
Sydney, Australia

**Qiushi Ren**

Department of Biomedical  
Engineering  
Peking University  
Beijing, China

**Hui Shi**

IMM  
Peking University  
Peking, China

**Andrew M. Smith**

Department of Bioengineering  
University of Illinois at  
Urbana-Champaign  
Champaign, Illinois

**Yujie Sun**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Jie Wang**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Peng Xi**

Department of Biomedical  
Engineering  
Peking University  
Peking, China

**Hao Xie**

Department of Biomedical  
Engineering  
Peking University  
Peking, China

**Pingyong Xu**

Institute of Biophysics  
Chinese Academy of Sciences  
Beijing, China

**Maiyun Yang**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Yi Yang**

College of Chemistry and  
Molecular Engineering  
Peking University  
Peking, China

**Baoli Yao**

Xi'an Institute of Optics and  
Precision Mechanics  
Chinese Academy of Sciences  
Beijing, China

**Mohammad U. Zahid**

University of Illinois at  
Urbana-Champaign  
Champaign, Illinois

**Mingshu Zhang**

Institute of Biophysics  
Chinese Academy of Sciences  
Beijing, China

**Yanjun Zhang**

Tianjin Medical University  
Tianjin, China



---

# Contents

Preface.....	vii
Contributors.....	ix
<b>Chapter 1   Optical nanoscopy with stimulated emission depletion....</b>	<b>1</b>
<i>Peng Xi, Hao Xie, Yujia Liu, and Yichen Ding</i>	
<b>Chapter 2   Structured illumination microscopy .....</b>	<b>23</b>
<i>Dan Dan, Baoli Yao, and Ming Lei</i>	
<b>Chapter 3   Super-resolution imaging with stochastic optical reconstruction microscopy (STORM) and photoactivated localization microscopy (PALM) .....</b>	<b>61</b>
<i>Yujie Sun</i>	
<b>Chapter 4   Small-molecule labeling probes .....</b>	<b>85</b>
<i>Jie Wang, Jie Li, Yi Yang, Maiyun Yang, and Peng R. Chen</i>	
<b>Chapter 5   Fluorescent proteins for optical microscopy .....</b>	<b>111</b>
<i>Pingyong Xu, Mingshu Zhang, and Hao Chang</i>	
<b>Chapter 6   Single-molecule imaging with quantum dots.....</b>	<b>135</b>
<i>Mohammad U. Zahid and Andrew M. Smith</i>	
<b>Chapter 7   Fluorescence detection and lifetime imaging with stimulated emission.....</b>	<b>161</b>
<i>Po-Yen Lin, Jianhong Ge, Cuifang Kuang, and Fu-Jen Kao</i>	
<b>Chapter 8   Fiber optic microscopy .....</b>	<b>179</b>
<i>Jin U. Kang and Xuan Liu</i>	

**Chapter 9 Scanning ion conductance microscopy ..... 199**  
*Zhicong Fei, Hui Shi, Yanjun Zhang, and Yuchun Gu*

**Chapter 10 Advanced photoacoustic microscopy..... 215**  
*Yichen Ding, Qiushi Ren, and Changhui Li*

**Index ..... 241**

# Optical nanoscopy with stimulated emission depletion

Peng Xi, Hao Xie, Yujia Liu, and Yichen Ding

Contents

- 1.1 Introduction..... 1
- 1.2 Point spread function and diffraction limit..... 2
  - 1.2.1 Diffraction limit criteria..... 2
  - 1.2.2 PSF Measurement ..... 4
- 1.3 Stimulated Emission Depletion (STED)..... 5
  - 1.3.1 The STED Process ..... 6
  - 1.3.2 Resolution Scaling with STED ..... 8
  - 1.3.3 STED and Other Imaging Modalities ..... 12
    - 1.3.3.1 Time-gated STED ..... 12
    - 1.3.3.2 STED + 4Pi ..... 12
    - 1.3.3.3 ISO-STED ..... 12
    - 1.3.3.4 Multiphoton-STED..... 13
    - 1.3.3.5 Automatic Spatially Aligned STED ..... 13
- 1.4 Technical Notes on STED Optical Nanoscopy ..... 14
  - 1.4.1 Generation and Measurement of the PSFs..... 14
  - 1.4.2 Measurement of the Saturation Intensity ..... 16
  - 1.4.3 Sample Preparation ..... 16
  - 1.4.4 Resolution Measurement ..... 17
- 1.5 Conclusion ..... 17
- References..... 19

## 1.1 Introduction

Since its invention in the seventeenth century, optical microscopy has been applied to biological study. For the past 400 years it has been at the forefront of biological study, from the macroscopic level to the microscopic, cellular level. For “seeing is believing,” optical microscopy has been widely extended to almost all disciplines of scientific research. Yet, due to the fact that optics has a wave nature, the diffraction limit becomes the resolution barrier. In the past decade, numerous super-resolution

techniques have been created. Stimulated emission depletion (STED) is one of the techniques that employs the feature of fluorescence to generate a smaller effective point spread function (PSF), or PSF engineering.

In this chapter, we first discuss the diffraction limit and its appearance as a PSF; then, the STED process in a single molecule energy level, as well as its behavior in a fluorescent medium. The combination of other techniques with STED, such as time-gating, iso-STED, STED and 4Pi, two-photon excitation, and self-aligned STED techniques are discussed. Next, the technical notes on the experimental instrumentation of STED are presented. Finally, the applications of STED to a variety of biological specimens are listed.

## 1.2 Point spread function and diffraction limit

### 1.2.1 Diffraction limit criteria

Before breaking it, we need to investigate how the barrier of optical diffraction limit is formed. We need to take a history tour, to a beautiful small town in Germany called Jena. We understand that the development of optics requires three key elements: theory, engineer, and material. The time was back in the 1870s, when three giants met magically here. Ernst Karl Abbe, an optical theory master; Carl Zeiss, a great founder of optical engineering; and Otto Schott, a pioneer in optical materials. They have made concerted efforts to lead optics to a higher level for the whole world. Nowadays, the enterprises founded by Schott and Zeiss, respectively, are still renowned around the world.

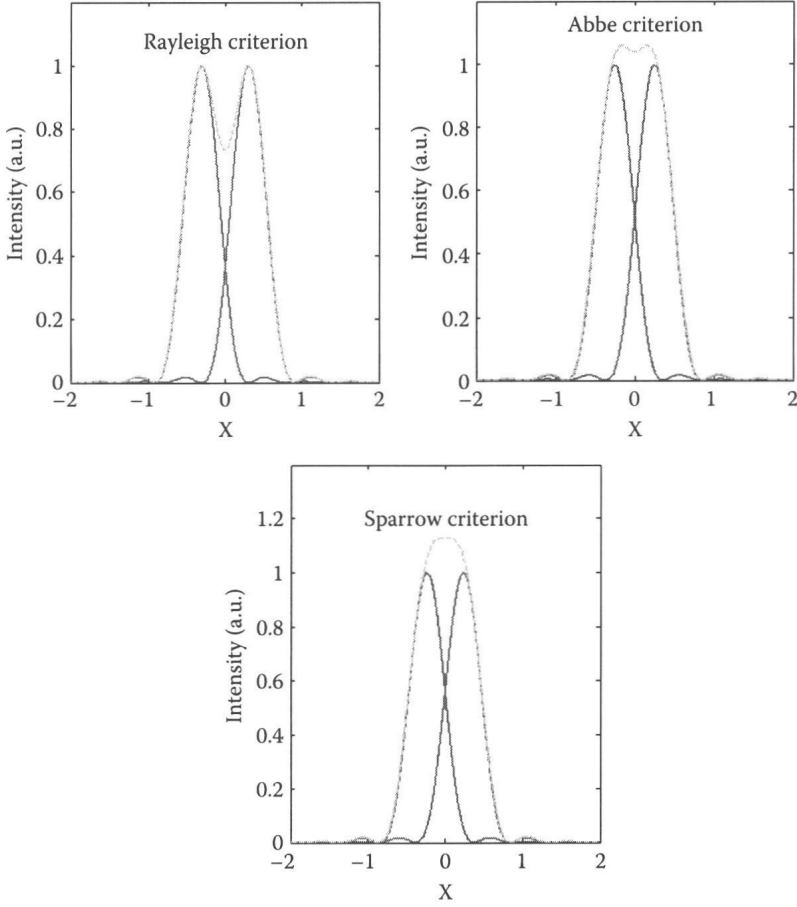
When light propagates through a barrier such as the aperture of objective, diffraction happens. With wave optics, one can describe the optical distribution of the focal plane as

$$I(0, v) = \left[ \frac{2J_1(v)}{v} \right]^2 I_0, \quad (1.1)$$

where

$$v = \frac{2\pi}{\lambda} NA \cdot r,$$

$NA = n \sin \alpha$  is the numerical aperture of the objective, and  $r$  is the radius of the point of interest.  $I_0$  and  $\lambda$  are the intensity and wavelength of the incident beam, respectively. This gives us a Gaussian-like function as illustrated in Figure 1.1, called the point spread function (PSF), that is, the image (spread function) of a perfect point (or equivalently, a planar wave). It is known as Airy pattern, or Airy disk in the two-dimensional form.



**Figure 1.1** (See color insert.) Comparison of the various diffraction limit criteria.

As one can see immediately from the plot, the closer two PSFs are, the more difficult it is to separate them. Lord Rayleigh formulated the Rayleigh criterion (Lord Rayleigh 1896):

$$d_{\text{Rayleigh}} = 0.61 \frac{\lambda}{NA} \quad (1.2)$$

This criterion is measured from the maximum of the PSF to the first minimum. More technically, the full width at half maximum (FWHM) is often used, which can be expressed as:

$$d_{\text{FWHM}} = 0.51 \frac{\lambda}{NA} \quad (1.3)$$



By putting two PSFs at the distance of FWHM, one can notice that, there is a dip of  $\sim 3\%$ . A more elegant expression is given by Abbe (Abbe 1873):

$$d_{\text{Abbe}} = \frac{\lambda}{2NA} \quad (1.4)$$

Clearly, to distinguish the two points, they cannot get too close to each other. However, there is still ambiguity as to how well these combined PSFs can be resolved. Alternative evaluations are like Sparrow's resolution limit (Sparrow 1919), which states that the resolution limit is where the dip disappears (becomes flat, or the second-order derivative equals zero).

$$d_{\text{Sparrow}} = 0.47 \frac{\lambda}{NA} \quad (1.5)$$

In a confocal setup, a pinhole is placed before a point-detector (such as a photomultiplier tube, PMT, or an avalanche photodiode, APD). Under ideal conditions (the pinhole is infinitely small), the FWHM of such a system can be described as:

$$d_{\text{Confocal}} = \frac{0.51\lambda}{\sqrt{2}NA} \approx \frac{0.36\lambda}{NA} \quad (1.6)$$

Note that this indicates the detector cannot get any photon since its size equals to zero. Increasing the pinhole will get more signal, with a scarification of resolution.

### 1.2.2 PSF Measurement

The PSF of a confocal system is the excitation PSF modulated by the pinhole's modulation spread function (MSF):

$$\text{PSF}_{\text{Confocal}} = \text{PSF}_{\text{Excitation}} \cdot \text{MSF} \quad (1.7)$$

When the pinhole size is a point (delta function), the MSF equals the PSF at the fluorescent wavelength. Since practically the size of the confocal pinhole is not infinitely small, the MSF can be calculated with the two-dimensional convolution of the PSF, and the size of the pinhole is given by:

$$\text{MSF} = \text{PSF}_{\text{Detection}} \otimes \text{Pinhole} \quad (1.8)$$

where  $\otimes$  denotes the convolution process.