# BIOLOGICAL ROLES OF PROTEIN PHOSPHORYLATION

PROCEEDINGS OF
A ROYAL SOCIETY DISCUSSION MEETING
HELD ON 8 AND 9 DECEMBER 1982

ORGANIZED AND EDITED BY S. V. PERRY, F.R.S., AND P. COHEN

LONDON
THE ROYAL SOCIETY
1983

# BIOLOGICAL ROLES OF PROTEIN PHOSPHORYLATION

PROCEEDINGS OF
A ROYAL SOCIETY DISCUSSION MEETING
HELD ON 8 AND 9 DECEMBER 1982

ORGANIZED AND EDITED BY S. V. PERRY, F.R.S., AND P. COHEN

LONDON
THE ROYAL SOCIETY

1983

# Printed in Great Britain for the Royal Society at the University Press, Cambridge

ISBN 0 85403 212 6

First published in *Philosophical Transactions of the Royal Society of London*, series B, volume 302 (no. 1108), pages 1-166

OSPHORYLATIO

### Copyright

© 1983 The Royal Society and the authors of individual papers.

It is the policy of the Royal Society not to charge any royalty for the production of a single copy of any one article made for private study or research. Requests for the copying or reprinting of any article for any other purpose should be sent to the Royal Society.

S.V. PERRY FRS. AND P. CC

Published by the Royal Society 6 Carlton House Terrace, London SW1Y 5AG

rondon

# CONTENTS [Two plates]

resussall L

PIC Vanne Debres of Freedom nongrational Contract Strainers in	PAGE
E. G. Krebs  Historical perspectives on protein phosphorylation and a classification system for protein kinases	3
HORMONE ACTION AND PHOSPHORYLATION	6
P. Cohen and the state of the s	000
Protein phosphorylation and the control of glycogen metabolism in skeletal muscle	13
Discussion: T. GOLDSTONE	25
HG. HERS	
The control of glycolysis and gluconeogenesis by protein phosphorylation	27
Discussion: P. Cohen	32
R. M. DENTON AND R. W. BROWNSEY	
The role of phosphorylation in the regulation of fatty acid synthesis by insulin and	
other hormones	33
Discussion: S. J. YEAMAN	44
P. J. RANDLE  Mitochondrial 2-oxoacid dehydrogenase complexes of animal tissues	47
CELL MOTILITY AND PHOSPHORYLATION	ARS S
S. V. Perry, F.R.S. and to the manner of the world with the annual residence of the state of the	onS.
Phosphorylation of the myofibrillar proteins and the regulation of contractile	
activity in muscle	59
R. C. Smith, W. Z. Cande, R. Craig, P. J. Tooth, J. M. Scholey and J. Kendrick-Jones	
Regulation of myosin filament assembly by light-chain phosphorylation	73
Discussion: N. Crawford	81
P. J. England	
Cardiac function and phosphorylation of contractile proteins	83
J. HAIECH AND J. G. DEMAILLE	
Phosphorylation and the control of calcium fluxes	91
GENERAL DISCUSSION	
R. J. P. WILLIAMS, F.R.S.	99

.

S.M. 1 YEARS 157

Kungaios-fonss

LINERAL DESCRIPTION.

Bigging N. CRAWFORD

Harren Ave J. G. DERMILLE

R. J. R. WILLIAMS, F.R. S. W. C.

Phosphorplation and the control of eviction there

MEMBRANE FUNCTION AND PROTEIN PHOSPHORYLATION	
Y. Nishizuka  Calcium, phospholipid turnover and transmembrane signalling	10!
J. Bennett  Regulation of photosynthesis by chloroplast protein phosphorylation  Discussion: P. Cohen, D. E. Hanke, L. Pinna	113
distorted perspectives on process phosphorelation and a classification system for	
PHOSPHORYLATION AND GENE EXPRESSION  T. Hunt Phosphorylation and the control of protein synthesis	127
KATHLEEN M. Rose, D. A. STETLER AND S. T. JACOB  Phosphorylation of RNA polymerases: specific association of protein kinase NII with RNA polymerase I  Discussion: L. A. Pinna	135
R A. LASKEY  Phosphorylation of nuclear proteins  Phosphorylation of nuclear proteins	143
PROTEIN PHOSPHORYLATION AND CANCER	Ear
R. L. ERIKSON, TONA M. GILMER, ELEANOR ERIKSON AND J. G. FOULKES Avian sarcoma viruses, protein kinases and cell transformation	151
BIOLOGICAL ROLE OF PROTEIN PHOSPHORYLATION	

E. SHACTER-NOIMAN, P. B. GHOCK AND E. R. STADTMAN

Physiological in a tipe a compatible province and the regulation of contraction

R. Cashitzis, W. Z. Candr, R. Craig, P. J. Tootis, J. M. Scholey And

Regulation of myosin filament secondly by high chain place baylarion

Protein phosphorylation as a regulatory device

# CONTENTS [Two plates]

Transmati A

	PAGE
E. G. Krebs  Historical perspectives on protein phosphorylation and a classification system for protein kinases	3
Tartes of Liphon a consequential consults they will be backed use placehood by	BET
HORMONE ACTION AND PHOSPHORYLATION	is the
P. COHEN THE MARKET AND IS IN THE PROPERTY OF	00
Protein phosphorylation and the control of glycogen metabolism in skeletal muscle	13
Discussion: T. GOLDSTONE	25
HG. HERS	
The control of glycolysis and gluconeogenesis by protein phosphorylation	. 27
Discussion: P. Cohen	32
[1] 하고 있다 이 경영 보고 그리고 있으면 하는 것이 되었다. 하는 사람들이 하는 사람들이 되었다. 그 그 사람들이 없는 것이 없는데 하는데 없는데 없는데 없다.	
R. M. DENTON AND R. W. BROWNSEY	
The role of phosphorylation in the regulation of fatty acid synthesis by insulin and other hormones	33
Discussion: S. J. YEAMAN	44
。 http://www.htm.com/schip.com/schi	A STATE
P. J. RANDLE  Mitochondrial 2-oxoacid dehydrogenase complexes of animal tissues	47
CELL MOTILITY AND PHOSPHORYLATION	
S. V. Perry, F.R.S.	025
Phosphorylation of the myofibrillar proteins and the regulation of contractile	
activity in muscle	59
R. C. Smith, W. Z. Cande, R. Craig, P. J. Tooth, J. M. Scholey and J. Kendrick-Jones	
Regulation of myosin filament assembly by light-chain phosphorylation	73
Discussion: N. Crawford	81
P. J. England  Cardiac function and phosphorylation of contractile proteins	83
J. HAIECH AND J. G. DEMAILLE	
Phosphorylation and the control of calcium fluxes	91
GENERAL DISCUSSION	
R. J. P. WILLIAMS, F.R.S.	99

## MEMBRANE FUNCTION AND PROTEIN PHOSPHORYLATION Y. NISHIZUKA Calcium, phospholipid turnover and transmembrane signalling 10! J. BENNETT Regulation of photosynthesis by chloroplast protein phosphorylation 113 Discussion: P. Cohen, D. E. Hanke, L. Pinna 124 Historical perspectives on protein phosphoralation and a classification system for PHOSPHORYLATION AND GENE EXPRESSION T. HUNT Phosphorylation and the control of protein synthesis 127 KATHLEEN M. ROSE, D. A. STETLER AND S. T. JACOB Phosphorylation of RNA polymerases: specific association of protein kinase NII with RNA polymerase I 135 Discussion: L. A. PINNA 142 The council of glycolysis and gluconcogenesis by protein objections of the R. A. LASKEY Phosphorylation of nuclear proteins R. M. Derrenium and Mill Recommended to the rest of the Research PROTEIN PHOSPHORYLATION AND CANCER

R. L. ERIKSON, TONA M. GILMER, ELEANOR ERIKSON AND J. G. FOULKES	
Avian sarcoma viruses, protein kinases and cell transformation	151

### BIOLOGICAL ROLE OF PROTEIN PHOSPHORYLATION

	DITT TO L.
E. SHACTER-NOIMAN, P. B. GHOCK AND E. R. STADTMAN	LIBO
Protein phosphorylation as a regulatory device	2.519 Vagag 157

Phosphoryish on of the myorbidian propries and the regulation of contractile

R. C. SMILL, W.Z. CARDE, R. CRAIG, P. J. TONTH, J. M. SCHOLEY-KER

HAVE OF BUILDING

Phosphorylation and the control of coloinin fluxe

# Historical perspectives on protein phosphorylation and a classification system for protein kinases

### By E. G. KREBS

Howard Hughes Medical Institute Laboratory, Department of Pharmacology, University of Washington, Seattle, Washington 98195, U.S.A.

Work on the phosphorylation of proteins as a dynamic process involved in the regulation of biological processes started in the 1950s with the finding that phosphorylase a and phosphorylase b are phospho and dephospho forms of the same enzyme. The field expanded sharply in the late 1960s with the discovery of the cyclic-AMP-dependent protein kinase, and it is now clear that phosphorylation-dephosphorylation constitutes a major type of regulation almost as common as allosteric control. The protein kinases, which catalyse the phosphorylation step in various phosphorylationdephosphorylation systems, can be divided into two main classes, the serine-threonine protein kinases and the tyrosine protein kinases. Each class can be subdivided into groups or entities depending on the nature of the agent(s) that regulate activity.

### INTRODUCTION

It is now well recognized that the phosphorylation and dephosphorylation of proteins is an important regulatory process applicable to many different cellular functions. The holding of this meeting and the organization of numerous other international symposia on protein phosphorylation during the past several years attest to this fact. How did a general appreciation of protein phosphorylation as a control mechanism develop? What is our present understanding of the diversity and properties of the enzymes that catalyse protein phosphorylation and dephosphorylation? Are there sets of principles that can serve as guidelines to investigators probing the possible role of protein phosphorylation in different systems? What is the future of the field? These and other general questions will constitute the theme of this paper, which is meant to serve as an introduction to this meeting.

# HISTORICAL ASPECTS

# Early work on glycogen metabolism

Before the 1950s the only known phosphoproteins were casein, phosvitin and related proteins involved in the feeding of the young or in early embryonic development. One exception was the proteolytic enzyme pepsin, which was known to contain a stoichiometric amount of phosphate (Northrop 1930). (In a sense it is ironical that the first phosphoenzyme should be one in which there is no known function of the bound phosphate, i.e. pepsin is an example of 'silent phosphorylation'.) The first developments that foreshadowed current concepts of protein phosphorylation as a dynamic process involved in regulation grew out of work on glycogen metabolism.

In the early 1940s Carl and Gerti Cori together with Arda Green had shown that the enzyme phosphorylase, which catalyses the first step in glycogen breakdown, exists in active and inactive forms (Cori & Green 1943; Cori & Cori 1945). They named these forms phosphorylase a and phosphorylase b and succeeded in demonstrating that phosphorylase a could be converted

to phosphorylase b in vitro. They were unable to achieve conversion of phosphorylase b to a in vitro but did obtain evidence that this reaction occurred in vivo. Thus the first 'interconvertible enzyme' system was established. The chemical nature of the interconversion reactions involving the forms of phosphorylase was not then appreciated.

In the early 1950s Earl Sutherland and his collaborators (Wosilait & Sutherland 1956; Rall et al. 1956), working with liver phosphorylase, and E. H. Fischer and myself (Fischer & Krebs 1955; Krebs & Fisher 1956), working with the muscle enzyme, discovered that the active form of phosphorylase is a phosphoenzyme and that the interconversion reactions involve enzyme-catalysed phosphorylation and dephosphorylation steps. The enzyme catalysing the phosphorylation of phosphorylase by ATP was named phosphorylase kinase, and the enzyme catalysing the hydrolytic dephosphorylation step was named phosphorylase phosphatase. These reactions, incorporating our present knowledge regarding stoichiometry, are as follows:

phosphorylase 
$$b+2$$
 ATP  $\xrightarrow{\text{kinase}}$  phosphorylase  $a+2$  ADP; (1)

phosphorylase phosphorylase 
$$a + 2 \text{ H}_2\text{O} \xrightarrow{\text{phosphatase}} \text{phosphorylase } b + 2 \text{ P}_i.$$
 (2)

Shortly after the elucidation of the biochemical mechanism of the phosphorylase interconversion reactions it was found that phosphorylase kinase is itself activated under phosphorylating conditions in a process that is stimulated by cyclic AMP (Krebs et al. 1959, 1964). Then in 1963 it was reported that glycogen synthase exists in two forms that are interconverted by phosphorylation and dephosphorylation (Friedman & Larner 1963). The phosphorylation of glycogen synthase, like that of phosphorylase kinase, was shown to be stimulated by cyclic AMP (Belocopitow 1961; Huijing & Larner 1966). It was of particular interest that, whereas phosphorylation had been shown to increase the activity of phosphorylase and phosphorylase kinase, both of which are involved in glycogen breakdown, phosphorylation decreased the activity of the biosynthetic enzyme, glycogen synthase. This reciprocal relation has been found to hold for other enzymes involved in biosynthesis and degradation and it would appear to be one of the 'principles' that has been established for the regulation of enzymes by phosphorylation—dephosphorylation. Philip Cohen, one of the organizers of this meeting, has effectively emphasized this point in recent years.

The mechanism of action of cyclic AMP in the regulation of phosphorylase kinase activation, and glycogen synthase inactivation, both resulting from enzyme phosphorylation, was not entirely clear at first. In connection with phosphorylase kinase, it was apparent that cyclic AMP enhanced the rate of phosphorylation and activation of the enzyme, but it was not known whether it interacted with phosphorylase kinase itself to stimulate an autophosphorylation reaction or whether a second protein kinase, i.e. a cyclic-AMP-dependent phosphorylase kinase kinase, was involved (Krebs et al. 1965). In connection with glycogen synthase, cyclic AMP also enhanced the rate of phosphorylation of the enzyme, and in this case, in that the substrate in the phosphorylation reaction is not a protein kinase, it was reasonably safe to conclude that the target for the regulatory agent was the synthase kinase itself (Huijing & Larner 1966 a, b). A less likely possibility was that the synthase might have contained two interacting kinases involved in a cascade. These uncertainties were resolved when it was determined that a separate phosphorylase kinase existed (Walsh et al. 1968) and when later it was shown that this enzyme and glycogen synthase kinase are the same enzyme (Schlender et al. 1969); Soderling et al. 1970).

The cyclic-AMP-dependent phosphorylase kinase kinase was not referred to by that name, but was instead called a cyclic-AMP-dependent protein kinase, since it was realized immediately that it could catalyse the phosphorylation of more than one protein (Walsh et al. 1968).

Because of widespread interest in cyclic AMP and its multiple functions, the finding of a cyclic-AMP-dependent protein kinase served to focus attention on the process of regulation by protein phosphorylation—dephosphorylation. Before this development many investigators had thought of the process as probably being limited to the control of glycogen metabolism, but it now appeared that it might have widespread implications. Noting an almost ubiquitous distribution of the cyclic-AMP-dependent protein kinase in various cell types, Kuo & Greengard (1969) postulated that all actions of cyclic AMP might be mediated by this enzyme. (This hypothesis is still valid for eukaryotic cells but does not hold for prokaryotes.) Another important finding that served to broaden interest in protein phosphorylation was the discovery by Lester Reed and his collaborators that pyruvate dehydrogenase is regulated by phosphorylation—dephosphorylation (Linn et al. 1969). Here was a clearcut example of a phospho—dephospho enzyme system well beyond the confines of glycogen metabolism.

### Present status of protein phosphorylation-dephosphorylation

When investigators became aware of the 'new' type of metabolic regulation and broadened their horizons to look beyond the confines of glycogen metabolism, work on the phosphorylation and dephosphorylation of enzymes expanded rapidly. This is illustrated in figure 1, in which the total number of enzymes known to undergo phosphorylation—dephosphorylation, or to contain covalently bound phosphate as isolated, is shown as a function of time. By early 1982 more than 40 enzymes belonged in this category. Table 1 lists the individual enzymes and literature citations for each entry.

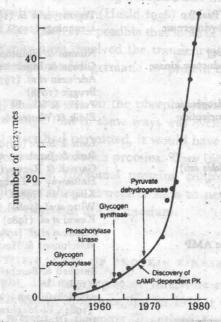


FIGURE 1. Enzymes reported to undergo phosphorylation-dephosphorylation.

Abbreviations: cAMP, cyclic AMP; PK, protein kinase.

seem technologica

### TABLE 1. ENZYMES REPORTED TO UNDERGO PHOSPHORYLATION-DEPHOSPHORYLATION

enzyme

glycogen phosphorylase

phosphorylase kinase† glycogen synthase† hormone-sensitive lipase†

fructose 1,6-bisphosphatase†
pyruvate dehydrogenase
hydroxymethylglutaryl-CoA reductase†
myosin light chain
acetyl-CoA carboxylase†
casein kinase II
RNA polymerase I†

### RNA polymerase II†

pyruvate kinase (liver)† cholesterol ester hydrolase† type II cyclic-AMP-dependent protein kinase regulatory subunit† tyrosine aminotransferase phosphofructokinase 1† tyrosine hydroxylase† phenylalanine hydroxylase† haem-inhibited elF-2 kinase cyclic-AMP-dependent protein kinase catalytic double-stranded RNA-dependent protein kinase cyclic-GMP-dependent protein kinase tryptophan hydroxylase type I cyclic-AMP-dependent protein kinase regulatory subunit SV40 large T antigen (an 'ATPase') NAD-dependent glutamate dehydrogenase (yeast) glycerophosphate acyltransferase† hydroxymethylglutaryl-CoA reductase kinase myosin light chain kinase† pp60ere† branched-chain ketoacid dehydrogenase calmodulin-dependent cyclic nucleotide phosphodiesterase† ATP citrate lyase† poly(A) polymerase isocitric dehydrogenase (Escherichia coli) casein kinase I EGF receptor-associated kinase P120gag-abl P90848-yes P140sag-fps insulin-stimulated low Km cyclic AMP phosphodiesterase† glucose-6-phosphatase†

### phosphofructokinase 2†

glycogen synthase kinase 3 (phosphatase activator, F<sub>A</sub>) ornithine decarboxylase (slime mould) Ca<sup>2+</sup>-transport ATPase cholesterol 7α-hydroxylase

### references

Fischer & Krebs (1955), Sutherland & Wosilait (1955), Rall et al. (1957) Krebs (1959) Friedman & Larner (1963) Rizak (1964), Corbin et al. (1970), Huttunen et al. (1970) Mendicino et al. (1966), Rious et al. (1977) Linn et al. (1969) Beg et al. (1973) Perrie et al. (1973) Carlson & Kim (1973) Walinder (1973) Jungman et al. (1974), Martelo & Hirsch (1974)Jungman et al. (1974), Martelo & Hirsch (1974) Ljungström et al. (1974) Trzeciak & Boyd (1974) Erlichman et al. (1974)

Lee & Nickol (1974)
Brand & Söling (1975)
Morgenroth et al. (1975)
Milstein et al. (1976)
Farrell et al. (1977)
Bechtel et al. (1977)

### Farrell et al. (1977)

deJonge et al. (1977) Hammon et al. (1977, 1978) Steinberg et al. (1977)

Tegtmeyer et al. (1977) Hemmings (1978)

Nimmo & Houston (1978) Gibson & Ingebritsen (1978) Adelstein et al. (1978) Brugge (1978) Parker & Randle (1978) Eldik & Watterson (1979), Sharma et al. (1980)

Linn & Srere (1979)
Rose & Jacob (1979)
Garnak & Reeves (1979)
Hathaway & Traugh (1979)
King et al. (1980)
Witte et al. (1980)
Kawai et al. (1980)
Feldman et al. (1980)
Marchmont & Houslay (1980)

Burchell & Burchell (1980), Begley & Craft (1981) van Schaftingen et al. (1981), Furuya et al. (1982), El-Maghrabi et al. (1982) Hemmings et al. (1981)

Atmar & Kuehn (1981) Caroni & Carafoli (1971) Sanghri et al. (1981)

Several points in relation to figure 1 and table 1 warrant discussion. Based on what is currently known, only about half of the enzymes in table 1 meet any substantial number of the criteria (Krebs & Beavo 1979) that should be satisfied to establish that a given enzyme undergoes physiologically significant phosphorylation-dephosphorylation from a regulatory standpoint. For instance, many of the enzymes listed undergo 'silent' phosphorylation, a term that has been used by Cohen (1982) to describe the phosphorylation of enzymes at sites that do not influence enzyme activity; examples of silent phosphorylation include the phosphorylation of tyrosine amino-transferase, 6-phosphofructo 1-kinase and ATP-curate lyase as well as that of a number of the protein kinases. It is known that the last group, protein kinases, characteristically catalyse their own phosphorylation, and in most instances the 'autophosphorylation' reaction has no demonstrable effects on enzyme activity. Also, for several of the enzymes listed in table 1 it has not actually been demonstrated that phosphorylation and dephosphorylation occur in vivo. Finally, even for enzyme that meet many of the criteria of physiological relevance, careful work has not always been carried out to correlate the state of phosphorylation of the enzyme in vivo with a physiological event that is presumably being regulated by phosphorylation-dephosphorylation. It should also be noted that several of the entries in table 1 are based on preliminary reports and may not withstand the test of time.

The regulation of cellular function as a result of protein phosphorylation—dephosphorylation reactions involves many proteins in addition to those that have been identified as having enzymic activity. For example, even as the role of this process in the control of glycogen metabolism was unfolding, it was recognized that non-enzymic nuclear proteins undergo dynamic phosphorylation—dephosphorylation (Kleinsmith et al. 1966), and a 'specific' histone kmase was discovered as early as 1967 (Langan & Smith 1967). After the finding of a cyclic-AMP dependent protein kinase (Walsh et al. 1968), it was shown that this enzyme and the histone kinase are apparently identical (Langan 1968). In addition to early studies on the phosphorylation of non-enzymic nuclear proteins, pioneering work had also been carried out on the rapid turnover of protein-bound phosphate in brain (Heald 1958) and in cell membranes in relation to ion transport (Judah & McLean 1962). It is possible that some of the protein phosphorylation observed in these later studies may have involved the transient phosphoenzyme intermediate of the Na<sup>+</sup>-K<sup>+</sup>ATPase and would not be germane to protein kinase-catalysed phosphorylation.

During the 1970s the number of reports on the phosphorylation of non-enzymic proteins underwent a dramatic upward surge that in some ways was even more impressive than that which occurred for enzymes. If space had permitted, it would have been of interest to consider in detail some of the countless examples of such proteins. They include cytosolic proteins, cell membrane proteins, nuclear proteins, ribosomal proteins, contractile proteins, cytoskeletal proteins and proteins of highly specialized structures such as nerve endings. Some of these topics will be discussed in other papers in this symposium.

### CLASSIFICATION OF PROTEIN KINASES

Until recently it was thought that only serine or threonine residues could serve as the phosphate acceptors in protein kinase substrates, but in 1979 it was shown that an activity phosphorylating tyrosine was present in polyoma T antigen immunoprecipitates (Eckhard et al. 1979). Shortly thereafter it was found that pp60<sup>sro</sup>, the transforming protein of the Rous

sarcoma virus (Brugge & Erikson 1977), which was known to be associated with protein kinase activity (Collet & Erikson 1978), was a tyrosine protein kinase (Hunter & Sefton 1980; Collet et al. 1980). Tyrosine protein kinase activity was then found to be associated with the epidermal growth factor (EGF) receptor (Ushiro & Cohen 1980). In addition, it has been shown that PDGF, the platelet-derived growth factor (Ek et al. 1982; Ek & Heldin 1982; Nishimura et al. 1982) and insulin (Kasuga et al. 1982a, b) stimulate the phosphorylation of tyrosine in certain proteins.

It appears that tyrosine protein kinases can be treated as a separate class of specific enzymes distinct from serine—threonine protein kinases. In this connection, however, it is of interest that pp60src and the catalytic subunit of the cyclic-AMP-dependent protein kinase have been shown to be structurally related (Barker & Dayhoff 1982). In addition to serine—threonine and tyrosine protein kinases it is possible that a third class of protein kinases, enzymes manifesting activity toward histidine and lysine residues, may exist; however, work on these enzymes has been limited (Smith et al. 1974).

Many serine-threonine protein kinases (table 2) other than the cyclic-AMP-dependent protein kinase(s) are regulated by specific effectors, a property that has been useful in their classification (Krebs & Beavo 1979). So far, enzymes of this class have been described that are regulated by (1) cyclic AMP, (2) cyclic GMP, (3) Ca<sup>2+</sup>, (4) diacylglycerol, (5) double-stranded RNA, (6) haemin, (7) acetyl CoA, NADH, pyruvate and ADP, and (8) polyamines.

### Table 2. Subclassification of serine-threonine protein kinases (PKs)

regulatory agent(s)

cyclic AMP

cyclic GMP Ca<sup>2+</sup> (calmodulin)

diacylglycerol (Ca<sup>2+</sup>, phospholipid) double-stranded RNA haemin (inhibition) acetyl-CoA, NADH, pyruvate, ADP polyamines unknown specific entities

type I, type II (heart) and type II (brain) cyclic-AMP-dependent PKs cyclic-GMP-dependent PK phosphorylase kinase (glycogen synthase myosin light-chain kinase liver glycogen synthase kinase (Soderling) protein kinase C double-stranded RNA-dependent PK eIF, kinase pyruvate dehydrogenase kinase polyamine-dependent PK casein kinase 1 (NI, Ck-S) casein kinase 2 (NII, Ck-TS, glycogen synthase kinase 5) glycogen synthase kinase 3 (factor F<sub>A</sub>) glycogen synthase kinase 4

In addition to the cyclic-nucleotide-dependent enzymes, the protein kinases that are regulated by calcium are especially prominent. Phosphorylase kinase was the first enzyme that was found to be dependent on calcium ions for activity (Meyer et al. 1964; Ozawa et al. 1967), a property that can now be accounted for by the presence of calmodulin as one of the subunits of the enzyme (Cohen et al. 1978). In other members of this group, calmodulin also serves as the component that mediates the calcium effect, but in these enzymes calmodulin is not an integral subunit in the same sense that it is in phosphorylase kinase. It is of interest that the several calcium (calmodulin)-dependent protein kinases (table 2) have distinctly different catalytic subunits and different specificities but share a common regulatory subunit, whereas the different

cyclic-AMP-dependent protein kinases share a common catalytic subunit but have different regulatory subunits. In this meeting we shall be hearing about several of the calcium (calmodulin)-dependent protein kinases and will also be hearing about protein kinase C, which requires calcium but not calmodulin.

A major subset of serine-threonine protein kinases consists of those enzymes for which no regulatory agents or effectors are known to exist. This does not necessarily mean, however, that these enzymes, or the reactions that they catalyse, are not subject to regulation, because regulation could occur by any of several mechanisms other than through the direct interaction of the protein kinase with an effector molecule. For example, regulation could be effected at the level of the protein substrate(s), i.e. it could be brought about through alterations in substrate conformation caused by the binding of effectors. Also, protein kinases for which no known effectors exist might be regulated by covalent modification, e.g. by serving as substrates for other protein kinases, A possible example of this type would be hydroxymethylglutaryl-CoA reductase kinase (Gibson et al. 1978). Finally, a protein phosphorylation reaction catalysed by a non-regulated protein kinase might be effectively controlled through regulation of the phosphoprotein phosphatase acting on the phosphorylated substrate.

# Table 3. Subclassification of tyrosine protein kinases

regulatory agent(s) Egg A. M. & Jacob S. T. Mar EGF A CONT. 19812

PDGF insulin unknown

Mender, M. M., Wel, Shallon, N

Toler Sear, Chr. Inglischet Masson Line and

An ve specific entities A see A and A S first and

London J. D., Relevant, E. M., Walth,

I'v. B., Westerman, B., Warranger A.

EGF receptor A Tron M.O. month & M.M. candala (PDGF receptor)
(insulin receptor) pp60ere P140gag-fps and P105gag-fps

P140gag-tps and P105gag-tps
P90gag-yes and P80gag-yes P85gag-fes and P110gag-fes

Flocker, E. H. & Ercke, E. L. aggs-yndrel, Chan City Mills 122.7 Academic M. S. Herrett D. A. J. of Table 3 categorizes the tyrosine protein kinases by the same system that was used for the serine-threonine protein kinases. Thus one can speak of EGF-dependent, PDGF-dependent and insulin-dependent tyrosine protein kinases. For EGF-stimulated tyrosine phosphorylation there is good evidence that the growth factor receptor itself is the protein kinase (Buhrow et al. 1982), and this suggests that the PDGF and insulin receptors may also be protein kinases. Supporting this view is the finding that in each case these factors stimulate the phosphorylation of tyrosine in a membrane protein with the molecular mass of the receptor, and it is known (see above) that protein kinases characteristically catalyse an autophosphorylation reaction. The virus-associated tyrosine protein kinases are not known to be regulated by specific effectors and would be analogous to the comparable group of serine-threonine protein kinases. It is of interest in this connection that one of these, pp60src, is known to be phosphorylated by the cyclic-AMPdependent protein kinase (Collett et al. 1979), suggesting the possibility of regulation by covalent modification. Table 3 indicates the existence of five types of viral onc-gene related tyrosine protein kinases, which are designated by using a system recently agreed upon by a number of authors (Coffin et al. 1981). For each viral-encoded entity listed in table 3 a normal cellular counterpart would also exist, making a total of ten distinct, albeit related, enzymes in this group. 5, M. G. & Flacture, R. W. 1985 Birchim. hisphys. Acta 20, 149-167.

Krein, E. G., Connida, G., Perner, J. B., Love, D. S., Branold, G. E. & Eischer, E. H. 1964. In Old Foreland

Amsterna on Course of Garages Advantages, pp. 200-210. Landon: L & A. Churchill.

### and the liquid of plates no References

Adeletein, R. S., Conti, M. A., Hathaway, D. R. & Klu, C. B. 1978 J. biol. Chem. 253, 8347-8350.

Atmar, V. J. & Kuehn, G. D. 1981 Proc. natn. Acad. Sci. U.S.A. 78, 5518-5522.

Barker, W. C. & Dayhoff, M. O. 1982 Proc. natn. Acad. Sci. U.S.A. 79, 2836-2839.

Bechtel, P. J., Beavo, J. A. & Krebs, E. G. 1977 J. biol. Chem. 252, 2691-2697.

Beg, Z. H., Allmann, D. W. & Gibson, D. M. 1973 Biochem. biophys. Res. Commun. 54, 1362-1369.

Begley, P. J. & Craft, J. A. 1981 Biochem. biophys. Res. Commun. 103, 1029-1034.

Belocopitow, E. 1961 Archs Biochem. Biophys. 93, 457.

Brand, I. A. & Soling, H. E. 1975 FEBS Lett. 57, 163-168.

Brugge, J. S. & Erikson, R. L. 1977 Nature Lond. 268, 346-348.

Brugge, J., Erikson, E., Collett, M. S. & Erikson, R. C. 1978 J. Virol. 26, 773-782.

Buhrow, S. A., Cohen, S. & Staros, J. V. 1982 J. biol. Chem. 257, 4019-4022.

Burchell, A. & Burchell, B. 1980 FEBS Lett. 118, 180-184.

Carlson, C. A. & Kim, K. H. 1973 J. biol. Chem. 248, 378-380.

Caroni, P. & Carafoli, E. 1981 J. biol. Chem. 256, 9371-9373.

Coffin, J. M., Varmus, H. E., Bishop, J. M., Essex, M., Hardy, W. D., Martin, G. S., Rosenberg, N. E., Scolnick, E. M., Weinberg, R. A. & Vogt, P. K. 1981 J. Virol. 40, 953-957.

Cohen, F. 1982 Nature, Lond. 296, 613-620.

Cohen, P., Burchell, A., Foulkes, J. G., Cohen, P. T. W., Vanaman, T. C. & Nairn, A. C. 1978 FEBS Lett. 92, 287-293.

Collett, M. S., Erikson, E. & Erikson, R. L. 1979 J. Virol. 29, 770-781.

Collett, M. S., Purchio, A. F. & F. Ison, R. L. 1980 Nature, Lond. 285, 167-169.

Corbin, J. D., Reimann, E. M., Walsh, D. A. & Krebs, E. G. 1970 J. biol. Chem. 245, 4849-4851.

Cori. G. T. & Coii, C. F. 1945 J. biol. Chem. 158, 321-332. Cori, G. T. & Green, A. A. 1943 J. biol. Chem. 151, 31-38.

Damuni, Z. & Cohen, P. 1982 Eur. J. Biochem. 129, 57-65.

deJonge, H. R. & Rosen, O. M. 1977 J. biol. Chem. 252, 2780-2783.

Eckhart, W., Hutchinson, M. A. & Hunter, T. 1979 Cell 18, 925-933.

Fk, B. & Heldon, C.-H. 1982 J. biol. Chem. 257, 10486-10492.

Ek, B., Westermark, B., Wasteson, A. & Heldin, C.-H. 1982 Nature, Lond. 295, 419-420.

Fldik, L. J. V. & Watterson, D. M. 1979 Fedn Proc. Fedn Am. Socs exp. Biol. 38, 789.

El-Maghrabi, M. R., Claus, T. H., Pilkis, J., Fox, E. & Pilkis, S. J. 1982 J. biol. Chem. 257, 7603-7607.

Erlichman, J., Rosenfeld, R. & Rosen, O. M. 1974 J. biol. Chem. 249, 5000-5003.

Farrell, P. J., Balkow, K., Hunt, T. & Jackson, R. J. 1977 Cell 2, 187-200.

Feldmann, R. A. 1980 Cell 22, 757.

Fischer, E. H. & Krebs, E. G. 1955 J. biol. Chem. 216, 121-132.

Friedman, D. L. & Larner, J. 1963 Biochemistry, Wash. 2, 669-675.

Furuya, E., Yokoyoma, M. & Uyeda, K. 1982 Proc. natn. Acad. Sci. U.S.A. 79, 325-329.

Chrinak, M. & Reeves, H. C. 1979 Science, Wash. 203, 1111-1112. Gibson, D. M. & Ingebritson, T. S. 1978 Life Sci. 23, 2649-2664.

Hamon, M., Bourgoin, S., Artaud, F. & Hery, F. 1977 J. Neurochem. 28, 811-818.

Hamon, M., Bourgoin, S., Hery, F. & Simonnet, G. 1978 Molec. Pharmac. 14, 99-110.

Hathaway, G. M. & Traugh, J. A. 1979 J. biol. Chem. 254, 762-768.

Heald, P. J. 1959 Biochem. J. 73, 132-141.

Hemmings, B. A. 1978 J. biol. Chem. 253, 5255-5258.

Hemmings, B. A., Yellowlees, D., Kernohan, J. C. & Cohen, P. 1981 Eur. J. Biochem. 119, 443-451.

Huijing, F. & Larner, J. 1966a Biochem. biophys. Res. Commun. 23, 259.

Huijing, F. & Larner, J. 1966 b Proc. natn. Acad. Sci. U.S.A. 56, 647-653.

Hunter, T. & Sefton, B. M. 1980 Proc. natn. Acad. Sci. U.S.A. 77, 1311-1315.

Huttunen, J. K., Steinberg, D. & Mayer, S. E. 1970 Proc. natn. Acad. Sci. U.S.A. 67, 290-295.

Judah, J. D. & McLean, A. M. 1962 Biochem. Pharmac. 2, 593-602.

Jungmann, R. A., Hiestand, P. C. & Schweppe, J. S. 1974 J. biol. Chem. 249, 5444-5451.

Kasuga, M., Zick, Y., Blithe, D. L., Crettaz, M. & Kahn, C. R. 1982a Nature, Lond. 298, 667-669.

Kasuga, M., Zick, Y., Blithe, D. L., Karlsson, F. A., Haring, H. V. & Kahn, C. R. 1982 b J. biol. Chem. 257, 9891-9894.

Kawai, S., Yoshida, M., Segawa, K. & Sugiyama, H. 1980 Proc. natn. Acad. Sci. U.S.A. 77, 6199-6203.

King, L. E., Carpenter, G. & Cohen, S. 1980 Biochemistry, Wash, 19, 1524-1528.

Kleinsmith, L. J., Allfrey, V. G. & Mirsky, A. E. 1966 Proc. natn. Acad. Sci. U.S.A. 55, 1182-1189.

Krebs, E. G. & Beavo, J. A. 1979 A. Rev. Biochem. 48, 923-959.

Krebs, E. G. & Fischer, E. H. 1956 Biochim. biophys. Acta 20, 150-157.

Krebs, E. G., Gonzalaz, C., Posner, J. B., Love, D. S., Bratvold, G. E. & Fischer, E. H. 1964 In Ciba Foundation Symposium on Control of Glycogen Metabolism, pp. 200-210. London: J. & A. Churchill.

Krebs, E. G., Graves, D. J. & Fischer, E. H. 1959 J. biol. Chem. 234, 2867-2873.

Kuo, J. F. & Greengard, P. 1969 J. biol. Chem. 244, 3417-3419.

Langan, T. A. 1968 Science, Wash. 162, 579.

Langan, T. A. & Smith, L. K. 1967 Fedn Proc. Fedn Am. Socs exp. Biol. 26, 103.

Lee, K. L. & Nickol, J. M. 1974 J. biol. Chem. 249, 6024-6026.

Linn, T. C., Pettit, F. H. & Reed, L. J. 1969 Proc. natn. Acad. Sci. U.S.A. 64, 227-234.

Linn, T. C., Pettit, F. H. & Reed, L. J. 1979 Proc. natn. Acad. Sci. U.S.A. 62, 234-241.

Linn, T. C. & Srere, P. A. 1979 J. biol. Chem. 254, 1691-1698.

Ljungström, O., Hjelmquist, G. & Engström, L. 1974 Biochim. biophys. Acta 358, 289-298.

Marchmont, R. J. & Houslay, M. D. 1980 Nature, Lond. 285, 904-906.

Martelo, O. J. & Hirsch, J. 1974 Biochem. biophys. Res. Commun. 58, 1008-1015.

Mendicino, J., Beaudreau, C. & Bhattacharyya, R. N. 1966 Archs Biochem. Biophys. 116, 436-445.

Meyer, W. L., Fischer, E. H. & Krebs, E. G. 1964 Biochemistry, Wash. 3, 1033-1039.

Milstein, S., Abita, J. P., Chang, N. & Kaufman, S. 1976 Proc. natn. Acad. Sci. U.S.A. 73, 1591-1593.

Morgenroth, V. H. III, Hegstrand, L. R., Roth, R. H. & Greengard, P. 1975 J. biol. Chem. 250, 1946-1948.

Nimmo, H. G. & Houston, B. 1978 Biochem. J. 176, 607-610.

Nishimura, J., Huang, J. S. & Deuel, T. F. 1982 Proc. natn. Acad. Sci. U.S.A. 79, 4303-4307.

Northrop, J. H. 1930 J. gen. Physiol. 13, 739.

Ozawa, E., Hosoi, K. & Ebashi, S. 1967 J. Biochem., Tokyo 61, 531-533.

Parker, P. J. & Randle, P. J. 1978 FEBS Lett. 95, 153-156.

Perrie, W. T., Smillie, L. B. & Perry, S. V. 1973 Biochem. J. 135, 151-164.

Rall, T. W., Sutherland, E. W. & Berthel, J. 1957 J. biol. Chem. 224, 463-475.

Reed, L. J. 1981 Curr. Top. cell. Regul 18, 95-106.

Riou, J. P., Claus, T. H., Flockhart, D. A., Corbin, J. D. & Pilkis, S. J. 1977 Proc. natn. Acad. Sci. U.S.A. 74, 4615-4619.

Rizack, M. A. 1964 J. biol. Chem. 239, 392-395.

Rose, K. M. & Jacob, S. T. 1979 J. biol. Chem. 254, 10256-10261.

Sanghri, A., Grassi, E., Warty, V., Diven, W., Wight, C. & Lester, R. 1981 Biochem. biophys. Res. Commun. 103, 886-892.

Schlender, K. K., Wei, S. H. & Villar-Polasi, C. 1969 Biochim. biophys. Acta 191, 272-278.

Sharma, R. K., Wang, T. H., Weich, E. & Wang, J. H. 1980 J. biol. Chem. 255, 5916-5923.

Smith, D. L., Chen, G. C., Bruegger, B. B., Holtz, S. L., Halpern, R. M. & Smith, R. A. 1974 Biochemistry, Wash. 13, 3780-3785.

Soderling, R. A., Hickenbottom, J. P., Reimann, E. M., Hunkeler, F. L., Walsh, D. A. & Krebs, E. G. J. biol. Chem. 245, 6317-6328.

Steinberg, R. A., O'Farrell, P. H., Friedrich, V. & Coffino, P. 1977 Cell 10, 381-391.

Sutherland, E. W. Jr & Wosilait, W. D. 1955 Nature, Lond. 175, 169-171.

Tegtmeyer, P., Rundell, K. & Collens, J. K. 1977 J. Virol. 21, 647-657.

Trzeciak, W. H. & Boyd, G. S. 1974 Eur. J. Biochem. 46, 201-207.

Tsiapalis, C. M., Trangas, T. & Gounaris, A. 1982 FEBS Lett. 140, 213-218.

Ushiro, H. & Cohen, S. 1980 J. biol. Chem. 255, 8363-8365.

Van Schaftingen, E., Davies, D. R. & Hers, H. G. 1981 Biochem. Biophys. Res. Commun. 103, 362-368.

Walinder, 1973 Biochim. biophys. Acta 293, 140.

Walsh, D. A., Perkins, J. P. & Krebs, E. G. 1968 J. biol. Chem. 243, 3763-3765.

Witte, O. N., Dasgupta, A. & Baltimore, D. 1980 Nature, Lond. 283, 826-831.

### E.C. KREES

## PROTEIN KINASE: CLASSIFICATION AND HISTORY

```
Krein, E. C., Ornica, D. J. & Parker, E. H. 1989, J. Sel. Chee, 734, 1931-1973.
                                                                                                                                                                                King J. K. Schreiter and A. Leith and A. Lei
          Acquar, U. J. of Mandal, Mr. 1925, Nov. and Add Mile Land Street, Mr. 1965, April 2015, A. T. propried
          Langeau, T. A. S. Stoith, L. Ellington Was Prise Indianal No. 80, 18th 52 and C. M. Nedyell M. D. W. redraid
         Probably N. P. Perro, J. A. & Breit, R. D. Companies and Said Land Marge July Science & J. M. and
        Line, T. C., Berth, Folk, Michael L. Lady Steel and Askar Michael St. St. St. Co., Tone, M. J. C., and
        Line, T. G., Fright, R. H. & Raweringth 1894, Courselle Searchief Mr. Mr. a. ben. 244, San, March. C. J. J. Valley. Line, T. G. & Sirre, P. A. 1979, J. Sid, Chee, 188, 1091-651142, A. April . Added to the Course of the Course 
       Brown, J., Britan, L., Order, March and A. Chamilland and Angelia Series, J. Amil's M. J. O. observed
        Endrow, S. Apart Silv. Mr. Administration of the World Property of the State of the
        Buretall, A. & Serend, Bugonirron & many policient to the A. S. C. Indeed, A. A. Standard, M. M. Standard, W. Standard, M. S. Mandard, W. Standard, M. S. Mandard, M. Mandard, M. S. Mandard, M. Mandard, M. S. Mandard,
       Milatein, S., Anie, J. P., Courg, N. & Kaufmad, S. Pipel Projetical, & V. S.A. Pattion of pg. mained
Mangement, V. H. III, Regressed, L. R., Rock, W. W. & Greenwall, P. 1988, J. Rad Conf. 136, 1944, 1548.
Mineson M. Greenway, N. oppositional, J. 198, 601 and progression of the Conf. 188, 1944, 1548.
                                Nerdigmen, J., Hunner, J., & B. Deugi, T. D. 19th West with a self-trail 1948 79 4853 4854 1874 1874 1874
                                                                                                                                                                                                                                                                                                                                                                                Northrop, J. H. 1930 A. pen Priprial 13, 739.
      Colors, F., Surchall, A., Pairley, J.C., objects of the later, Managed Principles of Association of Society, St.
                                                                                                                                                                                                                                                                                                  Parter, S. L. & Randle, E. J. 1978, PARK Latt. 95, 139-160.
       Paris, W. T., Smille, L. R. & Perry, S. W. 1972 Street, A. 1992 on Apprilia 2 S. R., resided S. S. S. S.
      Staff, T. W., Sucherland, E. W. & Rentell J. ways White Show dock and and and A. R. A., or hard ple property of the Control of
      Taxa and Z. & Cobjet, R. Super Sec. J. Physics, 133, 57 art.
                                                                                                                                                                                                                                                                                                                                                   Minuck, M. A. 1964 J. birk Chris 339, 392-395.
      Rose, E. St. J. Jacob, S. T. 1972 J. Mal. Gills 38 Rolling Start School, A. 1771. M. O., march in H. R., marches
      Sainthel, A., Crass, R., Warry, V., Dives, W., Wighl, W. Strissen, Rivered. Indiana Strict, Ad. Colored. May.
     Statement H. R., Wei et M. W. Wilson Poles and Statement Statement and 181 Strategy of the control of the Statement of the St
      Sharma, R. M., War, T. Phy Well, M. Ravinne, 1945, 1989, N. See, Unit. 1985, 1971, 1982, S. V. L.
                                                                             Smith, D. L., Chart, C. C., Sangger E. S., Hole, S. L., Margarit, A. & Hendig, A. G., 1844, Sangara, C. and C. and
     Sedering, R. A., Hickenbouton, J. P. Reinsein, S. M. Hickers, T. E., Walls, T. R. & Rein, R. C. V. List.
Chen. 245, 6317-6331
     Steinberg, R. A., O'Fursell, N. M., Schulerds, V. McCallad P. Took Cale at 181 agent school R. M. R., actual
 Walsh, D. A., Perkins, J. P. & Kreitt, S. G. agos. A. Sevi. Ches. 343, 3765-3765. L. mirel. S. G. against all seviness of the State of 
 Karm, S., Younds, M., Corna, K. & Suchman, of 1950 Processed day, S. J. S.A. W. map-cycl.
Ring L E Company, G. & Comm. S. 1980 Discountry, West, 12, 1886 1886.
Kingtenerila Ladi, Alleien V. G. & Mirky, & E. 1965. Francisco dessent file of the state of the Ericky, D. G. & Bravel L. A. 1970. A Sec. dischen dis. 425-050.

Krone E. D. & Pikelor, E. M. 1986. Fishing biother. Acts. 20, 130-147.
```

Appropriate on Control of Giverne Metabolism, pp. 300-240. London L. & A. Charchill.