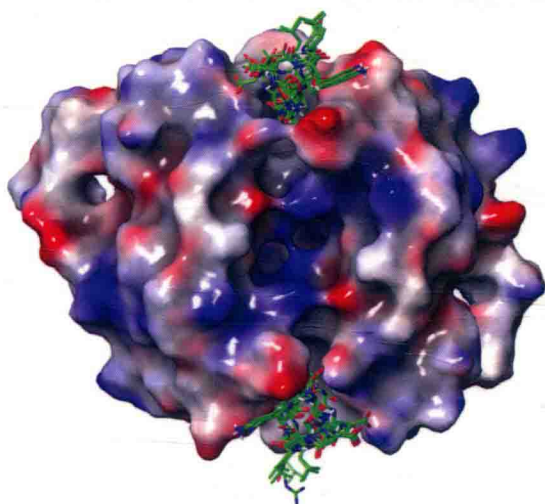


Advances in
PROTEIN CHEMISTRY
and **STRUCTURAL**
BIOLOGY

Insights into Enzyme Mechanisms and Functions
from Experimental and Computational Methods



VOLUME 105

Edited by
Christo Z. Christov





VOLUME ONE HUNDRED AND FIVE

ADVANCES IN PROTEIN CHEMISTRY AND STRUCTURAL BIOLOGY

Insights into Enzyme Mechanisms
and Functions from Experimental
and Computational Methods

Edited by

CHRISTO Z. CHRISTOV

*Faculty of Health and Life Sciences,
Northumbria University,
Newcastle upon Tyne,
United Kingdom*



AMSTERDAM • BOSTON • HEIDELBERG • LONDON
NEW YORK • OXFORD • PARIS • SAN DIEGO
SAN FRANCISCO • SINGAPORE • SYDNEY • TOKYO

Academic Press is an imprint of Elsevier



Academic Press is an imprint of Elsevier
50 Hampshire Street, 5th Floor, Cambridge, MA 02139, United States
525 B Street, Suite 1800, San Diego, CA 92101-4495, United States
The Boulevard, Langford Lane, Kidlington, Oxford OX5 1GB, United Kingdom
125 London Wall, London, EC2Y 5AS, United Kingdom

First edition 2016

Copyright © 2016 Elsevier Inc. All rights reserved.

No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopying, recording, or any information storage and retrieval system, without permission in writing from the publisher. Details on how to seek permission, further information about the Publisher's permissions policies and our arrangements with organizations such as the Copyright Clearance Center and the Copyright Licensing Agency, can be found at our website: www.elsevier.com/permissions.

This book and the individual contributions contained in it are protected under copyright by the Publisher (other than as may be noted herein).

Notices

Knowledge and best practice in this field are constantly changing. As new research and experience broaden our understanding, changes in research methods, professional practices, or medical treatment may become necessary.

Practitioners and researchers must always rely on their own experience and knowledge in evaluating and using any information, methods, compounds, or experiments described herein. In using such information or methods they should be mindful of their own safety and the safety of others, including parties for whom they have a professional responsibility.

To the fullest extent of the law, neither the Publisher nor the authors, contributors, or editors, assume any liability for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions, or ideas contained in the material herein.

ISBN: 978-0-12-804825-2

ISSN: 1876-1623

For information on all Academic Press publications
visit our website at <https://www.elsevier.com/>



**Working together
to grow libraries in
developing countries**

www.elsevier.com • www.bookaid.org

Publisher: Zoe Kruze

Acquisition Editor: Alex White

Editorial Project Manager: Helene Kabes

Production Project Manager: Surya Narayanan Jayachandran

Cover Designer: Mark Rogers

Typeset by SPi Global, India



VOLUME ONE HUNDRED AND FIVE

ADVANCES IN PROTEIN CHEMISTRY AND STRUCTURAL BIOLOGY

Insights into Enzyme Mechanisms
and Functions from Experimental
and Computational Methods

CONTRIBUTORS

G. Altankov

Institute for Bioengineering of Catalonia (IBEC); Institució Catalana de Recerca i Estudis Avançats (ICREA), Barcelona, Spain

A. Bidon-Chanal

Institute of Biomedicine (IBUB), Faculty of Pharmacy and Food Science, University of Barcelona, Santa Coloma de Gramenet, Spain

G.W. Black

Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

L. Capece

Universidad de Buenos Aires, Facultad de Ciencias Exactas y Naturales, INQUIMAE-CONICET, Ciudad Universitaria, Ciudad de Buenos Aires, Argentina

S.J. Charnock

Prozomix Limited, Haltwhistle, Northumberland, United Kingdom

N.M. Coelho

Institute for Bioengineering of Catalonia (IBEC), Barcelona, Spain

D.J. Cook

Prozomix Limited, Haltwhistle, Northumberland; Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

K. Cook

Thermo Fisher Scientific, Hemel Hempstead, Hertfordshire, United Kingdom

C. Estarellas

Institute of Biomedicine (IBUB), Faculty of Pharmacy and Food Science, University of Barcelona, Santa Coloma de Gramenet, Spain

D.A. Estrin

Universidad de Buenos Aires, Facultad de Ciencias Exactas y Naturales, INQUIMAE-CONICET, Ciudad Universitaria, Ciudad de Buenos Aires, Argentina

E. Fernández-de Gortari

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

J.D. Finnigan

Prozomix Limited, Haltwhistle, Northumberland; Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

E. Gallicchio

Brooklyn College, Brooklyn; The Graduate Center of the City University of New York, New York, NY, United States

L.J. Kay

Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

D. Kilburg

Brooklyn College, Brooklyn; The Graduate Center of the City University of New York, New York, NY, United States

V. Llopis-Hernández

Center for Biomaterials and Tissue Engineering, Universidad Politécnica de Valencia, Valencia, Spain; School of Engineering, University of Glasgow, Glasgow, United Kingdom

F.J. Luque

Institute of Biomedicine (IBUB), Faculty of Pharmacy and Food Science, University of Barcelona, Santa Coloma de Gramenet, Spain

J.L. Medina-Franco

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

O. Méndez-Lucio

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

A. Peña-Castillo

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

F.D. Prieto-Martínez

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

M. Salmerón-Sánchez

School of Engineering, University of Glasgow, Glasgow, United Kingdom

C. Seira

Institute of Biomedicine (IBUB), Faculty of Pharmacy and Food Science, University of Barcelona, Santa Coloma de Gramenet, Spain

T.K. Smulders-Srinivasan

Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

M. Soundararajan

Faculty of Health and Life Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

PREFACE

Enzymes play a crucial role in molecular processes in living organisms. They are characterized with the ability to increase considerably the rate of chemical reactions in the cells but also with high specificity toward their substrates. The key functions of enzymes in cell biology and unique properties make them important targets for drug design and applications in biotechnology, protein engineering, and synthetic biology.

Recent developments in experimental methods contribute for a rapid growth in our understanding about structure–function relationships in enzymes. Structural methods such as X-ray crystallography and NMR are currently widely used to determine the three-dimensional structures of enzymes, and their complexes with substrates and inhibitors. Homology modeling methods received growing applications to predict 3D structures of enzymes, when experimental structures are not available. In addition, spectroscopic and kinetic methods are broadly applied to complement structural information and understand enzyme structure–function relationships.

The applications of computational methods to enzyme mechanisms of functions received a large increase over the last years. The growth in computational power, the development of parallel supercomputers, and GPUs made possible modeling studies to be performed for very large biomolecular systems, containing hundreds of thousands of atoms. Nowadays, we can routinely perform molecular dynamics in order to understand the conformational flexibility of enzyme–ligand complexes in microsecond timescales, to apply combined quantum mechanical and molecular mechanical methods to explore the reaction mechanisms and the origins of the catalytic power in enzymes and to implement methods for prediction of the binding orientations and free energies of enzyme–ligand complexes.

The present thematic volume of *Advances in Protein Chemistry and Structural Biology* is aimed to address some of the recent developments in enzymes. The selected contributions are focused on state-of-the-art applications of computational and experimental methods to understand different aspects of structure–function relationships and mechanisms of enzymes and their applications in biotechnology, cell biology, drug design, and biomaterials.

CHRISTO Z. CHRISTOV

Faculty of Health and Life Sciences, Northumbria University,
Newcastle upon Tyne, United Kingdom

CONTENTS

Contributors

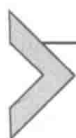
vii

Preface

ix

1. Molecular Modeling and Chemoinformatics to Advance the Development of Modulators of Epigenetic Targets: A Focus on DNA Methyltransferases	1
F.D. Prieto-Martínez, A. Peña-Castillo, O. Méndez-Lucio, E. Fernández-de Gortari, and J.L. Medina-Franco	
1. Introduction	2
2. Progress on Chemical Information	4
3. Chemoinformatic Studies of DNMTs	9
4. VS: Hit Identification and Optimization	16
5. Computer-Assisted Drug Repurposing	19
6. Food Chemicals as Potential Modulators of DNMTs and Other Epigenetic Targets	20
7. Concluding Remarks	21
Acknowledgments	22
References	23
2. Recent Advances in Computational Models for the Study of Protein–Peptide Interactions	27
D. Kilburg and E. Gallicchio	
1. Introduction	28
2. Theory and Methods	30
3. Sample Applications	39
4. Concluding Remarks	51
References	51
3. Structural Plasticity in Globins: Role of Protein Dynamics in Defining Ligand Migration Pathways	59
C. Estarellas, L. Capece, C. Seira, A. Bidon-Chanal, D.A. Estrin, and F.J. Luque	
1. Introduction	60
2. Ligand Migration Pathways in the Globin Superfamily: Structure–Function Relationships	63
3. The Coordination of the Heme Group and Its Role in Ligand Binding	73

4. Conclusions	76
Acknowledgments	77
References	77
4. Dynamic Reorganization and Enzymatic Remodeling of Type IV Collagen at Cell–Biomaterial Interface	81
N.M. Coelho, V. Llopis-Hernández, M. Salmerón-Sánchez, and G. Altankov	
1. Introduction	82
2. Material and Methods	85
3. Results and Discussion	91
4. Conclusive Remarks	100
References	101
5. Cytochromes P450: History, Classes, Catalytic Mechanism, and Industrial Application	105
D.J. Cook, J.D. Finnigan, K. Cook, G.W. Black, and S.J. Charnock	
1. Background and History of Cytochromes P450	106
2. P450 Classes	110
3. P450 Family Nomenclature	114
4. Catalytic Cycle	115
5. Uncoupling	117
6. P450s in Industry	119
References	123
6. Understanding the Multifaceted Role of Human Down Syndrome Kinase DYRK1A	127
L.J. Kay, T.K. Smulders-Srinivasan, and M. Soundararajan	
1. Introduction	128
2. Minibrain in <i>Drosophila</i>	129
3. Role of DYRK1A in Infection and Immunity	132
4. Cardiovascular Function of DYRK1A	136
5. Role of DYRK1A in Cancer	139
6. Understanding the Neuronal Function of DYRK1A	148
7. Concluding Remarks	157
References	158
<i>Author Index</i>	173
<i>Subject Index</i>	195



Molecular Modeling and Chemoinformatics to Advance the Development of Modulators of Epigenetic Targets: A Focus on DNA Methyltransferases

F.D. Prieto-Martínez, A. Peña-Castillo, O. Méndez-Lucio,
E. Fernández-de Gortari, J.L. Medina-Franco¹

Facultad de Química, Universidad Nacional Autónoma de México, México City, México

¹Corresponding author: e-mail addresses: medinajl@unam.mx; jose.medina.franco@gmail.com

Contents

1. Introduction	2
2. Progress on Chemical Information	4
3. Chemoinformatic Studies of DNMTs	9
3.1 Characterization of Chemical Space: ERCS	9
3.2 Chemoinformatic-Based Pharmacophore Model	12
3.3 Activity Landscape Modeling	14
3.4 Quantitative Structure–Activity Relationships	15
4. VS: Hit Identification and Optimization	16
4.1 Novel VS Hits	16
4.2 Follow-Up of VS Hits	18
5. Computer-Assisted Drug Repurposing	19
6. Food Chemicals as Potential Modulators of DNMTs and Other Epigenetic Targets	20
7. Concluding Remarks	21
Acknowledgments	22
References	23

Abstract

In light of the emerging field of *Epi-informatics*, ie, computational methods applied to epigenetic research, molecular docking, and dynamics, pharmacophore and activity landscape modeling and QSAR play a key role in the development of modulators of DNA methyltransferases (DNMTs), one of the major epigenetic target families. The increased chemical information available for modulators of DNMTs has opened up the avenue to explore the epigenetic relevant chemical space (ERCS). Herein, we discuss

recent progress on the identification and development of inhibitors of DNMTs as potential epi-drugs and epi-probes that have been driven by molecular modeling and chemoinformatics methods. We also survey advances on the elucidation of their structure–activity relationships and exploration of ERCS. Finally, it is illustrated how computational approaches can be applied to identify modulators of DNMTs in food chemicals.



1. INTRODUCTION

DNA methylation is one of the most important epigenetic regulation mechanisms and it is mediated primarily by the family of enzymes DNA methyltransferases (DNMTs). This process involves the addition of a methyl group at the C-5 position of a DNA cytosine residue by the cofactor S-adenosyl methionine (SAM). This family of enzymes is formed by DNMT1, DNMT3A, DNMT3B, and DNMT3L. DNMT1 is associated with the maintenance of methylation patterns in DNA. DNMT3A and DNMT3B are *de novo* DNMTs and are able to transfer a methyl group in nonmethylated CpG's islands. DNMT3L is related to DNMT3A and DNMT3B enhancing their activity (Robertson, 2001).

DNMTs are promising epigenetic targets for the treatment of a number of diseases. For instance, DNA hypermethylation is related to cancer metastasis by silencing the expression genes linked to cell division. DNMTs are implicated in autoimmune diseases and inherited disorders (Gros et al., 2012) and are also promising molecular targets for the treatment of other chronic and degenerative diseases such as Alzheimer's and psychiatric conditions (Gros et al., 2012), and diabetes (Arguelles, Meruvu, Bowman, & Choudhury, 2016). As such, inhibitors of DNMTs (DNMTis) are attractive compounds to be developed as clinical candidates for diverse diseases either alone or as part of combination therapies. Moreover, the development of DNMTis as *epi-probes* is also attractive for the further biochemical understanding of the role of DNMTs in epigenetics. In fact, there are not known good molecular probes for DNMTs although selective probe molecules have been developed for other epigenetic targets (Arrowsmith et al., 2015).

Thus far, 5-azacytidine and decitabine (Fig. 1) are DNMTis that have been approved by the USA Food and Drug Administration—FDA—for the treatment of myelodysplastic syndrome (Derissen, Beijnen, & Schellens, 2013). However, these two compounds are prodrugs that need to be incorporated into DNA. Their high toxicity and low specificity have prompted the search for specific inhibitors preferably of the nonnucleosidic type. To date, several nonnucleosidic inhibitors have been reported which

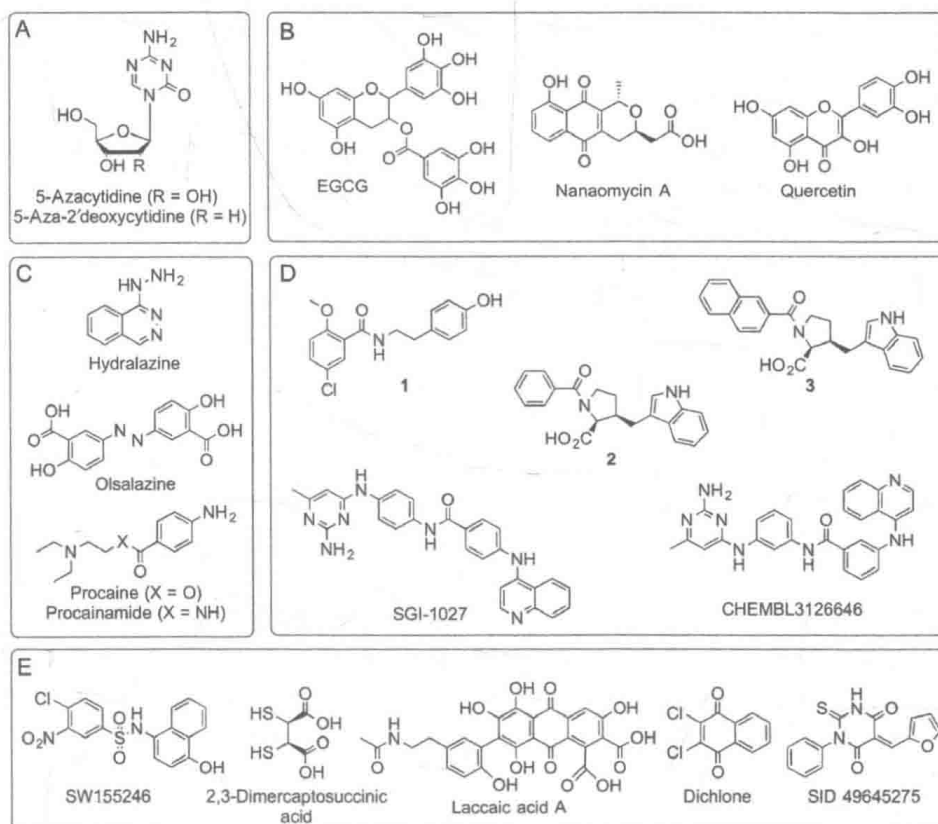


Fig. 1 Representative DNMT inhibitors and compounds with proposed demethylating properties. Compounds are classified by their source: (A) approved for clinical use, (B) natural products, (C) drugs approved for other indications, (D) synthetic compounds coming from optimization programs, and (E) molecules obtained from high-throughput screening.

have been identified from diverse sources such as drugs approved for other indications, natural products, virtual (in silico) screening, high-throughput screening, and synthetic compounds including molecules initially identified from computational screening and later optimized using medicinal chemistry approaches. The chemical structures of representative compounds are shown in Fig. 1. In the figure, compounds are classified by their source in five major groups, namely; DNMTis approved for clinical use (group A), natural products (B), drugs approved for other indications (C), synthetic compounds coming from optimization programs (D), and molecules obtained from high-throughput screening (E). The reader is also referred to reviews of DNMTis that have been published (Erdmann, Arimondo, & Guianvarc'h, 2016; Erdmann, Halby, Fahy, & Arimondo, 2014; Guianvarc'h & Arimondo, 2014).

Computational approaches have played a major role in the identification, optimization, and understanding of the biological activity of DNMTs at the molecular level. Application of *in silico* techniques continues to increase not only for DNMTs but also for several other epigenetic targets as reflected in the emerging research field called *Epi-informatics* (Medina-Franco, 2016). Reviews on the progress of computational approaches applied to DNMT have been published (Medina-Franco, Méndez-Lucio, Yoo, & Dueñas, 2015; Yoo & Medina-Franco, 2012). However, since these last reviews, major contributions from computational applications have been published in subsequent studies. In particular, a significant and relevant amount of structure–activity information (SAI) has been released and stored in public databases. Herein, we review recent advances on the chemical information resources that have been made available recently to chart the chemical space of DNMTs and elucidate the SAR of DNMTs. We also review major progress on the identification and optimization of hit compounds as well as on the computer-aided discovery of novel hits with putative novel binding sites. All together, these studies emphasize the continued synergy between molecular modeling and chemoinformatic to further advance epigenetic drug and probe development. Representative computational studies, techniques, and major outcomes discussed throughout this chapter are summarized in Table 1.



2. PROGRESS ON CHEMICAL INFORMATION

The SAI of compounds tested as DNMTs has been growing in the last few years. In a recent study, Fernández-de Gortari et al. surveyed public resources and collected chemical databases that contain SAI of DNMTs. Major compound collections included human epigenetic enzyme and modulator database (HEMD) (Huang et al., 2012), ChEMBL (Gaulton et al., 2012), and Binding Database (Liu, Lin, Wen, Jorissen, & Gilson, 2007). An additional database is EpiDBase (Loharch et al., 2015), a curated epigenetic database that includes 11,422 small molecules with activity against different epigenetic targets.

In addition to the availability of molecular databases with experimental data coming from research publications, advances in assay developments and rapid screening technologies for DNMTs (Medina-Franco, Yoo, & Dueñas-Gonzalez, 2015) are facilitating the access to medium- and high-throughput screening data. Table 2 summarizes the information of representative

Table 1 Representative Computational Studies on DNMT Is Recently Reported

Study	Approach	Major Outcomes	References
ERCS focused on DNMTs: comparison with other general compound collections	Chemoinformatic analysis based on physicochemical properties, molecular fingerprints, and scaffolds	The structure of DNMTis is diverse; occupy the traditional chemical space of drugs but also cover other regions. Four tentative privileged epigenetic scaffolds were identified	Fernández-de Gortari and Medina-Franco (2015)
ERCS focused on DNMTs: comparison with epigenetic reference databases	Chemoinformatic analysis including metrics of structural complexity	The structures of DNMTis are different from inhibitors of histone deacetylases and bromodomains. There is a large potential to develop DNMTis with increased complexity	Prieto-Martínez, Fernández-de Gortari, Méndez-Lucio, Medina-Franco (2016)
Chemoinformatic-based pharmacophore model	Pharmacophore models based on the predicted protein–ligand interaction profiles of selected chemical scaffolds enriched with active compounds	A model with three main pharmacophoric points was identified. The model had high sensitivity and specificity in a validation study	This chapter
Activity landscape modeling	Systematic comparison of structure similarity and potency difference of 280 compounds using SAS maps. The landscape of two major groups of inhibitors was assessed	Inhibitors related to the cofactor have a rough landscape with significant activity cliffs. Structure-based interpretation of an activity cliffs suggested distinct molecular interactions in the binding site. Also, a novel “activity landscape sweeping” approach was proposed	Naveja and Medina-Franco (2015)

Continued

Table 1 Representative Computational Studies on DNMT Is Recently Reported—cont'd

Study	Approach	Major Outcomes	References
Linear discriminant analysis-based QSAR and virtual screening	QSAR model for 47 molecules was used to classify natural products as active/inactive; predicted active were docked with DNMT1 and 3A	Six natural products as consensus hits from two docking programs were suggested as potential inhibitors of DNMTs	Maldonado-Rojas, Oliviero-Verbel, and Marrero-Ponce (2015)
Virtual screening with experimental validation	Docking-based screening of commercial compounds followed by analog searching	Identification of compound DC_05, DC_501, and DC_517 as low micromolar and selective inhibitors of DNMT1. DC_517 had activity in cell-based assays	Chen et al. (2014)
Molecular dynamics and virtual screening with experimental validation	Ensemble docking using conformations of the protein obtained from molecular dynamics. Multistep docking-based screening of a commercial compound collection	Two experimentally validated hits were identified: ASINEX ID BAS 12771472 and BAS 00872020. The former compound showed in vivo activity. A newly putative binding site in hDNMT1 is proposed	Joshi, Rajpathak, Narwade, and Deobagkar (2016)
Docking of newly designed selective compounds inspired in a virtual screening hit	Flexible docking	The selectivity of the most potent compounds toward DNMT3 over DNMT1 is rationalized at the molecular level	Aldawsari et al. (2016)
Toward computer-assisted drug repurposing	Similarity searching and data fusion	Potential DNMT inhibitors are identified	Naveja, Dueñas-González, and Medina-Franco (2016)

Table 2 Representative Confirmatory Assays in PubChem Related to DNMTs

AID	Name	Title	Assay Data	Source
1066238	Inhibition of human recombinant DNMT1 expressed in H19 cells assessed as inhibition of tritiated methyl incorporation from [3H]-labeled AdoMet into hemimethylated DNA duplex after 2 h by liquid scintillation counting analysis	Synthesis and evaluation of analogs of <i>N</i> -phthaloyl-L-tryptophan (RG108) as inhibitors of DNA methyltransferase 1	2 active, 9 tested	ChEMBL
461004	Inhibition of human recombinant DNMT3B expressed in baculovirus-insect cell system by scintillation counting	Novel and selective DNA methyltransferase inhibitors: docking-based virtual screening and experimental evaluation	2 active, 1 activity $\leq 1 \mu\text{M}$, 4 tested	ChEMBL
424255	Inhibition of human recombinant DNMT1	Constrained (L)-S-adenosyl-L-homocysteine (SAH) analogs as DNA methyltransferase inhibitors	18 active, 2 activity $\leq 1 \mu\text{M}$, 25 tested	ChEMBL
736572	Inhibition of human DNMT1 using AdoMet and poly dI-dC after 2 h by radioactive assay	Synthetic approaches to DNMT inhibitor SGI-1027 and effects on the U937 leukemia cell line	1 active, 1 tested	ChEMBL
424222	Inhibition of human recombinant DNMT3b2 expressed in baculovirus infected high five insect cells	SAR around (L)-S-adenosyl-L-homocysteine, an inhibitor of human DNA methyltransferase (DNMT) enzymes	16 active, 7 activity $\leq 1 \mu\text{M}$, 20 tested	ChEMBL
675178	Inhibition of DNMT1 in human HeLa cell nuclear extract assessed as methylated substrate level at 10 μM by ELISA	New cytosine derivatives as inhibitors of DNA methylation	2 tested	ChEMBL

Continued

Table 2 Representative Confirmatory Assays in PubChem Related to DNMTs—cont'd

AID	Name	Title	Assay Data	Source
657293	Inhibition of human DNMT1 using oligonucleotide 2 as substrate after 5000 s by micro plate reader based real-time break-light assay	Development of rationally designed DNA N6 adenine methyltransferase inhibitors	1 active, 1 activity $\leq 1 \mu\text{M}$, 3 tested	ChEMBL
257909	Inhibitory activity against DNA methyl transferase in leukemic NALM6 cells	Discovery of two novel, small-molecule inhibitors of DNA methylation	2 tested	ChEMBL
631009	Inhibition of human recombinant DNMT1 expressed in Sf9 cells assessed as remaining activity at 2 mM after 3 h by scintillation counting in presence of [3H] S-adenosyl methionine	Synthesis and biochemical evaluation of delta(2)-isoxazoline derivatives as DNA methyltransferase 1 inhibitors	26 tested	ChEMBL
1073761	Cytotoxicity against human U937 cells after 48 h by trypan blue exclusion assay	Selective nonnucleoside inhibitors of human DNA methyltransferases active in cancer including in cancer stem cells	6 active, 6 tested	ChEMBL
602386	Dose response confirmation of DNMT1 inhibitors in a fluorescent molecular beacon assay	Dose-response confirmation of DNMT1 inhibitors in a fluorescent molecular beacon assay	179 active, 1 activity $\leq 1 \mu\text{M}$, 200 tested	Burnham Center for Chemical Genomics
613112	Inhibition of DNMT in cell-free system	Epigenetic profiling of the antitumor natural product psammapiin A and its analogs	1 active, 1 activity $\leq 1 \mu\text{M}$, 1 tested	ChEMBL
890	Confirmation concentration-response assay for epigenetic modulators	Confirmation concentration-response assay for epigenetic modulators	24 active, 1 activity $\leq 1 \mu\text{M}$, 51 tested	NCGC
597	qHTS assay for epigenetic modulators	qHTS assay for epigenetic modulators	59 active, 68401 tested	NCGC