GENERAL CYTOGENETICS

J. SYBENGA

GENERAL CYTOGENETICS

J. SYBENGA

University of Agriculture Wageningen



© North-Holland Publishing Company - 1972

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission of the copyright owner.

TWO I THE LIST OF S

This book was originally published in Dutch by A. Oosthoek Publishing Company, Utrecht, 1968, under the title "Cytogenetica".

ISBN North-Holland 0 7204 4119 6 ISBN American Elsevier 0 444 10398 8

20 tables, 94 illustrations

Publishers:

NORTH-HOLLAND PUBLISHING COMPANY - AMSTERDAM NORTH-HOLLAND PUBLISHING COMPANY, Ltd. - LONDON

Sole distributors for the U.S.A. and Canada:

AMERICAN ELSEVIER PUBLISHING COMPANY, INC. 52 Vanderbilt Avenue, New York, N.Y. 10017

Because of the sufficient of the examination of the description of the control of

PREFACE INNE TO JOHN A SOUL

The field of cytogenetics is wide and diversified. The different specializations (Drosophila polytene cytology, human karyotyping, plant cytogenetics, etc.) seem to have only few contact points and tend to be treated as separate entities. Yet they all have one common basis: the chromosome, its behaviour and the genetic consequences of this behaviour. It is gradually becoming clear that the chromosomes of all higher organisms, in composition and behaviour are fundamentally equal. Studying chromosomes in plants is quite relevant for understanding chromosomes in man and vice versa. It is important, therefore, to try to treat the entire science of cytogenetics as one unit. While such an approach may already be difficult with a simpler subject, it is truly complicated with this particularly wide field, certainly when the book must be kept within pocket size. There are a few ways out: one can generalize and consider only the major aspects in a simplified way that makes them understandable to the uninitiated student without further help. Or one can make the coverage wider and introduce some detail, hoping that the student has a sound basic knowledge of genetics and can count on someone to help him out when he gets baffled by the complexities of cytogenetic thought. The latter alternative was chosen, but is should be understood that the coverage still had to be far from complete.

The book contains a reasonable amount of systematically arranged information that may be used for reading assignments accompanying a cytogenetics course, and may perhaps replace extensive note-taking during lectures.

VIII Preface

Because of the suitability of plant chromosomes for experimental manipulation, their study has contributed considerably to the understanding of chromosome behaviour. It is for this reason that many examples have been taken from plant cytogenetics. This may perhaps help preventing unnecessary "rediscovery" in humans and other animals of phenomena that for decades have been known to occur in plants.

special responsibility of the companies en ann an an air an fhail an Immeleon an Immel an Air an ann ann an Air an Air an Air an Air an Air an Air an Aftering the production of the sustained to Assess by the party the Million of the reservoir of the three region by the making and the words in a region that "friendly activity in the company of the control of the co tring its at least property of the modern of the Marrian as the first transfer of the first property and the first property of the f selfine that and of the hydron - of the ear, required in their subjected, their without and recombined or their principles of their control of the solution of the amendment of significant and a store the new participation of small which to the state of real charge of learner travitation, and the property of the confidence of the forest and other the confidence of the confiden ne velloueren in a man de la company de la c with a titude, mortes of the many. It is the more of the expression of the college time only วิทุริยาร์ ซึ่งสติ ๆ กรียดการวิทุก การระบบสหรรายที่ เมื่อเกิด กรีบกรุงกรุงกระบายที่

CONTENTS

Preface	VII
CI I D I	3 8 6 9
Chapter 1, Background	· verself to 97 mile - 1
1.1. The material basis	is a limboli part of
1.1.1. Self-reproduction	A Company of the Company
	an emission are seen at legs. I
1.1.3. Replication	relative or a strong of sell 2
1.1.4. Transcription	5
	1 200 100 100 100 100 100 100 100 100 10
1.1.6. Gene, cistron, operon, factor, allele	William to the state of the sta
1.2. The organization of DNA. Chromosomes	
1.2.1. Viruses and bacteria	9
1.2.2. Fungi and yeasts	13
1.2.3. Higher organisms. Histones. Chromatin	15
1.2.4. Heterochromatin	16
1.2.5. Replicons	17
1.2.6. Semi-conservative replication	a the 18
1.2.7. Number of strands per chromosome	20
1.2.8. Repetitive DNA sequences	21
1.3. The division of the nucleus of higher organisms	22
1.3.1. Mitosis, karyokinesis	22
1.3.2. Duration of mitosis	27
1.4. Factors affecting nuclear division and chromosome	
and structure	assessment in the 29
1.4.1. Alterations in the course of mitosis	29
1.4.2. Colchicine	31
1.4.3. Polyploidy and polysomy	33
1.4.4. Terminology of chromosome-number variants	Carried English Line 1, 34
1.4.5. Chromosome-structural aberrations	La Francisco de la Carte de Ca
1.4.6. Ionizing radiations	36
1.4.7. Radiomimetic substance	* ReCord 1 2.85 41
1.4.8. Primary lesion. Types of aberrations	42
1.4.9. Dose –effect curves	47
1.4.10. Duration of treatment. Dose fractionation	48
1.4.11 Chromosome-structural variants	40

X Contents

1.5. The karyotype			52
1.5.1. Karyotype, Idio	gram. Karyogram		52
1.5.2. Constancy of the	e karyotype		63
1.5.3. Development of	the karyotype. Cytotaxonomy		64
Chapter 2, Recombin	nation		69
, , , , , , , , , , , , , , , , , , ,			
2.1. The exchange of DNA			69
2.1.1. Why recombinat			69
	of DNA exchange. Crossing-over		70
2.2. Crossing-over in prokar	ryotes		72
2.2.1. Viruses			72
2.2.2. Bacteria			74
2.3. Recombination in euka	aryotes		75
2.3.1. Fungi. Yeasts			75
2.3.2. Alternation of g			76
2.4. Reduction division. Me			80
2.4.1. Function and de			80
2.4.2. Duration of mei	osis		88
2.4.3. The two types o	f recombination		89
2.5. Segregation on the bas	is of chromosome recombination		89
2.5.1. Monofactorial se			89
2.5.2. Segregations inv	volving more than one factor w	rith chromosome	
recombination		No. of the last	95
2.6. Segregations involving	crossing-over		101
2.6.1. Chiasmata and c	rossing-over in eukaryotes	managed by 1970 1	101
2.6.2. The mechanism			105
2.6.3. Reciprocity of e	xchange. Conversion		111
2.6.4. Terminology: ex	change, chiasma, crossing-over, rec	om bination, link-	
age			112
2.7. Estimating recombinat	ion and crossing-over percentages		114
2.7.1. Test-cross			114
2.7.2. F ₂			115
2.7.3. Double crossing-	over. Maximum crossing-over perc	entage	117
2.8. Interference			119
2.8.1. Chromatid inter	ference. Chiasma interference		119
2.8.2. Coincidence			123
2.8.3. Negative interfer			123
2.8.4. Interchromosom	ne effects		123
2.8.5. Chiasma localiza	tion		124
2.9. Factors affecting chias			125
2.9.1. Interference and	chromosome pairing		125
2.9.2. Temperature			126
2.9.3. Age			127
2.9.4. Water content			128
2.9.5. Sex			128
2.9.6. Heterochromatin	n .		128

Contents	XI

2.9.7. Ionizing radiation	7177 4 7 14 129
2.9.8. Chemicals	129
2.9.9. Variation within the organism	130
2.9.10. Genetic factors	130
2.9.11. Shifts in populations	132
2.10. Uniformity and variation	133
2.10.1. The one-dimensional structure of the chromos	ome 133
2.10.2. Three moments of central importance	133
Chapter 3, Specialized chromosomes	135
Chapter of Specialized enfolicement	hitaly cutok
3.1. Adaptational forms of normal chromosomes	135
3.1.1. Polytene giant chromosomes	135
3.1.2. Lampbrush chromosomes	140
3.2. Permanently specialized chromosomes	143
3.2.1. Sex chromosomes. Genetic sex determination	143
3.2.2. X-chromosome neterochromatinization. Sex chr	omatin, Barr-body,
Drumsticks	149
3.2.3. Hemizygosity	151
3.2.4. Deviating segregations of genes in sex chromosom	
	morni plana and Lide 153
3.2.6. Nucleolar chromosomes	155
3.2.7. B-chromosomes	159
3.2.8. Holokinetic chromosomes	162
	n
Chapter 4, Chromosome structural variants	165
TPL.	10 90 00
4.1. Introduction	165
	165
4.2.1. Types	165
4.2.2. Phenotypic effect	166
4.2.3. Chromosome pairing. Recombination. Genetic co	
4.3. Duplications	170
4.3.1. Types	170
4.3.2. Phenotypic effect	17.0
4.3.3. Chromosome pairing. Recombination. Secondary	
4.4. Inversions	174
4.4.1. Types	174
4.4.2. Phenotypic effect. Position effect	174
4.4.3. Chromosome pairing	at a moral dated 177
4.4.4. Recombination. Consequences of exchange	An Square 177
4.4.5. Transmission	181
4.4.6. Inversions in natural populations	183
4.5. Translocations	185
4.5.1. Types	185
4.5.2. Phenotypic effect	186
4.5.3. Chromosome pairing, Exchange, Configurations	187

XII Contents

	4.5.4. Two and more translocations			193
	4.5.5. Orientation			195
	4.5.6. Semisterility			199
	4.5.7. Translocations in man			201
	4.5.8. Genetic consequences of translocati	ons		203
	4.5.9. Translocations in natural population			207
	4.5.10. Interchromosome effects. Position			212
	4.5.10. Interentomosome effects, rosition	chicci		212
01				212
Ch	apter 5, Numerical variants			213
- >	Assembleday			213
£ 1	Aneuploidy			213
3.1.	Primary trisomics			
	5.1.1. Origin and phenotypic effects			213
	5.1.2. Chromosome pairing. Crossing-over			218
	5.1.3. Orientation. Centromere mis-divisio	n		221
347	5.1.4. Transmission			223
	5.1.5. Genetic segregations			224
	5.1.6. Double reduction		2 1 1111	226
5.2.	Derived types		April 1 mars 1	228
	5.2.1. Secondary trisomics			228
	5.2.2. Telocentric trisomics			230
5.3.	Tertiary and interchange trisomics			233
	5.3.1. Types and origin			233
	5.3.2. Chromosome pairing			234
	5.3.3. Transmission. Genetic consequences	3		235
	Euploidy			236
5.4.	Haploids			236
	5.4.1. Origin			236
	5.4.2. Chromosome pairing. Genetic conse	quences		238
5.5.	Triploids			240
	5.5.1. Origin			240
	5.5.2. Chromosome pairing. Chiasmata			241
	5.5.3. Anaphase I distribution			242
5.6	Higher polyploids		*	245
0.0.	5.6.1. Autopolyploids. Origin and phenoty	nic effects		245
	5.6.2. Chromosome pairing. Chiasmata. M		tions Fertility	247
	5.6.3. Natural autopolyploids	delotte configura	itions. Termity	250
	5.6.4. Genetic consequences. Tetrasomy			252
	5.6.5. Crossing-over percentages			256
57	5.6.6. Inbreeding			258
5.7.	Allopolyploids	The same of the sa		260
	5.7.1. Types			260
	5.7.2. Diploidization			263
	5.7.3. Genetic regulation of chromosome			263
	5.7.4. Allopolyploidization of autopolyplo	oids		265
	5.7.5. Genome analysis			267
	5.7.6. Genetic consequences. Deviating seg	regations		271

	Contents	XIII
577 A	euploids in allopolyploids	271
	entification of homoeologues	273
	romosome addition	275
	hromosome substitution	276
	ene localization	279
	ntroduction of genes from other species	280
Chapter 6	Genetic chromosome maps	282
6.1. Gene ma	28	282
	ree point test	282
	pping functions	282
	ra-gene maps	284
6.1.4. Vi		285
6.1.5. Ba		287
	thromosome maps in prokaryotes with a single chromosome	290
6.2.1. Vi	The state of the s	290
6.2.2. Ba		291
AT CONTRACTOR OF THE PARTY OF T	elation of genes with chromosomes in eukaryotes	292
	romosomes and linkage groups	292
	ploidization	292
	ne-chromosome correlation from abnormal segregations	295
	ization of genes within chromosomes	296
	ntromere mapping	• 296
	her cytological markers	300
	netic chromosome maps from cytological data	305
Chapter 7	Deviant cytogenetic systems ,	307
7.1. Apomixi		307
7.1.1. Pla		307
7.1.2. Au	imals	309
7.2. Unusual	segregations	310
	ocentric activity	310
	iotic drive. Affinity. Distributive pairing	313
7.3. Other sy	tems	315
7.3.1. Ch	romosome elimination	315
7.3.2. So	matic segregation	316
7.3.3. Ep	isomes	318
7.4. Variation	in chromosome mechanisms	319
References		321
Index		334

Chapter 1

BACKGROUND

1.1. The material basis

1.1.1. Self-reproduction

An essential characteristic of living material is its capacity for self-reproduction. In its absence, life, once originated, would not escape extinction: there are numerous internal and external processes that carry out an efficient break-down. This self-reproduction must be extremely exact as the vital processes of even the simplest forms of life are so complicated that slight deviations from an established pattern, proven to be efficient, will almost certainly result in a reduction of fitness.

All living material, from virus to complex multicellular organism contains a single basic substance that combines two essential functions: (1) exact replication (self-reproduction) and (2) strict regulation of the vital processes. The two functions are carried out in sequence, not simultaneously. This basic substance is usually DNA (deoxyribonucleic acid). In some cases (plant viruses for instance) it is the related substance RNA (ribonucleic acid). The exact replication of the basic regulating substance is the root of the self-reproduction of the species.

1.1.2. DNA, RNA

DNA and RNA consist of long molecular chains of *nucleotides*, which are the mono-phosphate esters of *nucleosides*. Nucleosides contain a pentose sugar (deoxyribose in DNA and ribose in RNA) and one of four nitrogenous bases: adenine, guanine (both purines), thymine (only in DNA and substituted by uracil in RNA)

Fig. 1.1. The components of DNA and RNA. DNA contains the pyrimidines cytosine and thy midine, the purines adenine and guanine and the pentose sugar deoxyribose. RNA contains the pyrimidines cytosine and uracil, the purines adenine and guanine and the pentose sugar ribose.

and cytosine (all three pyrimidines) (see fig.1.1). Thus only four types of nucleosides occur: adenosine, guanosine, thymidine and cytidine in DNA; adenosine, guanosine, uridine and cytidine in RNA. As a consequence, DNA and RNA contain only four types of nucleotide. DNA and RNA principally differ only in respect to the pentose sugar and one of the four nitrogenous bases.

1.1.3. Replication

Polymerization of the nucleotides into the DNA (or RNA) chains occurs serially. The nucleotide to be built in is in triphosphate form, with its phosphate groups attached to the 5'C atom of the sugar. In the (enzymatic) process of attachment two of the three phosphate groups are removed and the remaining one binds to the 3'C atom of the previously attached nucleotide of the growing chain (fig.1.2). Thus the backbone of the DNA (and RNA) macromolecules is formed by successive (deoxy)ribose and phosphate groups, with the nitrogenous bases sticking out from the sides. The chain is not symmetrical: one end is the 3'C atom of the sugar, the other end the phosphate group at the 5'C atom: a

Fig.1.2. A fragment of a DNA double chain. The backbones of the two chains are formed by alternating sugar and phosphate groups. The polarity of the two chains is opposite. The nitrogenous bases are attached to the sugars and the complementary bases of the two chains are connected by hydrogen bridges. There are two between thymidine and adenine and three between cytosine and guanine. The double chain has the form of a helix with one revolution per 10 nucleotides.



Fig.1.3. Semi-conservative replication of the DNA double helix. After unwinding, one of the two original chains directly forms a new, complementary chain. The other first forms fragments (Okazaki fragments) in the opposite direction, i.e. the same direction in respect to polarity. The fragments are later linked together by ligases. The two daughter helices both contain one old and one new strand and are completely equivalent.

DNA (and RNA) molecule has a definite *polarity* which has important consequences.

When DNA re(du)plicates, the new chain is laid down alongside an old chain, which functions as a template. There is a positive—negative relation between the two chains, in the sense that the nucleotides of the new chain are not identical to those of the old chain, but of specific complementary types. Opposite to thymine in the old chain, adenine is built into the new chain and vice versa.

Opposite to cytosine comes guanine (and vice versa). For stereochemical reasons other combinations are excluded. Hydrogen bonds form between the bases of the old and the new chains: two between thymine and adenine and three between cytosine and guanine. The orientation of the new chain is such that its polarity is reversed compared to that of the old chain. The two chains normally remain together: DNA is double stranded (fig.1.2). Before new synthesis can take place, the two strands must be separated. It might be expected that both strands are synthesized continuously starting from one initiation point. Then, because of the opposite polarity of the two strands, synthesis in respect to polarity would be into one direction in one strand and into the other direction in the other strand. There are strong indications that this is not the case: one strand is synthesized continuously, but the other strand is synthesized in sections (Okazaki fragments) in the same direction in respect to polarity, i.e. into the opposite direction when considered from the initiation point. The fragments are subsequently (enzymatically) linked together by ligases (fig. 1.3).

Some important aspects of the structure of DNA are:

- (1) DNA normally is double stranded; when the base of the base
- (2) The two strands (the old and the new one) are not identical but complementary;
- (3) The two strands have opposite polarity;
- (4) In the total DNA there are equal quantities of adenine and thymine, and of guanine and cytidine, on a molecular basis.

 This is independent of the sequence or of the relative frequencies of nucleotides in each chain.

1.1.4. Transcription was and advantages with abundance and

In a comparable fashion but under different conditions and effected by different enzymes the DNA functions as a template for the formation of chains of RNA (transcription), After completion these RNA chains are released from the DNA without forming H-bonds. The RNA macro-molecule formed on the DNA is called *messenger* RNA (mRNA). It is carried to the cytoplasm where it is used as a template for the formation of polypeptide chains. This polypeptide synthesis is carried out by small bodies,

the ribosomes, that usually act in groups (polysomes), moving along the mRNA molecules in the process of polypeptide formation. The RNA moiety of the ribosomes (rRNA) consists of at least one small and two large RNA molecules that are transcribed on specific segments of the DNA. In the ribosome they are combined with proteins. The ribosomes pick up a third type of RNA: the transfer RNA (tRNA), that consists of much shorter chains, folded in a specific manner and often containing special base-types. These tRNAs are formed on their own DNA segments. The tRNA molecules have two recognition sites: on one an amino acid is attached enzymatically by a specific synthetase. The other site consists of a group of three nucleotides. Type and sequence of these nucleotides appear to be specific for the amino acid attached to the other site. Thus, for each amino acid there exists (at least) one specific tRNA characterized by a distinct group of three nucleotides. After having picked up a tRNA molecule with an amino acid, the ribosome first attaches the tRNA to the mRNA: the three specific tRNA nucleotides are fitted on a complementary set of three nucleotides in the mRNA. The first is placed on a specific initial site and the second follows on the next three nucleotides of the mRNA. The ribosome must select the correct tRNA for each site. For fitting the tRNA nucleotide triplets onto the mRNA triplets the same rules are followed as with transcription: adenine on uracil (which in RNA takes the place of thymine in DNA) etc. The row of amino acids is threaded together to a polypeptide chain and then the tRNA molecules are released from the mRNA and from the amino acids. Now both RNA forms are available for a new cycle, or are broken down. The sequence of amino acids in the polypeptide thus depends on the nucleotide sequence in the RNA, and consequently on that in the DNA, three nucleotides corresponding to one amino acid. This triplet of nucleotides in the mRNA contains the code for a specific amino acid and is named codon. Some codons have specific functions, such as interpunction. One amino acid may correspond to more than one codon: the code is degenerated. The codons UAA, UAG and UGA indicate the end of the polypeptide chain, the codons AUG and GUG the beginning.

For the replication of DNA both strands are used. For transcription only one is available: this prevents that two different polypeptide chains are derived from one DNA double strand. Which one is read off is determined by specific base sequences.

1.1.5. Mutation

Some polypeptide chains are built into structural proteins, but others are used to make enzymes. Small changes in the DNA, resulting from loss or from doubling of single nucleotides, can alter the entire pattern of transcription, since always three consecutive nucleotides form one codon. When the reading frame is shifted, all following codons are changed. A polypeptide may be formed, but it may not be expected to be functional as part of an enzyme. When no alternative intact DNA chain is available, a functional enzyme is not formed and the character conditioned by this enzyme is not expressed. Such a heritable change in a character is called a mutation, in this case a "frame shift" mutation.

Other changes in the DNA only locally affecting the reading frame such as inversion of a small segment or translocation from the original site to another location very nearby, or the replacement of one nucleotide by another, all will yield minor effects if the corresponding polypeptide segment is of minor importance. Whenever amino acid sequences in corresponding polypeptides have been studied in related species, small variations without great consequences have been detected. Occasionally, however, even simple amino acid substitutions, the consequence of single nucleotide replacements, have been found to have drastic effects. Both, harmless variations and drastic changes due to single amino acid substitution (sickle-cell anaemia for instance) have been found in haemoglobin and other components of the blood of man.

All such DNA changes (mutations) are transmitted with each cycle of DNA replication: they are heritable. In nature, mutations occur "spontaneously" (cause unknown) in a low frequency. They can be induced experimentally in high frequencies by chemicals (such as ethyl-methane-sulphonate, EMS, and other alkylating agents), by ionizing radiations, and by ultraviolet radiation. A high spontaneous mutation frequency would thoroughly disturb the