

RSC Drug Discovery

Human-based Systems for Translational Research

Edited by Robert Coleman

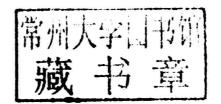


Human-based Systems for Translational Research

Edited by

Robert Coleman

Drug Discovery Consultant, Falmouth, UK Email: robt.coleman@btinternet.com







RSC Drug Discovery Series No. 41

Print ISBN: 978-1-84973-825-5 PDF eISBN: 978-1-78262-013-6

ISSN: 2041-3203

A catalogue record for this book is available from the British Library

© The Royal Society of Chemistry 2015

All rights reserved

Apart from fair dealing for the purposes of research for non-commercial purposes or for private study, criticism or review, as permitted under the Copyright, Designs and Patents Act 1988 and the Copyright and Related Rights Regulations 2003, this publication may not be reproduced, stored or transmitted, in any form or by any means, without the prior permission in writing of The Royal Society of Chemistry or the copyright owner, or in the case of reproduction in accordance with the terms of licences issued by the Copyright Licensing Agency in the UK, or in accordance with the terms of the licences issued by the appropriate Reproduction Rights Organization outside the UK. Enquiries concerning reproduction outside the terms stated here should be sent to The Royal Society of Chemistry at the address printed on this page.

The RSC is not responsible for individual opinions expressed in this work.

The authors have sought to locate owners of all reproduced material not in their own possession and trust that no copyrights have been inadvertently infringed.

Published by The Royal Society of Chemistry, Thomas Graham House, Science Park, Milton Road, Cambridge CB4 0WF, UK

Registered Charity Number 207890

For further information see our web site at www.rsc.org

Printed and bound by CPI Group (UK) Ltd, Croydon, CR0 4YY

Human-based Systems for Translational Research

RSC Drug Discovery Series

Editor-in-Chief:

Professor David Thurston, King's College, London, UK

Series Editors:

Professor David Rotella, Montclair State University, USA Professor Ana Martinez, Medicinal Chemistry Institute-CSIC, Madrid, Spain Dr David Fox, Vulpine Science and Learning, UK

Advisor to the Board:

Professor Robin Ganellin, University College London, UK

Titles in the Series:

- 1: Metabolism, Pharmacokinetics and Toxicity of Functional Groups
- 2: Emerging Drugs and Targets for Alzheimer's Disease; Volume 1
- 3: Emerging Drugs and Targets for Alzheimer's Disease; Volume 2
- 4: Accounts in Drug Discovery
- 5: New Frontiers in Chemical Biology
- 6: Animal Models for Neurodegenerative Disease
- 7: Neurodegeneration
- 8: G Protein-Coupled Receptors
- 9: Pharmaceutical Process Development
- 10: Extracellular and Intracellular Signaling
- 11: New Synthetic Technologies in Medicinal Chemistry
- 12: New Horizons in Predictive Toxicology
- 13: Drug Design Strategies: Quantitative Approaches
- 14: Neglected Diseases and Drug Discovery
- 15: Biomedical Imaging
- 16: Pharmaceutical Salts and Cocrystals
- 17: Polyamine Drug Discovery
- 18: Proteinases as Drug Targets
- 19: Kinase Drug Discovery
- 20: Drug Design Strategies: Computational Techniques and Applications
- 21: Designing Multi-Target Drugs
- 22: Nanostructured Biomaterials for Overcoming Biological Barriers
- 23: Physico-Chemical and Computational Approaches to Drug Discovery
- 24: Biomarkers for Traumatic Brain Injury
- 25: Drug Discovery from Natural Products
- 26: Anti-Inflammatory Drug Discovery
- 27: New Therapeutic Strategies for Type 2 Diabetes: Small Molecules
- 28: Drug Discovery for Psychiatric Disorders
- 29: Organic Chemistry of Drug Degradation
- 30: Computational Approaches to Nuclear Receptors
- 31: Traditional Chinese Medicine

- 32: Successful Strategies for the Discovery of Antiviral Drugs
- 33: Comprehensive Biomarker Discovery and Validation for Clinical Application
- 34: Emerging Drugs and Targets for Parkinson's Disease
- 35: Pain Therapeutics; Current and Future Treatment Paradigms
- 36: Biotherapeutics: Recent Developments using Chemical and Molecular Biology
- 37: Inhibitors of Molecular Chaperones as Therapeutic Agents
- 38: Orphan Drugs and Rare Diseases
- 39: Ion Channel Drug Discovery
- 40: Macrocycles in Drug Discovery
- 41: Human-based Systems for Translational Research

How to obtain future titles on publication:

A standing order plan is available for this series. A standing order will bring delivery of each new volume immediately on publication.

For further information please contact:

Book Sales Department, Royal Society of Chemistry, Thomas Graham House, Science Park, Milton Road, Cambridge, CB4 0WF, UK

Telephone: +44 (0)1223 420066, Fax: +44 (0)1223 420247

Email: booksales@rsc.org

Visit our website at www.rsc.org/books

Preface

Experimental animals have been a mainstay in the discovery and development of new medicines for human diseases for over half a century, so why do we now need to consider the use of humanised systems?

Historically, the pharmaceutical industry has been hugely successful, bringing a wide range of powerful and effective medicines to the market. Medicines to treat a wide range of disorders for which there had previously been, at best, inadequate treatment have been discovered and added to the clinician's armamentarium. It seemed that nothing could stop the almost exponential increase in the pharmaceutical industry's output. However, the last two to three decades have seen a marked decline in the productivity of this once prolific industry, 1,2 the effects of which have been stark, leading to a complete restructuring of the sector. Unfortunately, however dramatic such restructuring has been, it has not resulted in the hoped for increases in output of new safe and effective medicines per unit investment. Increasingly, when potential medicines developed using animal models are tested in human subjects, they are being found to fail for reasons of either lack of efficacy or associated use-limiting side effects and frank toxicity.

While lack of efficacy is of course a real problem, even more serious is the capacity of new medicines to do harm. In the early part of the 20th century, human medicines could be introduced onto the market with no legal requirement for the manufacturer to explore and report their safety profiles, but this changed following the Elixir sulfanilamide scandal. In 1937, a novel preparation of the antibacterial drug, sulfanilamide, was introduced to the US market, and was responsible for in excess of 100 fatalities. This led to Congress passing the 1938 Food, Drug, and Cosmetic Act, requiring companies to present safety data obtained in experimental animals on any proposed new drug, and to submit the data for approval by the FDA before

RSC Drug Discovery Series No. 41 Human-based Systems for Translational Research Edited by Robert Coleman © The Royal Society of Chemistry 2015 Published by the Royal Society of Chemistry, www.rsc.org viii Preface

being allowed to proceed to market. And so the use of animals to identify potential human hazards became routine within the pharmaceutical industry. The idea of using animals as human surrogates was based on the physiological parallels between humans and other animals, particularly those most closely related in evolutionary terms, i.e. mammalian species. While this was a clearly rational approach, it did not totally resolve the issue, and this was most clearly demonstrated in the 1950s with the thalidomide disaster.⁵ Thalidomide had been introduced as a mild sedative, particularly useful in reducing morning sickness in pregnancy, but was subsequently identified as the causative agent in the sudden explosion of cases of serious birth defects in the offspring of women who had been prescribed the drug. It is not that thalidomide had not been tested for side effects in experimental animals, it was simply the case that none had been performed on pregnant females to assess effects of the drug on the offspring, and interestingly, even if it had, because of species differences in this effect, the problem would probably not have been picked up. However, it was the thalidomide affair that consolidated the need for the rigorous safety evaluation of new potential medicines, and led to the increased use of animals as patient substitutes that persists to the present day.

As first-line testing for safety issues in human patients or volunteers was and remains ethically unacceptable, the logic behind using experimental animals on the basis of overall physiological similarity was uncontroversial. However, as time has passed, it has become increasingly clear that nonhuman species do not always respond to drugs in the same way as the humans that they are meant to represent, and their ability to reflect both clinical efficacy and safety issues has been shown to be highly variable, depending on particular drug type and disease target. Despite this, there has been a general acceptance by the medical establishment, the pharmaceutical industry and the regulators that overall, despite their shortcomings, they are doing a fair job, and with the lack of any obvious alternative, we have continued to rely on the animal-based status quo to assure the public of the safety of new medicines. But in the light of ever-increasing numbers of highprofile failures, including, but by no means restricted to, phen-fen, various statins^{7,8} and COX2 inhibitors, ^{9,10} this situation has become untenable, and it has become essential to do something to improve our ability to ensure the lack of safety problems with new medicines.

Much effort has been expended in exploring alternative approaches to new medicines R&D, and official organisations such as ECVAM (Europe) and ICCVAM (USA) have been established to determine their value. Unfortunately the output from these organisations is disappointingly low, ¹¹ and relatively few alternatives have received the necessary level of validation required for regulatory authorities to accept these tests as viable contributors to preclinical safety testing. Furthermore, even in cases where tests have been approved for use in drug submissions, there remains the issue of persuading the pharmaceutical industry to abandon their established animal-based methods in favour of the validated alternative.

Preface ix

Although a rigorous validation process is applied to new technologies, it is a fact that few if any of the animal-based tests that form the basis of our current safety testing paradigm have themselves been subjected to such scrutiny; their value has been taken as a given, despite the wealth of evidence of their shortcomings. Indeed, if one looks for peer-reviewed publications providing support for the status quo, there is really only one, and that merely reports an overall modest (approximately 70%) concordance between toxicity seen in animal species and in human subjects. In contrast, there is a wealth of publications providing data on the lack of value of animal data in identifying potential clinical safety issues. Indeed, a recent publication has explored the value of dogs as predictors of human safety profiles using more sophisticated measures of predictive power (likelihood ratios) than simple concordance, and have concluded that while demonstration of toxicity can in some cases reflect toxicity in humans (in line with Olson's findings), absence of toxicity in dogs provides no useful indication of absence in man. In the subject of the provides in the subject of the safety is applied to the subject of the safety is applied to subject to the safety in the safety is applied to subject t

If testing in animals is an unreliable indicator of potential clinical safety issues, then why are they still demanded and relied upon? A comprehensive answer to this question is outside the scope of this volume, but one key factor is the widespread belief that while flawed, they are superior to whatever else is available. 18 While there have long been proponents of a wider use of in vitro human systems, most moves in this direction have been rebuffed by claims that it is impossible to reflect the complexity of a whole integrated organism by looking at its parts in isolation, thus necessitating the use of animal surrogates. Such an attitude may be justified if there really are no viable alternatives available, and if the level of inaccuracy can be regarded as acceptable. For a long time, this attitude was (and in some quarters still is) the case, but technology moves on, and the seemingly impossible of vesterday becomes the possible of today, and the commonplace of tomorrow. With this in mind, together with the increasingly obvious inadequacies of the status quo, the exploration of human-based approaches seems unavoidable. Indeed, even the FDA, the primary regulatory authority in the USA, has come to this conclusion, stating on its website: "Consideration should be given to the use of appropriate in vitro alternative methods for safety evaluation. These methods, if accepted by all ICH regulatory authorities, can be used to replace current standard methods."19

It is fair to say that at this moment, we are not in a position to replace the current *in vivo* animal-based paradigm of assessing the potential safety and efficacy of new medicines with methods based on human *in vitro* models; there is much more work to do. However, we do now have greatly improved access to voluntarily donated viable human cells and tissues, an ever-increasing ability to generate a wide variety of tissue types from induced stem cells, and huge advances being made in a wide range of other technologies to apply to these materials, including those presented in this volume. With all this, we are now ready to perform proper evaluations of approaches which alone and in combination may offer superior means of establishing safety and efficacy of new medicines, consistent with 21st century science.

X Preface

The purpose of this book therefore is not to present a list of established assays that can reliably identify the potential efficacies and safety issues associated with new medicines, but rather to illustrate what is currently possible in humanising medicines R&D, which with sufficient commitment of time, effort and imagination will provide a basis on which to establish more rational and reliable means of developing safe and effective drugs for the future than those on which we currently rely.

Robert Coleman

References

- 1. J. W. Scannell, A. Blanckley, H. Boldon and B. Warrington, *Nat. Rev. Drug Discov.*, 2012, **11**, 191.
- 2. F. Sams-Dodd, Drug Discov. Today, 2005, 10, 139.
- 3. M. D. Breyer, Expert Opin. Drug Dis, 2014, 9, 115.
- 4. J. C. Geiger, Cal. West. Med., 1937, 47, 353.
- 5. A. E. Rodin, L. A. Koller and J. D. Taylor, J. Can. Med. Assoc., 1962, 86, 744.
- 6. H. M. Connolly, J. L. Crary, M. D. McGoon, D. D. Hensrud, B. S. Edwards, W. D. Edwards and H. V. Schaff, N. Engl. J. Med., 1997, 337, 581.
- 7. C. D. Furberg and B. Pitt, Curr Contr. Trials C, 2001, 2, 205.
- 8. R. S. Rosenson, Am. J. Med., 2004, 116, 408.
- 9. E. M. Antman, J. S. Bennett, A. Daugherty, C. Furberg, H. Roberts and K. A. Taubert, *Circulation*, 2007, **115**, 1634.
- 10. H. M. Krumholz, J. S. Ross, A. H. Presler and D. S. Egilman, *Br. Med. J.*, 2007, 334, 120.
- 11. M. Leist, N. Hasiwa, M. Daneshian and T. Hartung, Toxicol. Res., 2012, 1, 8.
- H. Olson, G. Betton, D. Robinson, K. Thomas, A. Monro, G. Kolaja,
 P. Lilly, J. Sanders, G. Sipes, W. Bracken, M. Dorato, K. Van Deun,
 P. Smith, B. Berger and A. Heller, Regul. Toxicol. Pharm., 2000, 32, 56.
- 13. P. Pound, S. Ebrahim, P. Sandercock, M. B. Bracken and I. Roberts, *Br. Med. J.*, 2004, **328**, 514.
- 14. A. Knight, Rev. Recent Clin. Trials, 2008, 3, 89.
- 15. R. A. J. Matthews, J. R. Soc. Med., 2008, 101, 95.
- 16. P. J. van Meer, M. Kooijman, C. C. Gispen-de Wied, H. Moors and H. Schellekens, *Regul. Toxicol. Pharmacol.*, 2012, **64**, 345.
- 17. J. Bailey, M. Thew and M. Balls, Altern. Lab. Anim., 2013, 41, 335.
- 18. http://www.animalresearch.info/ (accessed 12 May 2014).
- 19. http://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatory Information/Guidances/UCM194490.pdf (accessed 7 August 2014).

Contents

Chapter 1	Access to Human Cells and Tissues	
	Gerry Thomas	
	1.1 Ethics and Law Regarding Collection of Human	
	Samples	1
	1.1.1 The Four Cornerstones of Ethics and	
	Biological Samples	1
	1.1.2 Legal Issues	2
	1.2 Practical Issues Regarding Human Tissue	
	Collection and Annotation	8
	1.2.1 Obtaining Tissue with Consent for Research	9
	1.2.2 Accessing Material from a Diagnostic	
	Archive	11
	1.3 Improving Access and Annotation	12
	1.3.1 Access Policies	12
	1.3.2 Annotation	13
	1.4 Summary	14
	References	14
Chapter 2	Functional Studies with Human Isolated Tissues to Better	
-	Predict Clinical Safety and Efficacy	17
	David C. Bunton	
	2.1 Introduction	17
	2.2 Sourcing, Storing and Transporting Human Fresh	
	Tissues: The First Step is the Most Important	21

RSC Drug Discovery Series No. 41

Human-based Systems for Translational Research
Edited by Robert Coleman
© The Royal Society of Chemistry 2015
Published by the Royal Society of Chemistry, www.rsc.org

xii Contents

	2.3 Common Experimental Approac	enes to	
	Investigations in Isolated Fresh		23
	2.3.1 Tissue Baths and Wire M	Myographs 2	23
	2.3.2 Perfusion Myographs	2	25
	2.3.3 Organoculture Systems a	and Precision-Cut	
	Tissue Slices		25
	2.3.4 Membrane Transport an	d Ussing Chambers:	
	Skin, Lung Mucosa, Gas		
	and Glandular Tissues		28
	2.4 Applications of Functional Tiss		
	Development: Safety, Efficacy ar		
	Medicines		29
	2.4.1 Predicting Efficacy in Cl		
	Human Tissues		30
	2.4.2 Predicting Safety and To		00
	Functional Tissues		31
	2.4.3 Functional Tissues and		J L
	Personalised Medicines	_	32
	2.5 Summary		35
	Acknowledgements		35
	References		35
		`	00
Chapter 3	Translational Research in Pharmacol	ogy and Toxicology	
Chapter 3	Translational Research in Pharmacol Using Precision-Cut Tissue Slices		38
Chapter 3		3	38
Chapter 3	Using Precision-Cut Tissue Slices	3	38
Chapter 3	Using Precision-Cut Tissue Slices	rs, and P. Olinga	38 38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu	rs, and P. Olinga	
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction	rs, and P. Olinga	38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice	rs, and P. Olinga ces Sources of Human	38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different	rs, and P. Olinga ces Sources of Human	38 38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues	rs, and P. Olinga ces Sources of Human and Incubate	38 38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a	rs, and P. Olinga ces Sources of Human and Incubate ces from Various	38 38
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice	rs, and P. Olinga ces Sources of Human and Incubate ces from Various	38 38 40
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and	38 38 40
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Sl	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and	38 38 40 41
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and	38 38 40 41
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology 3.3 Application of Human Precision	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and	38 38 40 41
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology 3.3 Application of Human Precision Disease Models	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and a-Cut Tissue Slices in testine	38 38 40 41 43 47
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slice 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology 3.3 Application of Human Precision Disease Models 3.3.1 Fibrosis in Liver and Internal	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and a-Cut Tissue Slices in testine ses	38 38 40 41 43 47 47
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slices 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology 3.3 Application of Human Precision Disease Models 3.3.1 Fibrosis in Liver and Integration 3.3.2 Obstructive Lung Disease	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and 1-Cut Tissue Slices in testine ses	38 38 40 41 43 47 47 50
Chapter 3	Using Precision-Cut Tissue Slices G. M. M. Groothuis, A. Casini, H. Meu 3.1 Introduction 3.1.1 Precision-Cut Tissue Slices 3.1.2 Availability of Different Organs and Tissues 3.1.3 Techniques to Prepare a Precision-Cut Tissue Slice Organs and Tissues 3.2 Human Precision-Cut Tissue Slice Toxicology 3.3 Application of Human Precision Disease Models 3.3.1 Fibrosis in Liver and Interpretation 3.3.2 Obstructive Lung Disease 3.3.3 Cancer Research	rs, and P. Olinga ces Sources of Human and Incubate ces from Various ices in ADME and a-Cut Tissue Slices in testine ses	38 38 40 41 43 47 47 50 54

Contents xiii

Chapter 4	Modelling the Human Respiratory System: Approaches for in Vitro Safety Testing and Drug Discovery Human-Derived Lung Models; The Future of Toxicology Safety Assessment	66
	Zoë Prytherch and Kelly BéruBé	
	4.1 Introduction	66
	4.1.1 Overview of the Respiratory System	66
	4.1.2 Inhalation Toxicology	67
	4.1.3 Status Quo in Safety Assessment	68
	4.1.4 In Vitro Toxicology	68
	4.1.5 Trends and Technology Uptake	69
	4.2 Current Human-Based Models of the Respiratory	
	System: An Overview	70
	4.2.1 Cells and Tissues	70
	4.2.2 General Culture Conditions	76
	4.3 Discussion	80
	References	81
Chapter 5	Complex Primary Human Cell Systems for Drug Discovery Ellen L. Berg and Alison O'Mahony	88
	5.1 Introduction	88
	5.1.1 Challenges of Target-Based Drug Discovery	88
	5.1.2 Kinase Inhibitors – An Example Target Class	89
	5.1.3 Complex Primary Human Cell Systems for	
	Translational Biology	90
	5.2 Case Studies - Anti-Inflammatory Kinase Inhibitors	95
	5.2.1 JAK Kinase Inhibitors	95
	5.2.2 SYK Kinase Inhibitors	101
	5.3 Conclusions	105
	Acknowledgements	106
	References	106
Chapter 6	Human in Vitro ADMET and Prediction of Human	
	Pharmacokinetics and Toxicity Liabilities at the	
	Discovery Stage	110
	Katya Tsaioun	
	6.1 Introduction	110
	6.2 The Science of ADMET	112
	6.3 The ADMET Optimisation Loop	113
	6.4 Impact of Early Human Pharmacokinetics	
	Prediction	116

xiv Contents

	6.5 New H	uman ADMET Prediction Tools	117
	6.5.1	The Blood-Brain Barrier Challenge	117
	6.5.2	Mechanisms of Human Toxicity	121
	6.5.3	The Power of Multiparametric Screening	
		in Toxicology	125
	6.6 Bridgin	ng the Gap between in Vitro and in Vivo	126
		sions and Future Directions	128
	References		129
Chanter 7	'Rody-on-a-C	hip' Technology and Supporting	
onapter /	Microfluidic		132
		h, C. J. Long, C. McAleer, X. Guo, M. Esch,	132
		L. Shuler, and J. J. Hickman	
	J. 141. 1 100, 141.	. L. Shater, and J. J. Hickman	
	7.1 Introdu		132
	7.2 Multi-C	Organ <i>in Vitro</i> Models	133
		onal Single-Organ <i>in Vitro</i> Models	136
	7.3.1	Heart/Cardiac Tissue	137
	7.3.2	Lung	137
	7.3.3	Gastrointestinal (GI) Tract	139
	7.3.4	Liver	141
	7.3.5	Kidney	143
	7.3.6	Adipose Tissue	143
		Central Nervous System (CNS)/Peripheral	
		Nervous System (PNS)	144
		Skeletal Muscle	150
	7.3.9	Blood Surrogate	150
	7.4 Microfl		151
	7.5 Conclu	ding Remarks	153
	Acknowledge	8	153
	References		153
Chapter 8	Utility of Hu	ıman Stem Cells for Drug Discovery	162
ompter o	•	awar, Martin Graf, and Zameel Cader	
	8.1 Introdu		162
		g Approaches to Drug Discovery	164
		es in Human Stem Cell Technology	165
		Embryonic Stem Cells	165
		Reprogramming Somatic Cells	166
		ased Disease Models	167
	8.4.1	iPSC-Based Neurological and Psychiatric	
		Disease Models	167
		iPSC-Based Cardiovascular Disease Models	169
	8.4.3	Other Examples of iPSC-Based Disease	
		Models	172
	8.4.4	Genome Editing Tools	173

Contents

	8.5	Use of	iPSCs in Drug Efficacy Assessment	173
		8.5.1	Target-Based Screening versus Phenotypic	
			Screening	173
		8.5.2	Examples of Drug Testing to Validate iPSC-	
			Based Models	175
		8.5.3	Drug Screens on iPSC-Based Models	176
	8.6	Toxicit	y Testing Using iPSC-Based Models	177
		8.6.1	Cardiotoxicity	177
		8.6.2	Hepatotoxicity	178
		8.6.3	Neurotoxicity	178
	8.7	Integra	ation of iPSCs in Drug Discovery	179
		8.7.1	Challenges	179
		8.7.2	Future Directions	180
	8.8	Emerg	ing Resources of Diseased iPS Cell Lines	181
		8.8.1	StemBANCC (Stem Cells for Biological	
			Assays of Novel Drugs and Predictive	
			Toxicology)	181
		8.8.2	EBiSC (European Bank for Induced	
			Pluripotent Stem Cells)	182
		8.8.3	HipSci (Human Induced Pluripotent Stem	
			Cells Initiative)	183
	8.9	Summ	ary	183
	Refe	rences		183
Chapter 9	In Si	<i>lico</i> Sol	utions for Predicting Efficacy and Toxicity	194
•			att and Kevin P. Cross	
	9.1	Introd	uction	194
	9.2	Repres	enting Chemical Structures and	
		Associ	ated Data	196
		9.2.1	Overview	196
			Chemical Representation	196
			Biological Data Representation	199
			Calculated Data	199
			ing Chemical Databases	199
	9.4		ating Molecular Fragment Descriptors	201
			Overview of Fragment Types	201
			Predefined Features	202
			Structural Alerts	203
			Common Chemical Scaffolds	204
			Scaffolds Associated with Data	204
	9.5		standing Structure–Activity or Toxicity	
			onships	205
			Overview	205
		9.5.2	Classification Based on Substructures	205

	9.5.3 Clustering	205
	9.5.4 Decision Trees	207
	9.6 Quantitative Structure–Activity Relationship (QSAR)	
	Modelling	208
	9.6.1 Overview	208
	9.6.2 Building Models	208
	9.6.3 Applying Models	210
	9.6.4 Model Validation	211
	9.6.5 Explaining the Results	211
	9.7 Regulatory in Silico Case Study	212
	9.8 Prediction of Human Adverse Events Case Study	215
	9.9 Conclusions	216
	References	217
Chapter 10	In Cilias Occase Madallina in Destination 1966	
Chapter 10	In Silico Organ Modelling in Predicting Efficacy and	240
	Safety of New Medicines Blanca Rodriguez	219
	Bluncu Rouriguez	
	10.1 Introduction	219
	10.2 State-of-the-Art Computational Cardiac	
	Electrophysiology	220
	10.2.1 Single Cardiomyocyte Models	220
	10.2.2 Whole Organ Heart Models	223
	10.2.3 Simulating Interactions of Medicines with	,
	Ionic Channels	226
	10.3 Investigating Variability: Population of Models	
	Approach	230
	10.4 Validation of <i>in Silico</i> Models and Simulations	234
	References	236
-		
Chapter 11	Human Microdosing/Phase 0 Studies to Accelerate Drug	
	Development	241
	R. Colin Garner	
	11.1 Introduction	241
	11.2 What is Microdosing (Human Phase 0 Studies) and	
	Where Does it Add Value?	243
	11.3 Scientific, Regulatory and Ethical Aspects of	
	Microdosing	244
	11.3.1 Scientific Considerations	244
	11.3.2 Regulatory	245
	11.3.3 Ethical Considerations in Conducting	_ 10
	Microdose Studies	247
	11.4 Analytical Technologies used for Microdosing	⊿ 1/
	Studies	247
	11.5 Applications of Microdosing	249