## Cell Culture Methods for Molecular and Cell Biology

David W. Barnes, David A. Sirbasku, and Gordon H. Sato, Editore

Volume 1: Methods for Preparation of Media, Supplements, and Substrata for Serum-Free Animal Cell Culture

## Methods for Preparation of Media, Supplements, and Substrata for Serum-Free Animal Cell Culture

# Editors David W. Barnes

Department of Biological Sciences University of Pittsburgh Pittsburgh, Pennsylvania

## David A. Sirbasku

Department of Biochemistry and Molecular Biology University of Texas Medical School Houston, Texas

## Gordon H. Sato

W. Alton Jones Cell Science Center Lake Placid, New York

### Address all Inquiries to the Publisher Alan R. Liss, Inc., 150 Fifth Avenue, New York, NY 10011

## Copyright © 1984 Alan R. Liss, Inc. Printed in the United States of America.

Under the conditions stated below the owner of copyright for this book hereby grants permission to users to make photocopy reproductions of any part or all of its contents for personal or internal organizational use, or for personal or internal use of specific clients. This consent is given on the condition that the copier pay the stated per-copy fee through the Copyright Clearance Center, Incorporated, 21 Congress Street, Salem, MA 01970, as listed in the most current issue of "Permissions to Photocopy" (Publisher's Fee List, distributed by CCC, Inc.), for copying beyond that permitted by sections 107 or 108 of the US Copyright Law. This consent does not extend to other kinds of copying, such as copying for general distribution, for advertising or promotional purposes, for creating new collective works, or for resale.

## Library of Congress Cataloging in Publication Data

Main entry under title:

Methods for preparation of media, supplements, and substrata for serum-free animal cell culture.

(Cell culture methods for molecular and cell biology;

v. 1)

Includes index.

1. Culture media (Biology) 2. Cell culture.

I. Barnes, David W. (David William), 1949-

II. Sirbasku, David A. (David Andrew), 1941-III. Sato, Gordon. IV. Title: Serum-free animal cell

III. Sato, Gordon. IV. Title: Serum-free animal ce culture. V. Series.

QH585.M46 1984 ISBN 0-8451-3800-6 591'.07'24

84-7203

## **Contributors**

Steven K. Akiyama, Membrane Biochemistry Section, Laboratory of Molecular Biology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [215]

Mario A. Anzano, Laboratory of Chemoprevention, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [181]

Richard K. Assoian, Laboratory of Chemoprevention, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [181]

David W. Barnes, Department of Biological Sciences, University of Pittsburgh, PA 15260 [xxi,245]

Ralph A. Bradshaw, Department of Biological Chemistry, California College of Medicine, University of California at Irvine, Irvine, CA 92717 [139]

Albert E. Chung, Department of Biological Sciences, University of Pittsburgh, Pttsburgh, PA 15260 [321]

Paula Fehnel, Laboratory of Tumor Immunology and Biology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [295]

Charles A. Frolik, Laboratory of Chemoprevention, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [181]

Denis Gospodarowicz, Cancer Research Institute and the Departments of Medicine and Ophthalmology, University of California Medical Center, San Francisco, CA 94143 [69,275]

Lawrence A. Greenstein, Endocrinology Section, Metabolism Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [111]

Richard G. Ham, Department of Molecular, Cellular, and Developmental Biology, University of Colorado, Boulder, CO 80309 [3]

Robert A. Harper, Department of Dermatology, Temple University School of Medicine, Philadelphia, PA 19140 [147]

John R. Hassell, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [231]

A. Tyl Hewitt, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [239]

Michael Klagsbrun, Department of Surgical Research, Children's Hospital Medical Center, Boston, MA 02115 [159]

Hynda K. Kleinman, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [231]

Steven R. Ledbetter, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [231]

Lilly Lee, Endocrinology Section, Metabolism Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [111]

Lance A. Liotta, Laboratory of Pathology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [295]

Thomas Maciag, Department of Cell Biology, Revlon Biotechnology Research Center, Springfield, VA 22151 [195]

**Shing Mai,** Department of Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260 [321]

George R. Martin, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [231,239]

Wallace L. McKeehan, W. Alton Jones Cell Science Center, Lake Placid, NY 12946 [209] Alan C. Moses, Endocrinology Diabetes Unit, Beth Israel Hospital, Boston, MA 02215 [111]

S. Peter Nissley, Endocrinology Section, Metabolism Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [111]

Mounanandham Panneerselvam, Laboratory of Tumor Immunology and Biology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [295]

Elaine W. Raines, Department of Pathology, University of Washington, Seattle, WA 98146 [89]

Matthew M. Rechler, Section on Biochemistry of Cell Regulation, Laboratory of Biochemical Pharmacology, NIADDK, National Institutes of Health, Bethesda, MD 20205 [111]

Anita B. Roberts, Laboratory of Chemoprevention, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [181]

Russell Ross, Departments of Pathology and Biochemistry, University of Washington, Seattle, WA 98146 [89]

**Jeffrey S. Rubin,** Department of Biological Chemistry, Washington University School of Medicine, St. Louis, MO 63110 [139]

Atul Sahai, Laboratory of Tumor Immunology and Biology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [295]

David S. Salomon, Laboratory of Tumor Immunology and Biology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [295]

Gordon H. Sato, W. Alton Jones Cell Science Center, Lake Placid, NY 12946 [xxi] C. Richard Savage, Jr., Department of Biochemistry, Temple University School of Medicine, Philadelphia, PA 19140 [147]

Yuen W. Shing, Department of Surgical Research, Children's Hospital Medical Center and Harvard Medical School, Boston, MA 02115 [159]

Patricia A. Short, Endocrinology Section, Metabolism Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [111]

Janet Silnutzer, Department of Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260 [245]

David A. Sirbasku, Department of Biochemistry and Molecular Biology, University of Texas Medical School, Houston, TX 77225 [xxi]

Michael B. Sporn, Laboratory of Chemoprevention, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [181]

**K.S. Stenn,** Department of Dermatology, Yale University School of Medicine, New Haven, CT 06510 [269]

Victor P. Terranova, Laboratory of Developmental Biology, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [295]

Hugh H. Varner, Laboratory of Developmental Biology and Anomalies, National Institute of Dental Research, National Institutes of Health, Bethesda, MD 20205 [239]

Charity Waymouth, The Jackson Laboratory, Bar Harbor, ME 04609 [23]

Robert Weinstein, Departments of Pathology and Medicine, Charles A. Dana Research Institute, Harvard-Thorndike Laboratory, Harvard Medical School and Beth Israel Hospital, Boston, MA 02215 [195]

Kenneth M. Yamada, Membrane Blochemistry Section, Laboratory of Molegular Blology, National Cancer Institute, National Institutes of Health, Bethesda, MD 20205 [215]

Yvonne W.-H. Yang, Section on Biochemistry of Cell Regulation, Laboratory of Biochemical Pharmacology, NIADDK, National Institutes of Health, Bethesda, MD 20205 [111]

## Contents of Volumes 2, 3, and 4

## Volume 2: Methods for Serum-Free Culture of Cells of the Endocrine System

- 1 Growth of GH<sub>3</sub>, a Rat Pituitary Cell Line, in Serum-Free, Hormone-Supplemented Medium Izumi Hayashi
- 2 Growth of Adrenocortical Cell Cultures in Serum-Free Medium Michael H. Simonian and Mark L. White
- 3 Primary Culture of Testicular Somatic Cells J.P. Mather and D.M. Phillips
- 4 Isolation and Growth of Adult Human Prostatic Epithelium in Serum-Free, Defined Medium Mukta M. Webber, Donna M. Chaproniere-Rickenberg, and Robert E. Donohue
- 5 Growth of Functional Primary and Established Rat Ovary Cell Cultures in Serum-Free Medium Joseph Orly
- 6 Growth of Rat Mammary Tumor Cells in Serum-Free, Hormone-Supplemented Medium Tamiko Kano-Sueoka
- 7 Growth of Normal Mammary Epithelium on Collagen in Serum-Free Medium William R. Kidwell, Mozeena Bano, and David S. Salomon

- 8 Isolation and Serum-Free Cultivation of Mammary Epithelial Cells Within a Collagen Gel Matrix Walter Imagawa, Yasuhiro Tomooka, Jason Yang, Raphael Gu2man, James Richards, and Satuabrata Nandi
- 9 Serum-Free Culture of the Isolated Whole Mammary Organ of the Mouse: A Model for the Study of Differentiation and Carcinogenesis Mihir R. Baneriee and Michael Antoniou
- 10 Growth of Human Mammary Epithelial Cells in Monolayer Culture Martha Stampfer
- Definition of Hormones and Growth Factors Required for Optimal Proliferation and Expression of Phenotypic Responses in Human Breast Cancer Cells Marc E. Lippman
- 12 Serum-Free Cell Culture of MCF7 Human Mammary Carcinoma David W. Barnes
- 13 General Methods for Isolation of Acetic Acid- and Heat-Stable Polypeptide Growth Factors for Mammary and Pituitary Tumor Cells
  Tatsuhiko Ikeda, David Danielpour, Peter R. Galle, and David A. Sirbasku

Volume 3: Methods for Serum-Free Culture of Epithelial and Fibroblastic Cells

## SUBLIM-FREE OULTURE OF EPITHELIAL CELLS

- Growth of Primery and Established Kidney Cell Cultures in Serum-Free Media Mary. Faub
- 2 Howmonally Defined, Serum-Free Medium for a Proximal Tubular Kidney Epithelial Cell Line, LLC-PK<sub>1</sub> Million H. Saier, Jr.
- 3 Serum-Free Organ Culture of Embryonic Mouse Metanephres Ellis D. Ayner, William E. Sweeney, Jr., and Demetrius Ellis
- 4 Primary Culture of Hepatocytes H.L. Leffert, K.S. Koch, and H. Skelly

- 5 Selective Growth of Human Small Cell Lung Cancer Cell Lines and Clinical Specimens in Serum-Free Medium Desmond N. Carney, Martin Brower, Virginia Bertness, and Herbert K. Ole
- 6 Primary Tissue Cultures of Human Colon Carcinomas in Serum-Free Medium: An In Vitro System for Tumor Analysis and Therapy Experiments Jürgen van der Bosch
- 7 Growth and Differentiation of Human Bronchogenic Epidermoid Carcinoma Cells in Serum-Free Media Kaoru Miyazaki, Hideo Masui, and Gordon H. Sato
- 8 Serum-Free Cell Culture of A431 Human Epidermoid Carcinoma David W. Barnes
- 9 Growth and Differentiation of Embryonal Carcinoma Cells in Defined and Serum-Free Media Angie Rizzino
- 10 α<sub>2</sub>-Macroglobulin, a Contaminant of Commercially Prepared Pedersen Fetuin: Isolation, Characterization, and Biological Activity David S. Salomon, Kathryn B. Smith, Ilona Losonczy, Mozeena Bano, William R. Kidwell, Giulio Alessandri, and Pietro M. Gullino

#### SERUM-FREE CULTURE OF FIRRORLASTIC CELLS

- On Deciding Which Factors Regulate Cell Growth
  Arthur B. Pardee, Paul V. Cherington, and Estela E. Medrano
- Purification of Pituitary and Brain Fibroblast Growth Factors and Their Use in Cell Culture Denis Gospodarowicz
- Preparation of Bovine Pituitary Fibroblast Growth Factor Sandra K. Lemmon and Ralph A. Bradshaw
- Preparation of Pitultary Acidic FGF Angelo A. Gambarini, Mari C S. Armelin, and Hugo A. Armelin
- 15 Growth of SV40 BALB/c-3T3 Cells in Serum-Free Culture Medium G.A. Rockwell

- 16 Use of Hormone-Toxin Conjugates and Serum-Free Media for the Isolation and Study of Cell Variants in Hormone Responses Nobuyoshi Shimizu
- 17 Growth of Human Fibroblast Cultures in Serum-Free Medium Richard G. Ham
- Serum-Free Cell Culture for Growth of NIH 3T3 and 10T1/2 Mouse Embryo Fibroblast Cell Lines, SV40 Virus Propagation and Selection of SV40-Transformed Cells Lin-Chang Chiang, Janet Silnutzer, James M. Pipas, and David W. Barnes

Volume 4: Methods for Serum-Free Culture of Neuronal and Lymphoid Cells

#### SERUM-FREE CULTURE OF NEURONAL CELLS

- Culture Methods for Growth of Neuronal Cell Lines in Defined Media Jane E. Bottenstein
- 2 Preparation of a Chemically Defined Medium for Purified Astrocytes Richard S. Morrison and Jean de Vellis
- 3 Growth and Differentiation of Pheochromocytoma Cells in Chemically Defined Medium R. Goodman
- Differentiated Mouse Fetal Hypothalamic Cells in Serum-Free Medium
   A. Faivre-Bauman, J. Puymirat, C. Loudes, and A. Tixier-Vidal
- 5 Regulation of Pigmentation and Proliferation in Cultured Melanocytes John M. Pawelek
- 6 Neuron-Glia Interaction in Mammalian Brain: Preparation and Quantitative Bioassay of a Neurotrophic Factor (NTF) From Primary Astrocytes Wilfried Seifert and Hans Werner Müller
- 7 Preparation and Assay of Nerve Growth Factor Thomas L. Darling and Eric M. Shooter

#### SERUM-FREE CULTURE OF LYMPHOID CELLS

- 8 Production and Purification of Interleukin-2 for the Initiation and Maintenance of T-Cell Lines
  Diane Mochizukt and James D. Watson
- 9 Methods for Production and Purification of Human T-Cell Growth Factor
  M.G. Sarngadharan, R.C. Ting, and R.C. Gallo
- 10 Preparation of Thymosins
  Teresa L.K. Low and Allan L. Goldstein
- Culture of Lymphocytes and Hemopoietic Cells in Serum-Free Medium
  N.N. Iscove
- 12 Growth of Lymphoid Cells in Serum-Free Medium Frederick J. Darfler and Paul A. Insel
- 13 Serum-Free Cultivation of Plasmacytomas and Hybridomas Hiroki Murakami
- 14 Culture of Human Lymphocytes in Serum-Free Medium John Mendelsohn, Alendry Caviles, Jr., and Janice Castagnola
- Studies of Growth and Differentiation of Human Myelomonocytic Leukemia Cell Lines in Serum-Free Medium Theodore R. Breitman, Beverly R. Keene, and Hiromichi Hemmi
- Serum-Free Growth of SP2/0-AG-14 Hybridomas Kathelyn Sue Steimer

## **Preface**

For several of the past decades, experimental approaches to the study of cellular physiology were limited to either studies in the whole animal, or short-term studies with isolated cells or slices from various tissues. On the one hand, animal studies have provided our basic understanding of organ system physiology, but on the other hand this approach was limited because the in vivo measurements represented the net effect of many homeostatic changes that occurred during the experiments. In short, what was needed was a new experimental approach to cellular physiology in which isolated, functionally differentiated cells could be maintained in culture under conditions that allowed direct manipulations of the environment and measurement of the resulting changes in the function of a single cell type. The maintenance of cells in vitro has proven to be a major scientific undertaking of great importance, and it is now universally accepted that in vitro methods are indispensible to the study of cellular physiology.

Cell culture methods have been evolving rapidly. The first researchers in the field recognized that to establish even short-term cultures of animal cells in vitro, a basic requirement was the design of chemically defined synthetic media that contained the necessary mixtures of nutrients, vitamins, ions, and other essential metabolites, all of which were then to be maintained at the proper pH for life functions to be expressed. Pioneers such as Ted Puck, Charity Waymouth, Harry Eagle, Richard Ham, and others, led the efforts to provide all the workers of this field with formulations useful for specific cell types or purposes, and other formulations of synthetic media that were useful with a wide variety of cell types.

As these types of studies progressed, attention began to turn toward the problem of the most undefined component used in tissue culture media, namely the animal serum or plasma supplement. For a variety of cells in culture, it was widely accepted that the addition of this fluid was xxII Preface

essential for growth. Serum obviously was growth-stimulatory for some cell types; what was less obvious was that the serum supplement was toxic to other cell types or inhibitory to some physiological responses of interest. In nearly all cases, serum is the source of both growth-promoting and growth-inhibiting substances and, taken as a whole, serum-supplemented media greatly favor proliferation of mesenchymal-origin cells (i.e., fibroblasts) over growth of other cell types. Problems of apparent cellular "dedifferentiation" or fibroblast overgrowth were encountered by many researchers, using conventional culture techniques, and this led to a widely held view that cells which expressed differentiated functions could not be maintained in culture. This position was in part proven incorrect by workers demonstrating that functionally differentiated tumor cell types could be maintained in culture and that these cells continued expression of tissuespecific functions. But even with these initial successes, the problems remained of how to establish highly differentiated normal cells in culture selectively and how to support their proliferation in the absence of the undefined serum supplement.

In these four volumes, a different and unifying approach to cell culture is described. The concepts basic to this approach evolved from ideas contributed by investigators in areas of physiology and endocrinology, as well as those in fields more traditionally associated with cell culture methodology, and spring from attempts to replace serum in culture medium with purified components chosen to serve for cells the role usually provided by serum components. The methods are based on the experimentally established concepts that the function of the serum supplement is to supply hormones, growth factors, cell attachment proteins, extracellular enzymes and enzyme inhibitors, metal ion transport proteins and other binding proteins, lipoproteins, essential fatty acids, and other specific nutritional requirements for growth and tissue-specific expression by cells in culture. By supplementing media with appropriate mixtures of these defined substances, various differentiated cell types can be maintained in culture and be shown to express tissue-specific functions not previously identified in vitro.

In an attempt to satisfy the needs of both the novice and seasoned investigators wishing to utilize serum-free culture methods, we have organized the series beginning with this volume, which contains a description of the general methods of preparation of many of the common media components, followed by three other volumes, each dealing with detailed formulations for specific cell types. In this first volume, methods are presented for establishing the basal nutrient requirements of various cells and preparation of serum-free media, as well as complete methods

Preface xxIII

of preparation of several of the most common polypeptide growth factors used in defined media. Also, in this first volume, several important methods are described for the preparation of cell attachment proteins and extracellular matrix, which have proven to be essential components of many formulations of defined media. Careful evaluation of the properties and functions of the components described in Volume 1 will greatly aid the reader in the process of selecting which are likely to be important additions to the serum-free media desired. Volume 1 is organized to act as a companion to any of the other three, allowing the researcher to select one or more additional specialized volume of interest.

Beyond this general volume, the other three incorporate useful examples of how to apply these methods to individual problems. For example, in the second volume, entitled "Methods For Serum-Free Culture of Cells of the Endocrine System," methods are described for specific formulations effective with adrenal, pituitary, testicular, ovary, prostate, and mammary cells. The section on mammary cell culture is especially useful, since it is one of the most complete available on serum-free methods for cells from this organ.

Volume 3 of this series, "Methods for Serum-Free Culture of Epithelial and Fibroblastic Cells," describes much of what is currently known about the growth of these important cell types in a defined environment. This volume addresses the issues of culturing both normal and malignant epithelial cells, and the conditions that must be met to maximize expression of tissue-specific differentiated functions. Methods also described in Volume 3 deal directly with the uses of defined media to characterize the phenotypic and genotypic changes in untransformed and transformed fibroblasts. This field of research has received new interest with the discovery of oncogenes using in vitro transformation systems, and formulations of serum-free defined media for normal and transformed cells may become even more important with applications in these new directions.

Volume 4, the final of this series, is entitled "Methods for Serum-Free Culture of Neuronal and Lymphoid Cells." The first part of the volume focuses on applications to neuronal-origin cells that have been difficult to maintain in conventional serum-containing media. In addition, the preparation of neurotrophic factors is discussed in detail. The second part of Volume 4 addresses the methods of preparation of several of the important lymphokines, details growth conditions for lymphocytes, and also describes approaches to the growth of hybridomas and production of monoclonal antibody in serum-free, defined media. The latter method promises wide applications and is a broadly useful technical advance.

It has been our goal to provide a collection of culture methods that are

detailed and useful at the laboratory bench. For this reason, less emphasis has been placed in this series on the conceptual importance and implications of the various aspects of the serum-free systems described. More thorough treatments of these aspects can be found in several recent publications: "Growth of Cells in Hormonally Defined Media" (Cold Spring Harbor Press, 1982) and "The Use of Serum-Free and Hormonally Defined Media" (J. Mather, ed., Plenum Press, 1984). The editors thank the contributors for sharing their detailed methods and for their helpful comments which have greatly improved these volumes. We hope that the information provided here will serve as a catalyst for future advances and that these volumes will inspire new applications moving well beyond our present-day technology.

David W. Barnes David A. Sirbasku Gordon H. Sato

## Contents

	Contributors	XI
	Contents of Volumes 2, 3, and 4	xv
	Preface	. xxi
	METHODS FOR PREPARATION OF BASAL NUTRIENT MEDIA	9
1	Formulation of Basal Nutrient Media Richard G. Ham	. 3
2	Preparation and Use of Serum-Free Culture Media Charity Waymouth	. 23
3	Preparations and Uses of Lipoproteins to Culture Normal Diploid and Tumor Cells Under Serum-Free Conditions Denis Gospodarowicz	69
	METHODS FOR PREPARATION OF MITOGENIC PEPTIDES	
4	Preparation of Human Platelet-Derived Growth Factor Elaine W. Raines and Russell Ross	. 89
5	Purification of Multiplication-Stimulating Activity Lawrence A. Greenstein, S. Peter Nissley, Alan C. Moses, Patricia A. Short, Yvonne WH. Yang, Lilly Lee, and Matthew M. Rechler	111
6	Preparation of Guinea Pig Prostate Epidermal Growth Factor Jeffrey S. Rubin and Ralph A. Bradshaw.	139

vili	Contents
7	Purification of Human Epidermal Growth Factor From Urine C. Richard Savage, Jr. and Robert A. Harper
8	Isolation of Growth Factors From Human Milk Yuen W. Shing and Michael Klagsbrun
9	Purification of Type β Transforming Growth Factors From Nonneoplastic Tissues Anita B. Roberts, Charles A. Frolik, Mario A. Arızano, Richard K. Assoian, and Michael B. Sporn
10	Preparation of Endothelial Cell Growth Factor Thomas Maciag and Robert Weinstein
	METHODS FOR PREPARATION OF SUBSTRATA
11	Use of Basic Polymers as Synthetic Substrata for Cell Culture Wallace L. McKeehan
12	Preparation of Cellular Fibronectin Kenneth M. Yamada and Steven K. Akiyama
13	Isolation of Laminin Steven R. Ledbetter, Hynda K. Kleinman, John R. Hassell, and George R. Martin
14	Isolation of Chondronectin Hugh H. Varner, A. Tyl Hewitt, and George R. Martin
15	Human Serum Spreading Factor (SF): Assay, Preparation, and Use in Serum-Free Cell Culture  Janet Silnutzer and David W. Barnes
16	Purification of Epibolin From Human Plasma K.S. Stenn
17	Preparation of Extracellular Matrices Produced by Cultured Bovine Corneal Endothelial Cells and PF-HR-9 Endodermal Cells: Their Use in Cell Culture Denis Gospodarowicz