biotechnology of amino acid production

EDITED BY

ICHIRO CHIBATA

KIYOSHI NAKAYAMA

KOICHI TAKINAMI

and
HIDEAKI YAMADA

progress in industrial microbiology



biotechnology of amino acid production

EDITED BY

KŎ AIDA

Ochanomizu University, Tokyo, Japan

ICHIRO CHIBATA

Tanabe Seiyaku Co., Ltd. Osaka, Japan

KIYOSHI NAKAYAMA

Zama Research Laboratories, Bior Inc., Zama, Japan

KOICHI TAKINAMI

Ajinomoto Co., Inc., Kawasaki, Japan

and

HIDEAKI YAMADA

Kyoto University, Kyoto Japan

progress in industrial microbiology

1986





KODANSHA LTD. Tokyo



ELSEVIER Amsterda Copublished by KODANSHA LTD., Tokyo

and
ELSEVIER SCIENCE PUBLISHERS B. V., Amsterdam

exclusive sales rights in Japan KODANSHA LTD. 12-21, Otowa 2-chome, Bunkyo-ku, Tokyo 112, Japan

for the U.S.A. and Canada ELSEVIER SCIENCE PUBLISHING COMPANY, INC. 52 Vanderbilt Avenue, New York, NY 10017

for the rest of the world ELSEVIER SCIENCE PUBLISHERS B. V. 25 Sara Burgerbartstraat, P.O. Box 211, 1000 AE Amsterdam, The Netherlands

ISBN 0-444-99502-1 (Vol. 24) ISBN 0-444-41666-8 (Series)

ISBN 4-08-201742-3 (Japan)

Copyright @ 1986 by Kodansha Ltd.

All rights reserved.

No part of this book may be reproduced in any form, by photostat, microfilm, retrieval system, or any other means, without the written permission of Kodansha Ltd. (except in the case of brief quotation for criticism or review).

PRINTED IN JAPAN

List of Contributors

Numbers in parentheses indicate the Chapters to which the authors contributed. Editors are indicated by an asterisk.

- *Kō Aida (overview), Department of Nutrition and Food Science, Ochanomizu University, Eunkyo-ku, Tokyo 112, Japan
- Kázumi Araki (22), Tokyo Research Laboratory, Kyowa Hakko Kogyo Co., Ltd., Machida-shi, Tokyo 194, Japan
- Teruhiko Beppu (3), Department of Agricultural Chemistry, Faculty of Agriculture, University of Tokyo, Bunkyo-ku, Tokyo 113, Japan
- *Ichiro Chibata (13, 20), Research and Development Headquaters, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Osaka 532 Japan
 - Hitoshi Enei (25), Applied Research Department, Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki-shi 210, Japan
 - Yuji Goto (8), Process and Engineering Development Laboratories, Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki-shi 210, Japan
 - Yoshio Hirose (6), International Research and Development, Ajinomoto Co., Inc., Chuo-ku, Tokyo 104, Japan
 - Shigeaki Ichikawa (27), Pharmaceutical R & D Laboratory, Asahi Chemical Industry Co., Ltd., Asahi-cho, Nobeoka-shi 882, Japan
 - Toshio Kakimoto (20), Analytical Chemistry Research Laboratory, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Osaka 532, Japan
 - Isao Karube (7), Research Laboratory of Resource Utilization, Tokyo Institute of Technology, Midori-ku, Yokohama 227, Japan
 - Masakazu Kikuchi (9), Biotechnology Laboratories, Central Research Division, Takeda Chemical Industries, Ltd., Yodogawa-ku, Osaka 532, Japan
 - Akira Kimura (28), Applied Microbiology Research Institute for Food Science, Kyoto University, Uji-shi, Kyoto 611, Japan
 - Masahiko Kisumi (2, 21), Research Laboratory of Applied Biochemistry, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Osaka 532, Japan

- Saburo Komatsubara (21), Research Laboratory of Applied Biochemistry, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Osaka 532, Japan
- Hidehiko Kumagai (23), Department of Food Science and Technology, Faculty of Agriculture, Kyoto University, Sakyo-ku, Kyoto 606, Japan Yasuji Minoda (5), Emeritus professor, University of Tokyo, Bunkyo-ku, Tokyo 113, Japan
- Yasushi Morinaga (18), Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki-shi 210, Japan
- Toru Nagasawa (19), Department of Agricultural Chemistry, Faculty of Agriculture, Koyto University, Sakyo-ku, Kyoto 606, Japan
- Shigeru Nakamori (15), Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki-shi 210, Japan
- Yoshio Nakao (9), Applied Microbiology Laboratories, Central Research Division, Takeda Chemical Industries, Ltd., Yodogawa-ku, Osaka 532, Japan
- *Kiyoshi Nakayama (1), Zama Research Laboratories, Bior Inc., Zamashi, Kanagawa-ken 228, Japan
 - Heiichi Sakai (26), Faculty of Agriculture, Tamagawa University, Machida-shi, Tokyo 194, Japan
 - Tadashi Sato (13), Research Laboratory of Applied Biochemistry, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Oaska 532, Japan
 - Mitsuru Shibukawa (27), Pharmaceutical R & D Laboratory, Asahi Chemical Industry Co., Ltd., Chiyoda-ku, Tokyo 100, Japan
 - Isamu Shiio (17), Central Research Laboratories, Ajinomoto Co., Inc., Totsuka-ku, Yokohama 244, Japan
 - Kenji Soda (4, 16), Institute of Chemical Research, Kyoto University, Uji-shi, Kyoto 611, Japan
 - Shuichi Suzuki (7), Saitama Institute of Technology, Okabe, Oosato, Saitama 369-02, Japan
 - Takashi Tachiki (11), Department of Food Science and Technology, Faculty of Agriculture, Kyoto University, Uji-shi, Kyoto 611, Japan
 - Satomi Takahashi (24), Biochemical Research Laboratories, Kanegafuchi Chemical Industry Co., Ltd., Takasago-shi, Hyogo-ken 676, Japan
- *Koichi Takinami (14), Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki-shi 210, Japan
 - Hidehiko Tanaka (4), Institute for Chemical Research, Kyoto University, Uji-shi, Kyoto 611, Japan
 - Tatsurokuro Tochikura, Department of Food Science and Technology, Faculty of Agriculture, Kyoto University, Sakyo-ku, Kyoto 606, Japan
 - Tetsuya Tosa (13, 20), Research Laboratory of Applied Biochemistry, Tanabe Seiyaku Co., Ltd., Yodogawa-ku, Oaska 532, Japan

- Osamu Tosaka (14), Kyushu Factory, Ajinomoto Co., Inc., Saga-gun, Saga-ken 840-21, Japan
- *Hideaki Yamada (18, 19, 25), Department of Agricultural Chemistry, Faculty of Agriculture, Kyoto University, Sakyo-ku, Kyoto 606, Japan Hajime Yoshida (12), Tokyo Research Laboratory, Kyowa Hakko Kogyo Co., Ltd., Machida-shi, Tokyo 194, Japan
 - Tohru Yoshimura (4), Department of Applied Biology, Faculty of Textile Science, Kyoto Institute of Technology, Sakyo-ku, Kyoto 606, Japan Fumiliro Yoshinaga (10), Planning and Development Department, Aiinomoto Co., Inc., Chuo-ku, Tokyo 104, Japan

In 1957 several novel bacteria with high productivity of L-glutamic acid were found in various sources, and a new fermentation method called L-glutamic acid fermentation commenced in the field of applied microbiology. Since this discovery many investigators have undertaken studies on the microbial production of other amino acids, including L-lysine, L-isoleucine, L-leucine, L-valine, L-threonine, L-aspartic acid L-alanine, L-serine, L-arginine, L-tryptophan, L-phenylalanine, L-proline and L-histidine. Over the past quarter of a century, great advances have been made in breeding microorganisms capable of hyperproduction of these amino acids.

In 1972 the first monograph on amino acid fermentation, The Microbial Production of Amino Acids edited by Yamada et al. (Kodansha), was published. This was a significant publication in the field of industrial microbiology which helped to establish amino acid fermentation subsequent to antibiotics fermentation.

Over the past few decades, microbiologists and biochemists have introduced molecular biology and genetic engineering to the field of applied and industrial microbiology. Amino acid fermentation is the practical result of biotechnology which integrates microbiology, biochemistry and chemical engineering.

This volume discusses the general aspects of amino acid fermentation fundamental to industrial use. The microorganisms and their biosynthetic pathways and related enzymes are described in detail in order to clarify the regulatory mechanism of the microbial metabolism. In particular, the various amino acids are treated separately in individual chapters, including descriptions of the development of bio-

x PREFACE

chemical processes. The book covers general and specific aspects of industrial microbiology research, past, present and future.

Kō Aida Ichiro Chibata Kiyoshi Nakayama Koichi Takinami Hideaki Yamada

An Overview of the Microbial Production of Amino Acids

Kō AIDA
Ochanomizu University, Tokyo

Since the discovery of monosodium glutamate (MGS) as a seasoning by Kikunae Ikeda in 1908, demand in the food industry has increased steadily throughout the world. Formerly, it was industrially obtained from wheat, soybean and other plant proteins or from Steffen's waste. However, the fact that appreciable quantities of L-glutamic acid may be synthesized and accumulated in bacterial culture broth was first reported in Japan in 1957, and one company soon began its commercial production by a fermentative process, to be followed later by various manufacturers throughout the world.

Stimulated by this success, systematic research on amino acid biosynthesis by microorganisms quickly developed. In 1959, the Association of Amino Acid Fermentation was established by the researchers in this field, and it held annual meetings and issued a biannual bulletin in Japanese. In 1963, the name of the Association was changed to the "Association of Amino Acid and Nucleic Acid" to accommodate the new results on the fermentative production of nucleic acid-related substances such as 5'-inocinic acid and 5'-guanvlic acid which were reported in 1959 and produced commercially since 1961. As commemoration publications, The Microbial Preduction of Amino Acids and Microbial Production of Nucleic Acid-Related Substances were published in 1972 and 1976, both by Kodansha, Tokyo. The Association merged with the Japanese Association of Industrial Fermentation in 1979. The April and October issues of its bulletin, "Fermentation and Industry" were published as special issues under the former title Amino Acid and Nucleic Acid. At the time of the merger, Bibliography of Amino Acid and Nucleic Acid Fermentation was published in 1980 covering original reports and reviews issued from 1958 to 1979.

The trends in research concerning amino acid fermentation have been summarized by the numbers of papers published as shown in Table I. Predominant interest focused on glutamic acid fermentation. Lysine and tryptophan were also subjects of keen interest. At present, most of the usual amino acids can be produced by microbial or enzymatic methods, although these methods compete economically with

TABLE I. Recent Trend of Research Concering Biotechnology of Amino Acid Production

	^	-1 9 79†	'80	'81	'82	'83	'84	'85
. Origin	al Report							
1. Ami	no acid fermentation	49	3	2	4	5	4	1
A	lanine	18	2	5	3	0	2	
A	rginine	46	4	3	4	4	5	
A	spartic acid	0	0	0	0	.1	0	1
A	sparagine	9	1	3	0	0	1	- 1
C	itrulline	11	i	3	4	4	3	
C	ystine, Cystein	5 76	7	9	19	8	4	
G	lutamic acid	21	4	3	22	4	3	
G	lutamine	0	0	1	1	1	0	
G	lycine	21	0	0	1	0	1	
H	istidine	0	0	0	0	0	1	
H	iomoserine	35	1	0	0	0	0	
Is	oleucine	84	1	4	0	12	2	
L	eucine	24	1	4	. 4	2	0	
L	ysine	234	14	3	4	8	6	1
N	1ethionine	25	2	2	6	5	3	1
C)rnithine	13	i	0	1	1	0	
P	henylalanine	29	0	1	3	5	5	
P	roline	28	5	3	3	1	4	
S	erine	17	2	2	8	3	2	
T	'hreonine	56	1	4	1	I	1	
Т	`ryptophan	133	· 4	2	9	13	8	1
Т	yrosine	34	4	2	1	0	1	
V	Valine	70	0	2	0	1	0	
ind	neral amino acid, ustrial technology, material	95	16	8	15	8	2	
am	zyme related to ino acid biosynthesis I metabolism	178	57	161	23	36	107	1:
4. Rec	combinant DNA		7	13	12	18	62	I
5. An	alytical method	34	9	36	12	7	5	
B. Revie	:w							
I. Pro	oduction	112	5	19	11	50	24	
	synthesis and tabolic control	62	6	11	5	8	1	
3. Ap	plication	81	2	8	8	5	2	

[†] Described in Bibliography of Amino Acid and Nucleic Acid Fermentation, Association of Amino Acid and Nucleic Acid, Japan (1980).

other methods such as hydrolysis of natural proteins and chemical synthesis.

Microbial Production of Amino Acids

In 1957, two research groups, Kinoshita et al. and Asai et al., reported that L-glutamic acid can be produced in culture broth in appreciable quantities. Thereafter, the ability to produce glutamic acid has been found in many microorganisms, including Corynebacterium glutamicum. Brevibacterium flavum, B. lactofermentum, B. thiogenitalis and Microbacterium ammoniaphilum. The general characteristics of strains giving yields greater than 30% of glutamic acid from sugars were as follows: gram positive, non-sporulating, non-motile, coccal or rod-like; all required biotin for growth. Biotin was found to play a critical role in glutamic acid fermentation. Therefore, the function of biotin in glutamic acid fermentation has been studied extensively from both academic and industrial points of view. To utilize raw materials such as molasses which contain large amount of biotin, the addition of penicillin during fermentation was found to be effective. Several detergents were also found to be useful for the same purpose. The role of biotin was found to regulate the cell membrane permeability of glutamic acid through changes in cell membrane phospholipids. Based on these studies, a new type of glutamic acid-producing mutant was induced. It is a glycerol requiring mutant of Corynebacterium alkanolyticum, and the cell membrane permeability of glutamic acid can be regulated by the concentration of glycerol added to the medium. Glutamic acid fermentation is very interesting as a typical example whose most important characteristic is the regulation of cell membrane permeability of the fermentation product. Studies on the regulatory mechanism of glutamate biosynthesis have also been conducted in detail.

Following the success of glutamic acid fermentation, various methods have been developed for microbial production of amino acids. These may be classified as follows:

- a) Methods employing wild strains (yielding glutamic acid, alanine, valine)
- b) Methods employing mutants (yielding lysine, threonine, arginine, citrulline, ornithine, homoserine, tryptophan, phenylalanine, tyrosine, histidine, etc.)
- c) Precursor addition method (yielding threonine, isoleucine, tryptophan, etc.)
- d) Enzymatic method (yielding lysine, aspartic acid, alanine, D-p-hydroxyphenyl-glycine, etc.)

At first, wild strains from various environments were screened. but microorganisms which can produce amino acids in quantities sufficient to warrant commercial exploitation were extremely limited. Glutamic acid was the exception. Since amino acids are essential components of microbial cells and their biosynthesis is teleologically regulated at an optimum level, they are normally synthesized only in limited quantities. Terminal or near-terminal amino acids are under strict metabolic control, and special means must be devised to overcome such regulation in order to produce large quantities. The most successful technique so far developed involves the induction of auxotrophic mutants. analogue-resistant mutants, or combinations of these mutations. dustrial production of L-lysine was firstly accomplished by use of homoserine/threonine double auxotrophs of Corvnebacterium glutamicum. The induction of S-(2-aminoethyl)-L-cysteine (AEC)-resistant mutant of Brevibacterium flavum as a L-lysine-producing strain greatly stimulated the use of analogue resistant mutants thereafter.

The precursor addition method was successfully applied to L-threonine and L-isoleucine. For the former, the added substance was p-threonine in the case of Serratia marescens. For L-isoleucine, α-aminobutyric acid was added in the case of Bacillus subtilis etc. The enzymatic method involves microbial transformations and microbial enzymes. L-Aspartic acid can be produced almost quantitatively from fumaric acid and ammonia using aspartase of E. celi. Enzymatic production of L-lysine has been successful. pr-a-Amino-e-caprolactum is converted almost completely to L-lysine using L-a-amino-e-caprolactum hydrolase and α-amino-e-caprolactum racemase of Cryptococcus laurentii and Achromobacter obas, respectively, at the same time in the same vessel, DL-5-(phydroxyphenyl) hydantoin can be transformed to D-5-(p-hydroxyphenyl) hydantoin by asymmetric hydrolysis with hydantoinase of Pseudomonas putida. Under a slightly alkaline condition, residual L-form is transformed to p-form by spontaneous racemization. As a result, all pl-5-(p-hydroxyphenyl) hydantoin can be quantitatively converted to N-carbamoyl-pp-hydroxyphenyl-glycine, which is further converted to D-p-hydroxyphenylglycine by chemical decarboxylation with HNO₈ and HCl. This process has been industrialized and the product widely used as a raw material of semisynthetic antibiotics.

The present volume surveys all essential aspects of the biotechnology of amino acid production. Chapter I covers microbial and biochemical fundamentals, such as breeding of amino acid-overproducing mutants, intracellular genetic recombination, recombinant DNA method and related enzymes. Chapter 2 deals with biochemical engineering and fermentation technology, such as raw material, oxygen supply, automatic

analysis and computer control. The production of individual amino acids is described in detail in Chapter 3. Chapter 4 is devoted to the production of amino acid-related substances. So far as is known, no comparable treatise has yet been compiled in English.

Future Prospects

Any consideration of the future prospects of mass industrial production of amino acids by microbial or enzymatic processes cannot ignore the rival methods of chemical synthesis and protein hydrolysis. For example, DL-methionine, which is effectively used to strengthen animal fodder in large quantities, is produced by chemical synthesis. However, biotechnological methods have the imoprtant advantage of directly yielding the optically active L-form of amino acids.

Both amino acid fermentation and nucleic acid fermentation have been investigated mostly in Japan, and they are representative in the fermentation industry. Such unique techniques as the utilization of various kinds of mutants, increased permeability of cell membrane, artificial regulation of microbial metabolism, and the combination of enzymatic reaction and chemical synthesis have been developed. Over the past decade, significant progress has been made in the utilization of immobilized enzymes and cells for the microbial production of amino acids. Half-life of aspartase activity of immobilized E. coli cells increased to 680 days in a continuous enzymatic reaction system. Breeding of amino acid-overproducing strains has reached a state of very high development. Application of intracellular genetic recombination to strain improvement has been successful in several cases and recombinant DNA method and cell fusion method have been effectively applied to strain improvement in amino acid fermentation. It is reported that in the USSR L-threonine has been industrially produced by E. coli whose productivity of L-threonine was improved by recombinant DNA technique in 1979. The establishment of host-vector systems in genera Brevibacterium and Corynebacterium will further open new prospects in this field.

A principal factor which will control the future development of the amino acid fermentation industry is the exploitation of new uses for the amino acids produced. Efforts in this direction will be necessary in conjunction with basic research in the biotechnology of amino acid production.

Contents

	Prefa	ace ix
		Overview of the Microbial Production of Amino Acids xxi AIDA)
	PAR	T I MICROBIAL AND BIOCHEMICAL FUNDAMENTALS
l		ding of Amino Acid-overproducing Mutants oshi Nakayama)
	1.1	Efficient Permeation of Substrate Material into Microbia
	1.2	Cells 3 Efficient Procedures for the Metabolic Reaction Concerned 5
	1.3	Blocking Side Reaction 7
	1.4	9
	1.5	Efficient Leakage of Metabolite into Cell Environment 10
	1.6	Overproduction in Wild-type Strains 10
	1.7	Conclusion 12
		References 13
2	Stra	olication of Intracellular Genetic Recombination to in Construction
	(Ma	asahiko Kisumi)
	2.1	Advantages of Using Transductional Method 15
	2.2	Construction of Amino Acid-producing Strains 16
		2.2.1 Histidine-producing Strains 16
		2.2.2 Arginine-producing Strains 17
		2.2.3 Threonine- and Isoleucine-producing Strains 20
		2.2.4 Strains Producing Other Amino Acids 21
	2.3	
		References 23

List of Contributers

3	Bree	ding of Amino Acid-producing Strains 24
	(Ter	ruhiko Beppu)
	3.1	Recombinant DNA Technology 25 3.1.1 Trytophan 25 3.1.2 Threonine 26
		3.1.3 Lysine 28 3.1.4 Proline 28 3.1.5 Alanine 29 3.1.6 Glutathione 29
	3.2	
		 3.2.1 Host-vector Systems of Corynebacterium glutamicum 30 3.2.2 Host-vector Systems of Brevibacterium 32 3.2.3 Breeding of Coryneform Bacteria by Protoplast Fusion 33
	3.3	
	3.4	
4		ynthetic and Metabolic Enzymes of Amino Acids mji Soda and Tohru Yoshimura)
	4.1	Pyridoxal 5'-Phosphate-dependent Enzymes 36 4.1.1 Enzymes Catalyzing C _a -H Bond Cleavage 37
	4.2	NAD(P)+ Enzymes 42
	4.3	Flavin Enzymes 43
	4.4	Vitamin B ₁₂ Enzymes 44
	4.5	Hydrolytic Enzymes and Others 45 References 47

PART II BIOCHEMICAL ENGINEERING AND FERMENTATION TECHNOLOGY

5 Raw Materials for Amino Acid Fermentation (Yasuji MINODA)

5.1	Raw Materials Currently Used for Amino Acid Fermentation in Japan 52		
	5.1.1 Sugars 53		
	5.1.2 Starches 56		
5. 2	Raw Materials for Future Use in Amino Acid Fermentation		
	in Japan 57		
	5.2.1 Sugars 57		
	5.2.2 Starches 59		
	5.2.3 Cellulose 61		
	5.2.4 Ethanol 62		
5.3			
	5.3.1 Methanol 63		
	5.3.2 Carbon Dioxide 65		
	References 66		
Carl	hemical Effects of Oxygen Supply and bon Dioxide Removal 67 shio History)		
6.1	Determination of Oxygen and Carbon Dioxide 67		
6.2			
٠.٠	6.2.1 Effects of High Oxygen Tension on Cell Growth and		
	Amino Acid Formation 70		
	6.2.2 Relationship Between Amino Acid Productivity and		
	Degree of Satisfaction of Cell Oxygen Demand		
	(r_{ab}/KrM) 71		
	6.2.3 Effect of Carbon Dioxide Tension on Amino Acid		
	Fermentation 76		
6.3			
	References 79		
	omatic Analysis of Amino Acids Using Biosensors 81 o Karube and Shuichi Suzuki)		
7.1	Enzyme Sensor for Glutamic Acid 81		
	Microbial Sensor for Glutamic Acid 82		
7.3			
	References 89		
Computer Control of Fermentation Processes 90			
(Yuji Goro)			
8.1	Computers and Sensors 90		
	Process Control 92		

8.3 Advanced Control

References

	PAI	RT III PRODUCTION OF INDIVIDUAL AMINO ACIDS
9	-	amic Acid 101 sakazu Кікисні and Yoshio Nакао)
	9.1	Discovery of Glutamic Acid-producing Microorganisms and Their Features 102
	9.2	Microbial Production of Glutamic Acid 103 9.2.1 Production of Glutamic Acid from Sugar 103 9.2.2 Production of Glutamic Acid from Ethanol and Other Carbon Sources 104
		9.2.3 Production of Glutamic Acid from Acetic Acid 104 9.2.4 Production of Glutamic Acid from Benzoic Acid and Methanol 105
	9.3	9.2.5 Production of Glutamic Acid from Hydrocarbons 106 Regulation of Glutamic Acid Fermentation 107 9.3.1 Regulation of Membrane Permeability 107 9.3.2 Regulation of Biosynthesis 110
		9.3.3 Miscellaneous Regulation 111 Trends in Strain Improvement 112 Conclusion 113 References 114
10		oline 117 umihiro Yoshinaga) References 120
11		utamine 121 akashi Таснікі and Tatsurokuro Тоснікика)
	11.	1 Direct Fermentation 122 11.1.1 Production of Glutamine and N-Acetylglutamine by Glutamate-producing Bacteria 122 11.1.2 Production of Glutamine by Non-glutamate-producing Bacterium 124
	11.	.2 Enzymatic Synthesis of Glutamine 126 References 129
12		ginine, Citrulline, and Ornithine 131

94

98 *