Methods of DNA and RNA Sequencing

Edited by

Sherman M. Weissman, M.D.

Methods of DNA and RNA Sequencing

Edited by

Sherman M. Weissman, M.D.

Library of Congress Cataloging in Publication Data

Main entry under title:

ISBN 0-03-059174-0

Methods of DNA and RNA sequencing.

Includes bibliographical references and index.

Nücleotide sequence.
 Deoxyribonucleic acid—Analysis.
 Ribonucleic acid—Analysis.
 Weissman, Sherman M.
 QP625.N89M47
 1983
 574.87'328
 82-22261

Published in 1983 by Praeger Publishers CBS Educational and Professional Publishing a Division of CBS Inc. 521 Fifth Avenue, New York, New York 10175 U.S.A.

© 1983 by Praeger Publishers

All rights reserved

456789 052 98765432

Printed in the United States of America

LIST OF CONTRIBUTORS

Marvin H. Caruthers
Department of Chemistry
University of Colorado
Boulder, CO 80309
USA

David J. Galas
Département de Biologie Moléculaire
Université de Genève
Geneva
Switzerland

G. Nigel Godson
Biochemistry Department
New York University Medical
Center
New York, N.Y. 10016
USA

Yoshiyuki Kuchino
Biology Division
National Cancer Center Research
Institute
Tsukiji, Chuo-Ku
Tokyo, Japan

Allan Marshall Maxam Sidney Farber Cancer Institute Charles A. Dana Cancer Center Boston, MA 02115 USA Stephen M. Mount
Department of Molecular Biophysics
and Biochemistry
Yale University
New Haven, CT 06510
USA

Susumu Nishimura
Biology Division, National Cancer
Center Research Institute
Tsukiji, Chuo-Ku
Tokyo, Japan

Debra A. Peattie
Department of Biochemistry
Stanford University Medical Center
Stanford, CA 94305
USA

Erika Randerath
Department of Pharmacology
Baylor College of Medicine
Texas Medical Center
Houston, TX 77030
USA

Kurt Randerath
Department of Pharmacology
Baylor College of Medicine
Texas Medical Center
Houston, TX 77030
USA

Albert Schmitz
Départment de Biologie Moléculaire
Université de Genève
Geneva
Switzerland

Thomas Shenk
Department of Microbiology
Health Sciences Center
State University of New York
Stony Brook, N.Y. 11794
USA

Michael Smith
Department of Biochemistry
Faculty of Medicine
University of British Columbia
Vancouver, B.C. V6T 1W5
Canada

Joan A. Steitz
Department of Molecular Biophysics
and Biochemistry
Yale University
New Haven, CT 06510
USA

Sherman M. Weissman
Department of Human Genetics
Yale University School
of Medicine
New Haven, CT 06510
USA

PREFACE

The entire structure of a free-living biologic unit is, to a first approximation, encoded in its DNA. Increasingly efficient and universal approaches for cloning discrete fragments of genetic DNA defined by restriction endonuclease cleavage sites have made available in principle each part of even the most complex genomes in large amounts and of sufficient purity for detailed analysis. Development of DNA sequencing techniques based principally on restriction endonuclease-generated fragments and dependent on gel fractionation methods that are capable of resolving the oligonucleotides that differ by 0.2% or less in their chain length have made it possible for many laboratories to determine complete sequences for the genetic material of particular interest to their research. Fortunately for the field, the inventors have made available fairly detailed protocols that make it possible to routinely reproduce the methods with clean results. A particularly exemplary protocol has been provided by Drs. Maxam and Gilbert for the method of chemical sequencing of DNA. One purpose of the present volume has been to update a simple set of such protocols for the more commonly used methods of nucleic acid labeling and sequence analysis, together with a discussion of the principles underlying the methods and some presentation of discussion of common sources of technical difficulties.

The chemical synthesis of deoxynucleotides has become a fundamental tool, both in sequence analysis and in in vitro mutagenesis and further characterization of biologically active sequences. Dr. Caruthers has included a chapter providing an update on discussion of the current methods that are applicable for laboratories not specialized in organic chemistry, and Dr. Smith has prepared an extensive discussion of the application of synthetic oligonucleotides to problems of molecular biology. Dr. Godson has reviewed the enzymatic approaches to DNA sequencing, and Dr. Maxam has provided a historical account of the development of chemical methods for DNA sequencing. It has become an increasing challenge to interpret

sequences, once available, particularly through the use of valuable chemical methods that have been developed for studying interaction of purified proteins or protein complexes with nucleic acids, and these footprinting methods and extensive discussion of them by Drs. Schmitz and Galas are also included in the present volume.

Finally, a variety of techniques, including the classical techniques of prokaryotic genetics and *in vitro* mutagenesis of genes in functioning animal cells, followed by reintroduction of genes into cellular milieus *in vitro* enzymologic studies, have partly defined at least major subclasses of sequences responsible for transcription initiation and termination of prokaryotes and begun to provide a corresponding definition for eukaryotic sequences involved in these processes, as well as in the uniquely eukaryotic RNA splicing mechanisms. "State of the art" reviews on sequence features involved in initiation and termination of transcription (Dr. Shenk), and RNA splicing (Drs. Mount and Steitz) have been included.

The aim of the editor throughout has been to obtain experts to review their fields with the intention of describing methods in sufficient detail to facilitate ready application in laboratories without previous experience in the field and to provide discussion both of the theoretic underpinnings of the method and of the potential pitfalls. An effort has been made to minimize reduplication of material that has been authoritatively and lucidly reviewed elsewhere; for example, there is no discussion of in vitro mutagenesis, or of features of translation initiation sites, since several excellent reviews are available. One major feature of nucleic acid sequence analysis not covered in the present volume is the use of computers, both to expedite handling of the data and to reveal more detailed information than can be seen simply by visual inspection. This is a rapidly burgeoning subject in which many laboratories have their own favorite "data-crunching" systems, and increasing progress has been made in the theoretical aspects of problems related to the detection of potential base-pairing structures, analyses of homology among related sequences, etc. The editor can only refer the interested reader to primary papers such as the excellent collection recently published in Nucleic Acids Research (Jan., 1982) and to caution the novice that accessibility to computer analysis of data is almost essential for any extensive sequencing project. It is the hope of the authors that the present set of essays will expedite the work of young or novice investigators and provide a convenient reference or refresher for experienced workers in the field.

CONTENTS

| List of Contributors | برواوركان فكليات | v |
|--|------------------|------|
| Preface | | xvii |
| Acknowledgments | | 4/8 |
| 1 | | |
| NEW METHODS FOR CHEMICALLY SYNT DEOXYOLIGONUCLEOTIDES | THESIZING | 1 |
| Marvin H. Caruthers | | |
| General outline of DNA chemical synthmethodologies | esis | 2 |
| A procedure for the rapid chemical synt deoxyoligonucleotides | thesis of | 5 |
| The support | | 6 |
| The synthesis cycle | | 7 |
| Isolation of deoxyoligonucleotides | | 11 |
| Experimental section | | 13 |
| General methods | | 13 |
| Synthesis of the support | | 13 |
| Synthesis of deoxynucleosidephos | phoramidites | 15 |
| Outline of the synthesis cycle | | 16 |
| Isolation of synthetic deoxyoligon | ucleotides | 17 |
| Acknowledgments | | 17 |
| References | | 18 |

| PRO | THETIC OLIGODEOXYRIBONUCLEOTIDES AS DEES FOR NUCLEIC ACIDS AND AS PRIMERS IN UENCE DETERMINATION | 23 |
|------|--|----|
| Mich | nael Smith | |
| ٧ | Introduction | 23 |
| | Background | 24 |
| | Oligonucleotide length and duplex stability | 25 |
| | Effect of base composition on oligonucleotide duplex stability | 29 |
| | Stability of oligodeoxyribonucleotide- oligoribonucleotide duplexes | 31 |
| V | Influence of base-pair mismatch on oligonucleotide duplex stability | 34 |
| | Ionic strength, polyvalent cations, and oligodeoxyribonucleotide duplex stability | 37 |
| | Oligonucleotide duplexes form very rapidly | 38 |
| | Length of a genetically unique oligonucleotide | 39 |
| | Synthetic oligodeoxyribonucleotide as probe for genomic DNA fragment purification | 39 |
| | Oligodeoxyribonucleotides as probes for monitoring isolation of a genomic DNA clone | 41 |
| # | Oligodeoxyribonucleotides as probes for monitoring isolation of a cDNA clone | 46 |
| | Oligodeoxyribonucleotides as primers for reverse transcriptase to produce specific cDNA probes | 48 |
| | Use of synthetic oligodeoxyribonucleotides to prepare double-strand cDNAs | 52 |
| | Oligodeoxyribonucleotides as primers for DNA sequence determination | 53 |
| Ġ. | Use of synthetic oligodeoxyribonucleotides as primers for RNA sequence determination | 55 |
| | Conclusion | 57 |
| | References | 57 |

| 3 | | |
|------|---|-----|
| 13 | QUENCING DNA BY THE SANGER CHAIN | |
| | RMINATION METHOD | 69 |
| G. N | ligel Godson | |
| | Introduction | 69 |
| | Principle of primed synthesis sequencing and the chain termination method | 70 |
| | Use of the filamentous phage cloning vectors for chain termination sequencing | 73 |
| | A model system for sequencing a 2-kB piece of DNA | 74 |
| | Special strategies in cloning and sequencing | 77 |
| | Cloning DNA fragments in a directed orientation | 78 |
| | Cloning internal DNA fragments only | 79 |
| | Subcloning in situ | 79 |
| | Turning cloned fragments around | 79 |
| | Use of internal primers | 80 |
| | Checking the sequence with restriction enzymes | 81 |
| | Problems of random sequencing- | 82 |
| | Strategies for sequencing large pieces of DNA (10-100 kB) | 82 |
| | Appendixes | 83 |
| | Acknowledgments | 108 |
| | References | 108 |
| | | |
| 4 | | |
| NU | CLEOTIDE SEQUENCE OF DNA | 113 |
| Alla | n Marshall Maxam | |
| | Introduction | 113 |
| | Sequencing bases in nucleic acids | 114 |

115

116

Distinguishing the bases

Ordering nucleotides

| | Phase one: Tract and block methods | 123 |
|-----|---|-----|
| | Pyrimidine tracts | 125 |
| | Copying DNA into RNA | 126 |
| | Ribosubstitution | 128 |
| | Phase two: Wandering-spot methods | 129 |
| | Phase three: Base-specific nested-segment methods | 132 |
| | The chemical method | 133 |
| | Comparison and assessment of the chemical and enzymatic methods of DNA sequencing | 144 |
| | Double-stranded DNA | 144 |
| | Single-stranded DNA | 147 |
| | Amount and specific activity of DNA | 147 |
| | Sequencing labor | 148 |
| | Sequence accuracy | 148 |
| | Biologically significant DNA ends and modifications | 149 |
| | References | 150 |
| | Addendum by Sherman M. Weissman, M.D. | 164 |
| | References | 166 |
| | The first and the first in the first | |
| 5 | | |
| SEL | LECTED POSTLABELING PROCEDURES FOR BASE MPOSITION AND SEQUENCE ANALYSIS OF CLEIC ACIDS | 169 |
| | a Randerath & Kurt Randerath | |
| | Radioactive derivative (postlabeling) methods for structural analysis of nucleic acids | 169 |
| | Fluorography and screen-intensified autoradiography for enhanced detection of radioisotopes (³ H, ³² P, ¹²⁵ I) on chromatograms or gels | 173 |
| | General aspects | 173 |
| | Materials | 174 |
| | Fluorography for the detection of ³ H on thin-layer chromatograms | 175 |

| Detection of ³ H in polyacrylamide, mixed polyacrylamide/agarose, or agarose gels by fluorography | 176 |
|--|---------|
| Detection of ³² P and ¹²⁵ I on chromatograms or gels by screen-intensified autoradiography | 178 |
| ³ H derivative method for base analysis of RNA | 178 |
| General aspects | 178 |
| Outline of method | 179 |
| Materials | 180 |
| Procedure | 180 |
| Comments | 186 |
| Radioiodination method for ribonucleoside analysis | 188 |
| General aspects | 188 |
| Outline of method | 188 |
| Materials | 190 |
| Procedure | 190 |
| Comments | 192 |
| ³² P derivative method for base analysis of normal | Sec. 5. |
| and mutagen/carcinogen-modified DNA | 192 |
| General aspects | 192 |
| Outline of method | 193 |
| Materials | 194 |
| Procedures | 196 |
| Comments | 198 |
| ³ H derivative method for sequence analysis of RNA | 200 |
| General aspects | 200 |
| Outline of method | 200 |
| Combined ³ H/ ³² P derivative method for sequence | |
| analysis of RNA | 201 |
| General aspects | 201 |
| Outline of method | 201 |
| ³² P gel readout method entailing partial enzymatic | 000 |
| degradation of end-labeled RNA | 202 |
| Conoral aspects | 97(19 |

| | Outline of method | 203 |
|------|---|-----|
| | Materials | 204 |
| | Procedures | 205 |
| | Comments | 210 |
| | ³² P thin-layer readout method based on single-hit | |
| | chemical cleavages of RNA | 210 |
| Υ | General aspects | 210 |
| | Outline of method | 211 |
| | Materials | 212 |
| | Procedure | 214 |
| 7 | Comments | 220 |
| | Sequence analysis of large RNAs | 223 |
| | Conclusions | 224 |
| | Acknowledgments | 225 |
| | References | 225 |
| | | |
| 6 | | |
| CHA | ARACTERIZATION OF MODIFIED NUCLEOSIDES | |
| IN t | RNA | 235 |
| Susu | umu Nishimura & Yoshiyuki Kuchino | |
| | Identification of modified nucleosides by two- | |
| | dimensional thin-layer cellulose chromatography | |
| | and paper chromatography | 239 |
| | Identification of modified nucleosides by high- | |
| | pressure liquid chromatography (HPLC) | 243 |
| 7 | Analysis of modified nucleosides by mass spectrometry | 243 |
| | Analysis of modified nucleotides by postlabeling | |
| | procedures | 245 |
| | Identification of modified nucleotides during tRNA sequencing | 245 |
| | Identification of modified nucleotides in total | 07- |
| | tRNA by the postlabeling procedure | 251 |
| H- | Structural elucidation of unknown modified nucleosides | 252 |

| Mat | terials | 254 |
|------------------|--|-----|
| Ref | erences | 255 |
| | | |
| | | |
| 7 | · · · · · · · · · · · · · · · · · · · | |
| DIRECT END-LA | CHEMICAL METHOD FOR SEQUENCING ABELED RIBONUCLEIC ACIDS | 261 |
| Debra A | . Peattie | |
| | nciples of the chemical RNA sequencing ctions | 262 |
| | Chemical base modification | 264 |
| | Aniline-induced strand scission | 267 |
| En | d-labeling the RNA | 270 |
| | 3'-end-labeling with (5'-32P)pCp and T4 RNA ligase | 270 |
| | 5'-end-labeling with ³² P | 273 |
| | Practical considerations | 274 |
| Bas | se-specific chemical cleavage reactions | 275 |
| | Procedures | 275 |
| | Practical considerations | 284 |
| Usi | ing the chemical reactions to probe RNA | 286 |
| | Guanosine reaction | 288 |
| | Cytidine reaction | 288 |
| | Adenosine reaction | 289 |
| | Practical Considerations | 289 |
| Ac | knowledgments | 290 |
| Re | ferences | 291 |
| Ap | pendix | 297 |
| | Outline for four base-specific reactions for chemical sequencing of end-labeled RNA | |
| | molecules | 297 |
| | Diagnostic table for aid in correcting | 300 |

| THE STUDY OF PROTEIN-DNA BINDING SPECIFICITY: DNase FOOTPRINTING | 305 |
|--|-----|
| Albert Schmitz & David J. Galas | |
| Introduction | 305 |
| Bovine pancreatic deoxyribonuclease I (DNase I) | 306 |
| Description of the method | 307 |
| The reactions | 310 |
| Typical footprinting reaction | 311 |
| Assignment of the DNase I-produced bands | 312 |
| Other enzymes suitable for footprinting reactions | 313 |
| Interactions analyzed by footprinting | 315 |
| Footprint of lac repressor on operator DNA | 315 |
| Footprints of RNA polymerase on the <i>lac</i> promoter | 320 |
| Footprints of CRP on the lac control region DNA | 327 |
| Footprint of DNA gyrase | 331 |
| Nuclease specificities | 333 |
| Extensions of the method | 334 |
| Concluding remarks | 335 |
| Acknowledgments | 336 |
| References | 336 |
| Addendum | 344 |
| | |
| 9 | |
| TRANSCRIPTION INITIATION AND TERMINATION SIGNALS | 349 |
| Thomas Shenk | |
| Prokaryotic promoters | 349 |
| Eukaryotic RNA polymerases | 351 |
| Cell-free transcription by eukaryotic polymerases | 351 |
| Polymerase III | 352 |
| Polymerase II | 352 |

| | Polymerase I | | 353 |
|------|--|----|-----|
| | 5'-ends and initiation sites | | 353 |
| | Polymerase III | | 353 |
| | Polymerase II | | 353 |
| | Polymerase I | | 354 |
| Ŗ | Transcriptional control regions | | 355 |
| | Polymerase III | | 355 |
| | Polymerase II | | 357 |
| | Polymerase I | | 368 |
| | Regulation of transcription by eukaryotic polymerases | | 368 |
| | Polymerase III | 20 | 370 |
| | Polymerase II | | 372 |
| | Polymerase I | | 374 |
| | Termination of transcription in prokaryotes | | 374 |
| | Termination of transcription in eukaryotes | | 375 |
| | Acknowledgments | | 377 |
| | References | | 377 |
| | | | |
| 10 | | | |
| | NALS FOR THE SPLICING OF EUKARYOTIC SSENGER RNA TRANSCRIPTS | | 399 |
| Step | hen M. Mount & Joan A. Steitz | | |
| | Splice junction signals | | 400 |
| | Are junction signals sufficient? | | 410 |
| | The versatility of splicing | | 412 |
| | What recognizes splicing signals? | | 417 |
| | The biochemistry of splicing | | 419 |
| | Why splicing? | | 421 |
| | Acknowledgments | | 423 |
| | References | | 424 |
| | | | |
| Inde | ex | | 437 |

New Methods for Chemically Synthesizing Deoxyoligonucleotides

MARVIN H. CARUTHERS

age of the company of the con-

Recent developments in the recombinant DNA field have created a need for sequence-defined deoxyoligonucleotides. These compounds are being used as primers and probes for isolating natural genes (Wallace et al., 1981; Gillam et al., 1977), for experiments involving site-directed mutagenesis (Smith, 1980), and for the synthesis of genes and gene control regions (Agarwal et al., 1970; Khorana, 1979; Itakura et al., 1977; Goedell, Kleid, et al., 1979; Goedell, Heyneker, et al., 1979; Wetzel et al., 1980; Edge et al., 1981). However, until recently the synthesis and isolation of deoxyoligonucleotides has been a difficult and time-consuming task. Ideally, chemical methods should be simple, rapid, versatile, and accessible to the nonchemist. This chapter outlines a synthetic methodology that satisfies all these criteria. The approach has been used successfully by nonchemists in my own laboratory and more recently by other research groups with limited backgrounds in nucleic acid chemistry.

This chapter is divided into three sections. The first section provides a general outline of various approaches that have been used for synthesizing deoxyoligonucleotides. Such an outline is important since the field is highly specialized and not generally familiar to most biologists and biochemists. The second section outlines and discusses our methodology. The final section is a presentation of procedural details useful for those interested in duplicating our approach.