NMR in Chemistry

A Multinuclear Introduction

WILLIAM KEMP

NMR in Chemistry

A Multinuclear Introduction

WILLIAM KEMP



© W. Kemp 1986

All rights reserved. No reproduction, copy or transmission of this publication may be made without written permission.

No paragraph of this publication may be reproduced, copied or transmitted save with written permission or in accordance with the provisions of the Copyright Act 1956 (as amended).

Any person who does any unauthorised act in relation to this publication may be liable to criminal prosecution and civil claims for damages.

First published 1986

Published by MACMILLAN EDUCATION LTD Houndmills, Basingstoke, Hampshire RG21 2XS and London Companies and representatives throughout the world

Typeset by TecSet Ltd, Sutton, Surrey Printed in Hong Kong

British Library Cataloguing in Publication Data Kemp, William, 1932-NMR in chemistry: a multinuclear introduction.

1. Nuclear magnetic resonance spectroscopy

1. Title
541.2'8 QD96.N8
ISBN 0-333-37291-3
ISBN 0-333-37292-1 Pbk

Preface

In the last few years the horizons of nuclear magnetic resonance spectroscopy have all but been pushed out of sight as new and ever more sophisticated instruments are built and reports of new applications flood the literature.

For chemists who are active developers of the science, in whatever aspect, it is difficult merely to read all of the new publications. For chemists who use the method, among other methods, to solve their chemical problems it is well-nigh impossible to keep up to date.

This book is an attempt to introduce NMR to a wide audience of users, in such a cohesive way that all of its potential can be tapped and exploited: it is not a book about the physics and mathematics of NMR, but concerns the interpretation of NMR spectra by those who do not consider themselves particularly mathematical by inclination: the minimum prerequisites are some knowledge of general spectroscopic principles and a familiarity with the chemical properties of common functional classes.

Clear non-mathematical pictures of magnetic resonance must inevitably be distortions of more fundamental laws, but a worthwhile sacrifice is made in forming a bridge between physics and chemical application. The book is obviously not monographic in style or depth, but it will serve its intent if, having read it, the reader is thereby equipped and encouraged to tackle the necessarily more rigorous specialist works.

The emphasis is multinuclear, with a deliberate attempt being made to demystify the NMR studies of the less common NMR elements. Likewise, Fourier transforms and superconducting magnets are introduced very early, and are not given any special status as 'recent developments': continuous wave spectrometers are not referred to as 'conventional', and the use of the terms upfield and downfield is minimised for the same pedagogic reasons (being replaced by lower frequency and higher frequency respectively).

The problem of order of presentation, of finding a beginning, a middle and an end, is not unique to the study of NMR: the chapter sequence adopted should not therefore be taken as a recommended learning programme,

but merely as an approximate indicator of the way in which most teachers introduce the subject, building upon the familiar to construct a framework of advanced understanding.

Chapter 1 is set at the most elementary conceptual level, and is an obligatory launching point for the multinuclear approach.

Chapter 4 (on proton NMR) and chapter 5 (on carbon-13 NMR) form the mainstay of the subject; some teachers begin with proton while others begin with carbon-13. Surprisingly little theory is needed to interpret such spectra: for this reason, while chapter 2 is fairly full in explanation, most students will tend to attack this chapter in several short sorties, as the need to know becomes more expansive. The same is true of the instrument details given in chapter 3 which, while fairly detailed and self-contained, can profitably be absorbed in separate parts at different times.

The set of chapters 2, 3, 4 and 5 completes a study of NMR to intermediate level.

Chapters 6 and 7 interrelate with each other at the next most advanced conceptual level: here are introduced the theories and practice made accessible only by microprocessor controlled instrumentation. An argument can be made for introducing some of the chapter 6 theory at an earlier stage, and of course this is not precluded: but there is a strong case to be adduced for consolidating the basic applications in the early chapters before tackling the three-dimensional complexities of the rotating frames of reference, with all the perceptual traps involved.

The study of dynamic molecular processes by NMR is treated separately in chapter 8. Some of the simpler ideas of dynamic NMR are, however, interspersed in the earlier chapters, since to gather them all in a late chapter would be artificial and distorting.

Chapters 9-13 are all self-sufficient to a degree, and are the barest indications, by example, of the kind of information contained in the NMR spectra of a selection of other nuclei. Given the multinuclear treatment of the earlier chapters, there is no need here for extensive detail.

Chapter 14 very much embodies two personal views.

Many details of physics are easily forgotten (or may not have been learnt) and having them gathered together serves as a useful aide-memoire for non-physicists. We also often learn nothing about the people who helped to assemble the enormous construct of science, and the sparse biographical notes given here on a few of the famous names in NMR may whet an appetite or two.

Although informative data are supplied as necessary throughout the book, chapter 15 contains a large amount of reference data, which can be accessed for systematic or detailed needs. Extensive tables of chemical shifts, coupling constants and relaxation times are gathered here, with details of common NMR solvents.

There are many problem examples throughout the book. It has been established that one major reason for lack of success in problem solving is an inability to collect and identify only that information which is necessary

for solution. In an endeavour to side-step this difficulty, most of the problem examples are preceded by worked examples, so that confidence will rise through knowing which method to apply. Most problems are also seen within the context of the learning objectives under discussion (although other problem examples test more comprehensive understanding).

While it is demonstrably possible to interpret many NMR spectra without intimate study of theory or of spectrometer operation, hopefully the presentation of the subject matter in this book will stimulate an intellectual curiosity, such that a spectrum can be interpreted in the morning, and explained in the afternoon. If the book fulfils its purpose, the user will go back the following morning for more. There is always another experiment to be done.

Heriot-Watt University, Edinburgh

W.K.

Acknowledgements

Thanks are due to innumerable people for help and information: for the supply of spectra, or for the permission to reproduce copyright material used in the figures in this book, several companies (and their personnel) have contributed, and this is recognised with gratitude.

The photograph of Felix Bloch shown in the Frontispiece was kindly supplied by Stanford University, California: that of E. M. Purcell was furnished by E. M. Purcell himself.

Bruker Spectrospin published the material in figures 5.3, 5.13, 5.14, 6.12, 7.4, 7.14, 7.16 and 13.2.

Japan Electronic Optical Laboratories, JEOL, supplied the photographs of the spectrometers shown at the head of chapter 3, and the spectra in figures 1.13, 5.1(b), 5.2 (lower), 5.4, 6.14, head of chapter 7 (also used on the book cover) and 11.1.

Oxford Instruments supplied the photograph of the superconducting magnet core and are copyright holders of the diagram showing the cryostat system in figure 3.1.

Oxford Research Systems supplied the photographs of the brain images in figure 7.21 and of the whole-body NMR magnet shown at the head of chapter 10.

Perkin-Elmer published the spectra used in figures 4.18, 4.24, 8.2, 9.2 and at the head of chapter 6.

The figures at the head of chapters 8 and 12 were used with the permission, respectively, of the Royal

Society of Chemistry and the American Chemical Society.

Varian Associates granted permission to use the spectra from their NMR Spectra Catalog, namely figures 4.6, 4.7, 4.8, 4.9, 4.10, 4.14, 4.15, 4.20, 4.25, 4.26, 4.27 and 4.28. Other figures which used spectra published by Varian are figures 5.2, 5.11, 5.12, 7.2, 7.5, 7.9, 7.17, 10.2, 10.4, 11.3, 11.4, 12.2, 14.5(c), 13.1, 13.3, 13.4 and 13.5; also those at the heads of chapters 9, 11 and 13.

The following were reproduced, with permission, from Kemp, *Organic Spectroscopy*, 2nd edn, London, Macmillan (1986): 3.2, 4.1, 4.2, 4.3, 4.4, 4.5, 4.12, 4.13, 4.17, 4.23, 6.7, 8.6, 14.5(a).

Johnson and Jankowsky, Carbon-13 NMR Spectra, New York, Wiley (1972) was the source (with permission) of figures 5.1(a), 5.5, 5.6, 5.7, 5.8, 5.9, 5.10 and 9.4.

Dalton's table of atomic symbols, which appears at the head of chapter 15, is reproduced by permission of the Trustees of the Science Museum (London).

Many colleagues helped by argument, or by reading part or all of the manuscript (Bill Steedman, Kevin McCullough and Alan Boyd — who also recorded several of the spectra): all are thanked for their encouragement.

CONTENTS

Prefa Ackn	ce owledgements	xi xiii	2.13	Nuclei with Spin Quantum Number Greater than 1	22
	,		2 14		22
1	Introduction to Nuclear Magnetic Resonance	-		Nuclear Quadrupole Resonance Spectroscopy	23
	NMR	1	2.13		22
1.1	The Principles of NMR Spectroscopy	2	2 16	6 Boltzmann Distributions in NMR – Effects	23
1.2	The NMR Spectrometer	3	2.10	6 Th. 1 S. G	
1.3	Chemical Shifts	4	2 17		24
1.4	Field Strength and Frequency	6	2.17	**	25
1.5	The Units of Chemical Shift	7			25
1.6	Frequencý Standards	7	2 18	The Heisenberg Uncertainty Principle and the	26
1.7	Chemical Shift Assignments	8	2.10	2577 2-2 0 4 4	~~
1.8	Two Models of Nuclear Magnetic	8			27
	Resonance			2.18.1 Effect of T_1 and T_2 on NMR Line Widths	
	1.8.1 Quantum Mechanics	8		2 10 2 Tt. 01 15 1 4	27
	1.8.2 The Precessing Nucleus	9		2.10.2 Line Shape and Peak Areas	27
1.9	Coupling and Decoupling of Magnetic				
	Nuclei	11	3	The NMR Spectrometer	29
			3.1	NMR Magnets	30
2	The Fundamental Basis of Magnetic Resonance	6 1 4		2 1 1 Damman A Mariana	30
		~ 14		2 1 2 Flactnoments	30
2.1	The Spin of Electrons and Nuclei	15		212 6	30
2.2	Angular Momentum and Magnetic Moments				٠.
	of the Electron and the Proton	15		Danabatan to	31
2.3	Angular Momentum and Magnetic Moments			3.1.5 The Direction of B_0 — The z-axis	
•	of Other Nuclei	16		C	32
2.4	The Spin Quantum Number	16	3.2	Magnetic Field Homogeneity and Spectro-	
	2.4.1 The Dimension of Spin Angular			maken Beenfester	32
	Momentum	17			32
	2.4.2 The Direction of Spin Angular			3.2.2 Sample Spinning and Spinning Side-	
	Momentum	17		h	32
2.5	Nuclei with Spin Quantum Number = $\frac{1}{2}$	17	3.3	Manual Carbillan	33
2.6	The Nuclear Magnetic Moment Vector, μ	17	3.4	Field Sweep and Frequency Sweep -	,,
2.7	The Strengths of Nuclear Magnets — The			Continuous Warm NRAD Co.	33
2.0	Nuclear Magneton Unit, μ _N	19	3.5	Pulsed Radiofrequency — Fourier Transform	
2.8	The Magnetogyric Ratio $-\gamma$	19		NMP Constants	34
2.9	Precessional Frequency and γ – The Larmor			3.5.1 Fourier Transforms	34
2 10	Equation	19		3.5.2 Advantages of the Pulsed FT NMR	
	Summary of NMR Theory	20		Mathad .	35
2.11	Electron Spin Resonance Spectroscopy	20	3.6	Sensitivity in Multinuclear NMR	36
2.12	Nuclei with Spin Quantum Number = 1	20		3.6.1 Relative Abundance	36

CONTENTS

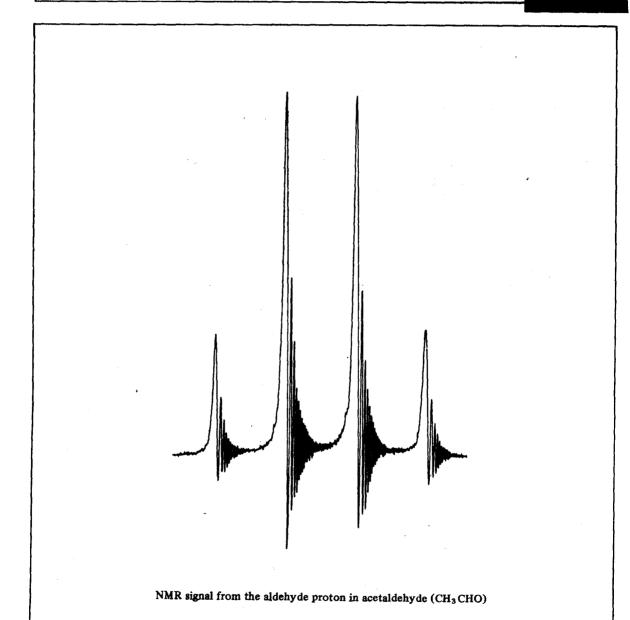
	3.6.2	Sensitivity and Nuclear Magnetic			4.4.2	Protons in Alkane Groups — lables	
	•	Moments	36			15.1, 15.2 and 15.3	55
3.7	Radio	frequency Circuits	36		4.4.3	Protons of Alkene, Alkyne and	
		Radiofrequency Sources in Multi-				Aromatic Systems — Tables 15.4	
		nuclear NMR	37			and 15.5	56
	3.7.2	Single Coil and Crossed Coil	•	4.5	Proton	s in OH, NH and SH Groups — Table	
		Detection	37		15.6		57
	3.7.3	Forced Induction and Free Induction	38		4.5.1	Hydrogen Bonding and Proton	-
		The Components of RF Signals	39			Chemical Shifts	57
		Phase Sensitive Detectors and	<i></i>		4.5.2	Quadrupolar Broadening in NH	
		Quadrature Detection	40			Signals	59
	3.7.6	RF Pulses and their Fourier	-10	4.6	Spin-S	Spin Coupling — Spin-Spin Splitting	59
		Transforms	40	4.0		Predicting Multiplicity from the	0,
3.8	Data F	landling in FT NMR	41		1.0.1	(n+1) Rule	59
	3.8.1		41		4.6.2	The Coupling Constant, J — Table	,
		The Fast Fourier Transform — Real	41		7.0.2	15.7	62
	5.0.2	and Imaginary Solutions	42		163	Theory of Spin-Spin Coupling — I	64
	383	Phases and Filters	42			AX ₂ , AX ₃ , and A ₂ X ₃ Coupling	0-
		Dynamic Range and Computer Word	42		4.0.4	Systems	
	3.0.4	Length	42		165		65
3.9	Vorioh	ole Temperature Probes	43 43		4.0.3	Coupling of Protons to Other Nuclei with Spin = $\frac{1}{2}$	
3.7	V ai ia c	ne Temperature Probes	43	47	Camar		60
		φ. φ.δ		4.7		al Features of Simple Coupling Systems	0
					4.7.1	Magnetic Equivalence in Proton	,,
4	Proton :	NMR Spectra	45		470	NMR	6
					4.7.2	Successive Branching and Pascal's	٠.
4.1		ing, Deshielding and Proton Chemical	16		4.50	Triangle	68
•	Shifts		46			Long Range Coupling	70
	4.1.1	Electronegativity and Proton	47		4.7.4	Coupling in Proton Exchange	
		Chemical Shifts	46			Processes	70
	4.1.2	•	45			AMX Coupling Systems	73
		Chemical Shifts	47	4.8	-	ing to Nuclei with Spin $I > \frac{1}{2}$	73
4.2		tropic Effects and Proton Chemical			4.8.1	Predicted Multiplicity for Spin = 1	
	Shifts		47			Nuclei	73
		Aromatic Ring Currents	47		4.8.2	Effects of Quadrupole Moments on	
		Double Bonds in Alkenes	49			Observed Splitting	74
		Carbonyl C=O Double Bonds	49	4.9	Non-fi	irst Order Spectra	7:
		Alkyne Triple Bonds	50	4.10	Simpli	ification of Complex Spectra	7
		Alkanes	51		4.10.1	Increased Field Strength	71
4.3	Chemi	ical and Magnetic Equivalence in			4.10.2	Chemical Shift Reagents	73
		n NMR	51		4.10.3	Spin Decoupling — Double and	
	4.3.1	Free Rotation and Molecular				Triple Irradiation	80
		Conformations	51	4.11	Additi	ional Problems	81
	4.3.2	Restricted Rotation in Alkenes and					
		Amides	52	_	a .	10 ND 60 0	
	4.3.3	Proton Exchange in Alcohols, Amines	,	5	Carbon-	13 NMR Spectra	84
		Carboxylic Acids and Keto-Enol		5.1	Recor	ding Carbon-13 NMR Spectra	85
		Systems	52		5.1.1	Natural Abundance and Isotopomers	85
	4.3.4	Deuterium Exchange	53		5.1.2	•	٥.
4.4		ns Attached to Carbon Atoms —				Ratio	85
		cal Shift Assignments	54		5.1.3	Coherent and Broad Band De-	9.
	4.4.1	Use of Correlation Tables 15.1-15.5	54			coupling - 13C{1H} NMR Spectra	87
						- Can I mark opposite	0/

						V	111
		Off Resonance Decoupling	88	6.2	Energy 6.2.1	Levels for an AX Coupling System The Sign of the Coupling Constant, J	111
5.2		ation Effects in Carbon-13 NMR	00			Factors Influencing the Coupling	112
	Specti		90		0.2.2	Constant, J	113
5.3		ical Shifts in Carbon-13 NMR	90	*	623	Non-first Order AB Spectra	114
	Specti		90	6.3		Levels for an AMX Coupling	117
	3.3.1	Theory of Carbon-13 Chemical Shifts	91	0.5	System		114
5.4	Cham	ical Shift Assignments in Carbon-13	71	6.4	•	nination of the Relative Signs of J	114
J. T		Spectra	92	0.4	6.4.1	Spin Tickling	115
	5.4.1	•	72			INDOR – Internuclear Double	
	3.4.1	Correlation Tables 15.8-15.16	93		o .	Resonance	115
	5.4.2		93	6.5	Net Ma	agnetisation and its Vector	
		Alkane Chemical Shifts — Table				entation	117
		15.8	94	6.6		s of Reference in NMR	118
	5.4.4	Alkene Chemical Shifts - Table				The Laboratory Frame	118
		15.9	95			Rotating Frames of Reference	118
	5.4.5	Alkyne Chemical Shifts — Table				Axes of Rotating Frames, x' , y' and	
		15.10	96			Z	119
	5.4.6	Aromatic and Heterocyclic		6.7	Pulsed	Radiofrequency in the Rotating	
		Chemical Shifts — Table 15.11	96		Frame		119
	5.4.7	Influence of Functional Substituents			6.7.1	Magnetisation on the y'-axis	119
		on Alkane Chemical Shifts — Table			6.7.2	Pulse Width and Flip Angle - 90°	
		15.12	97			and 180° Pulses	119
	5.4.8	Influence of Functional Substituents		6.8	Relaxa	tion in the Rotating Frame	122
		on Alkene and Aromatic Chemical			6.8.1		
		Shifts — Table 15.13	99			Lattice Relaxation $-T_1$	122
	5.4.9	Carbonyl Group Chemical Shifts —			6.8.2	Transverse Relaxation — Spin-Spin	
		Table 15.14	101			Relaxation $-T_2$	123
	5.4.10	Miscellaneous Multiple-bonded			6.8.3	T ₂ and Field Homogeneity	123
		Carbon Chemical Shifts — Table	101			Line Width, Ringing, T_1 and T_2	123
		15.15	101	6.9		uclear Overhauser Effect - NOE	124
	5.4.11	Methyl Group Chemical Shifts —	101		6.9.1		124
5.5	C1	Table 15.16	101		6.9.2	Theory of the NOE	126
0.0		ing Constants in Carbon-13 NMR	101		6.9.3	The Size of the NOE	126
	-	a — Table 15.17 — Satellites	101		6.9.4	The Sign of the NOE — Nuclei	
	3.3.1	Coupling of Carbon-13 to	102		605	with Negative γ	127
	552	Hydrogen and Deuterium Homonuclear Carbon Coupling	102		6.9.5	Time Dependence of Spin	
	5.5.3	Coupling of Carbon-13 to Other	102			Decoupling and the NOE	127
	3.3.3	** • •	102				
5.6	Additi	Nuclei onal Problems	102 103				
	Additi	onal i toolems	103	7		lse and Computational Methods in	
					NMR		128
ó	More A	dvanced Theories of NMR	108	7.1	Gatad	Decoupling and NOE	
5.1	Theory	y of Spin-Spin Coupling — II	109	,	7.1.1		129
		Electron-coupled Spin-Spin Inter-	10)		7.1.2	Gated Decoupling — NOE with	129
		actions between Directly-bonded			,,,,,,	Spin Splitting	100
		Nuclei	109		7.1.3	Inverse Gated Decoupling — Spin	129
	6.1.2	Electron-coupled Spin-Spin Inter-				Decoupling without NOE	121
		actions between Protons in CH ₂			7.1.4	Quantitative Analysis in Decoupled	131
		Groups	110			Carbon-13 NMR Spectra	131
			•			-	

7.2	Massuran	nent of T_1 – the Spin-Lattice (or			8.1.3	Line Shape (Band Shape) and	
7.2	Measuren	inal) Relaxation Time	131				163
	Zongitud	version Recovery Measurement			8.1.4	Relaxation Times and Exchange	,
		T_1	131			Rates	163
		ther Pulse Sequences for Measuring		8.2	Energy	y of Activation	164
	7.2.2 O	-	134		8.2.1	The Arrhenius Equation and the	
		ignificance of T ₁ Values	134			Eyring Equation	164
		eak Suppression Methods	135		8.2.2	Typical Values of Activation	
7.3	Measure	ment of T_2 — the Spin-Spin (or				Energies	165
1.5	Transver	se) Relaxation Time	136	8.3	Theor	y of Coalescence	165
	7.3.1 S	pin Echoes – the Simple Homo-		8.4	The N	MR Time Scale — What is it?	166
		nuclear Case	136	8.5		ically Induced Dynamic Nuclear	
		Carr-Purcell-Meiboom-Gill (CPMG)			Polari	sation - CIDNP	167
		Measurement of T_2	137		8.5.1	Applications of CIDNP	167
		Significance of T ₂ Values	138		8.5.2	The Nature of CIDNP	168
,		Coupling Constants and Phase					
1		Modulation in Spin Echoes	138	9	Elnorie	e-19 NMR Spectra	169
7.4		tion Transfer and Cross Polarisation	141	,			
/		Selective Population Inversion	142	9.1		ine-19 Chemical Shifts	170
7.5		rised Principles of Spin Echoes and		9.2	Fluor	ine-19 Coupling Constants	171
7.5		tion Transfer	143		9.2.1		
7.6		DEPT, and Spectrum Editing by				Fluorine-19	171
	ADEPT		143		9.2.2		
		Spectrum Editing — Carbon Sub-				Proton and Phosphorus-31	174
		spectra	143		9.2.3		
7.7		mensional NMR - 2D NMR	146			Carbon-13	174
,		J-resolved 2D NMR Spectra	148		9.2.4		
		Shift-correlated 2D NMR Spectra	149			Nitrogen-14	174
		Recording 2D FT NMR Spectra - τ					
		and the Second Time Domain Axis	149	10	Phospi	horus-31 NMR Spectra	176
		¹³ C Satellite Spectroscopy —			_		
		Carbon-Carbon Connectivity Plots		10.1		phorus-31 Chemical Shifts	177
		CCCP	149	10.2		phorus-31 Coupling Constants	177
7.8	Solid S	tate NMR Spectra	151		10.2	.1 Coupling of Phosphorus-31 with	,
	7.8.1	Magic Angle Spinning - MAS	152			Phosphorus-31	178
	7.8.2	Dipolar Decoupling and Cross			10.2	.2 Coupling of Phosphorus-31 with	. =0
	. 4	Polarisation	152			Proton and Fluorine-19	178
	7.8.	Applications of Solid State NMR	154		10.2	.3 Coupling of Phosphorus-31 with	4 600
' 7.9	Biglog	ical NMR	154			Carbon-13	178
	10.1	NMR Imaging - Zeugmatography,					
	Ø	Tomography	154	11	Nitro	gen-14 and Nitrogen-15 NMR Spectra	182
1	7.9.2	Surface Detection and Topical					183
38	,	Magnetic Resonance - TMR	156	11.1		ogen Isotopes – ¹⁴ N and ¹⁵ N	183
200					11.1	.1 Isotope Abundance	103
4. T					11.1	.2 Sensitivity Enhancement – INEPT	183
8	Dynami	ic NMR	158			and DEPT	184
			155	11.2		ogen Chemical Shifts	104
8.1		Processes and the Heisenberg			11.2	2.1 Reference Standards for	184
		tainty Principle	160		44.0	Nitrogen-15 NMR	104
	8.1.1	Chemical Shift and Exchange Rate			11.2	2.2 Factors Influencing Nitrogen-15	185
	8.1.2	Spin Coupling and Exchange Rates	163			Chemical Shifts	103

ĺΧ

. •	11.2.3 Influence of Protonation and Solven on Nitrogen-15 Chemical Shifts	t 185	15.2	Influence of functional group X on the chemical shift position (δ) of CH_3 , CH_2	
11.3	•	100		and CH protons β to X	212
	Nitrogen-15 NMR	187	15.3	Proton chemical shifts (δ) for CH ₂ and CH	
11.4	Nitrogen-15 Coupling Constants	187		groups bearing more than one functional	
				substituent	212
			15.4	Proton chemical shifts (δ) for protons	1
12	Oxygen-17 NMR Spectra	189		attached to unsaturated and aromatic	
12.1	Oxygen-17 Chemical Shifts	190	15.5	groups	213
12.2	Oxygen-17 Coupling Constants	191	13.3	Influence of functional group X on the chemical shift of protons on benzene rings	
			15.6	Proton chemical shifts (8) for OH, NH and	214
•	•		15.0	SH groups	214
13	NMR Spectra from Other Nuclei	193	15.7	Proton-proton coupling constants	215
13.1	Deuterium NMR Spectra	104		Francisco-F	#1W
13.2		194 195	Carbo	n-13 Data	
13.3	•	195		13C Chemical Shift Summary Chart	.
13.4		195	15.8	13C Chemical shifts (8) in alkane groups	216
13.5	•	196	15.9	13C Chemical shifts (δ) in alkenes	217
13.6	Selenium-77 NMR Spectra	196	15.10		218 219
13.7	Chlorine, Bromine and Iodine NMR Spectra	196	15.11	13C Chemical shifts (δ) in aromatic and	217
13.8	Other Elements and their NMR Spectra	197		heterocyclic molecules	220
			15.12	Influence of functional group X on the	
13				chemical shift of nearby carbons in alkane	:
14 🧝	Background Physics and Biography of NMR	200		chains	221
14.1	Vector Quantities - Torque and Angular		15.13	Brown it on the	
S	Momentum	201		chemical shift of nearby carbons in alkene	.,
14.2	Gyroscopic Behaviour	203	15 14	groups and benzene rings	222
14.3	Magnetic and Electrical Phenomena	204	15.14	¹³ C Chemical shifts (δ) in carbonyl groups ¹³ C Chemical shifts (δ) for carbons in	223
	14.3.1 Bar Magnets and Magnetic Moments	204	13.13	various multiple-bonded environments	-
,	14.3.2 Wire Loops and Spinning Charges	204	15.16	13C Chemical shifts (δ) for methyl groups	223
	14.3.3 Alternating Current	204		in common environments	224
	14.3.4 Signal-to-noise Ratios	205	15.17	¹³ C- ¹ H Coupling constants	224 225
14.4	Quadrupole Moments	206	15.18	¹³ C Relaxation times $-T_1$ (in seconds)	226
14.5	Historical Retrospects in NMR	206			220
14.6	14.5.1 Biographical Notes Nocturnal Processes	207	Other !	Note	
17.0	Noctuinal Processes	209		•	
				Solvents used in NMR work	227
1.0	Tables of Defenses Des	• • •	15.20	Greek Alphabet	227
15	Tables of Reference Data	210	E	w.D 12	
Proto	n Data			r Reading	228
115.1	Proton chemical shifts (δ) for CH ₃ , CH ₂			rs to Problem Examples dix of Magnetic Isotopes	230
	and CH groups attached to functional			ndium of Acronyms in NMR	233
	group X	211	Index	mamin of acronymical PMK	45
					43 T



This chapter presents an overview of the NMR experiment and the chemical information which it can provide; it will not be essential reading for those who have previously met the basics of the technique, but for others it will supply a perspective which would be lacking if the beginnings were treated too rigorously.

1.1 THE PRINCIPLES OF NMR SPECTROSCOPY

The atomic nuclei of many elements are magnetic because they are charged and because they behave as if they were spinning. We can investigate this magnetic property by studying the way in which these nuclei interact with an externally applied magnetic field, B_0 . Typical magnetic nuclei are those of hydrogen (1H), carbon (the ^{13}C isotope only), nitrogen (^{14}N and ^{15}N), oxygen (^{17}O only), fluorine (^{19}F), and phosphorus (^{31}P).

The simplest case is exemplified by the nucleus of

tions which result in two different orientations being allowed — either aligned with the field or opposed to the field. These two orientations clearly have different energies, the aligned position being of lower energy than the opposed.

Whenever an external magnetic field is present, some of the nuclei in the sample become aligned (adopt the lower energy configuration) while others become opposed (adopt the higher energy configuration).

The situation is summarised in figure 1.1.

The energy difference (ΔE) between the two nuclear configurations corresponds to a particular, precise electromagnetic frequency since, by the Bohr relationship, $\Delta E = h\nu$. These energy relationships are illustrated in figure 1.2. To take a specific example, if the external magnetic field strength is 2.35 tesla (T), then the energy gap (ΔE) for the hydrogen nucleus is approximately

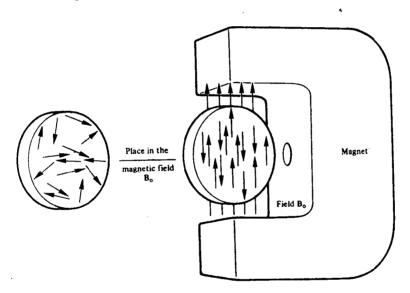


Figure 1.1 In the absence of the field B_0 , the magnetic nuclei in the sample are oriented randomly. In the magnetic field, they must adopt either the aligned Lientation (of lower energy) or the opposed orientation (of higher energy)

hydrogen (the proton) and also by the nucleus of carbon-13 (¹³C). These nuclei behave like bar magnets in an applied field, and in the manner of compass needles tend to align themselves along the same direction as the field. Unlike bar magnets and compass needles however, which always come to rest aligned with the field, magnetic nuclei such as ¹H and ¹³C have quantum restric-

6.6 \times 10⁻²⁶ J, and the corresponding frequency (ν) is 100 MHz (lying in the radiofrequency (RF) band of the electromagnetic spectrum). For ¹³C nuclei in the same magnetic field, the energy gap (ΔE) is 1.7 \times 10⁻²⁶ J, and the frequency ν = 25 MHz.

If a sample containing ¹H nuclei is placed in such a magnetic field, and the sample then irradiated with the

Ł

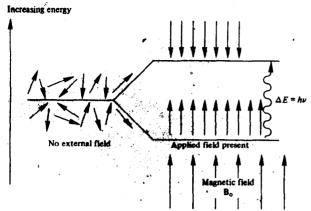


Figure 1.2 In the absence of B_0 the magnetic nuclei all have the same energy. When B_0 is applied, the aligned and opposed orientations correspond to different energies, the energy difference, ΔE , having the dimension $h\nu$

correct radiofrequency, we find that the nuclei interact with the radiofrequency.

Some of the lower-energy nuclei absorb radiation and move up to the higher-energy state: that is, they undergo a transition from being aligned with the field to become opposed to the field. At the same time, some of the higher-energy nuclei are stimulated to emit energy, and they therefore change their opposed orientation and become aligned with the field.

These transitions will only arise when the magnetic energy gap between the nuclear energy levels is matched exactly with the incoming radiofrequency, that is, when they are 'in resonance', and $\Delta E = h\nu$. The process can be pictured as in figure 1.3.

In Table 1.1 are listed a few important nuclei, with the corresponding RF energy necessary to bring about the nuclear magnetic resonance condition and to stimulate transitions among the available magnetic energy levels. (For more comprehensive details see the Appendix at the back of the book.)

The study of nuclear magnetic resonance (NMR) is concerned with these energy levels, and with the frequency of radiation absorbed during resonance.

1.2 THE NMR SPECTROMETER

A primitive lay-out for an NMR instrument is shown in figure 1.4, with a magnetic field strength of 2.35 T. Assuming that the sample contains carbon and hydrogen, then the 13 C nuclei will be partitioned between their two allowed energy levels (1.7 × 10^{-26} J apart); likewise the protons will be partitioned between their two allowed energy levels (6.6 × 10^{-26} J apart).

With the RF transmitter/receiver tuned to 25 MHz, the ¹³C nuclei will undergo upward and downward transitions between the levels and this resonance condition will be detected by observing the absorption of radiofrequency power.

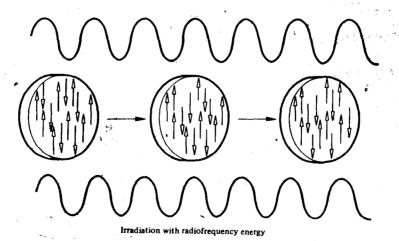


Figure 1.3 When the nuclei (in the magnetic field) are irradiated with radiofrequency energy of the appropriate frequency, some of them undergo transitions from the aligned to the opposed orientations and vice versa

Table 1.1 Common magnetic nuclei. The values shown for ΔE (in joules) and for ν (in MHz, 10^6 s^{-1}) are for nuclei in a magnetic field of 2.35 T

Nucleus	Natural abundance (%)	For $B_0 = 2.35 \text{ T}$			
		$\Delta E/\mathrm{J}$	ν/MHz		
¹H	99.98	6.6 × 10 ⁻²⁶	100		
13C	1.1	1.7×10^{-26}	25		
19F	100	6.2×10^{-26}	94		
31 P	100	2.7×10^{-26}	40.5		
14N	99.63	0.5×10^{-26}	7		
1 5 N	0.37	0.6×10^{-26}	10		
17O	0.037	0.9×10^{-26}	13.5		

Tuned to 100 MHz, the instrument will again record the absorption of RF power, this time because the protons are in resonance and undergoing transitions. The spectrometer will detect ¹⁹F at 94 MHz and ³¹P at 40.5 MHz, and so on.

By convention, the static magnetic field of the instrument is labelled B_0 , and the RF electromagnetic field is labelled B_1 .

1.3 CHEMICAL SHIFTS

Not all protons resonate at exactly 100 MHz in this instrument. Protons attached to carbon atoms for ex-

ample are surrounded by different electron densities compared with those attached to oxygen or nitrogen atoms; this affects their magnetic susceptibilities.

Depending on their chemical environment the precise resonance frequency will be shifted from 100 MHz by a few parts per million; we name this phenomenon *chemical shift*, and it is the most important reason for the development of NMR as a routine and powerful analytical technique in chemistry.

For a simple organic molecule like *tert*-butyl alcohol (see figure 1.5) there are protons in two different chemical environments — nine of them are in methyl groups and one is attached to oxygen. The nine methyl protons come to resonance about 130 Hz higher in frequency than 100 MHz (that is, about 1.3 parts per million to higher frequency) and the OH proton absorbs at about 400 Hz (about 4 ppm) higher frequency than 100 MHz.

Figure 1.5 is the proton magnetic resonance spectrum (¹H NMR spectrum) for *tert*-butyl alcohol; it is essentially a plot of RF absorption against frequency.

Note that the integrated area of the larger peak (for the methyl protons) is nine times that of the smaller (OH) signal; in other words

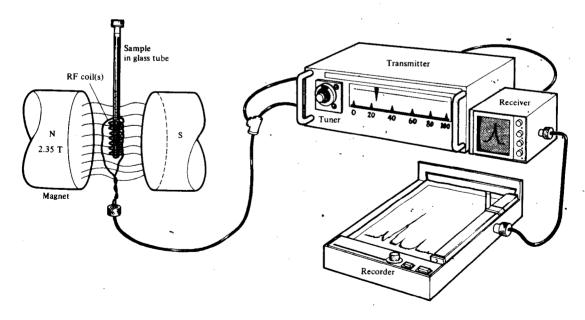


Figure 1.4 Nuclear magnetic resonance spectrometer. The sample is placed in the magnetic field; when the radio-frequency transmitter is tuned to the resonance frequency, a signal arises in the receiver

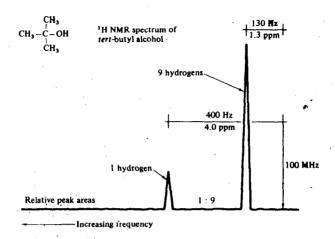


Figure 1.5 Chemical shift. In the proton magnetic resonance spectrum (¹H NMR spectrum) of tert-butyl alcohol, the nine methyl protons give a signal 1.3 ppm higher in frequency than 100 MHz. The hydroxyl proton signal is about 4.0 ppm higher than 100 MHz. The field strength for this 100 MHz proton NMR spectrum is 2.35 T

Signal intensity is proportional to the number of protons present in each of the chemical environments within the molecule.

Figure 1.6 is the carbon-13 NMR spectrum for tertbutyl alcohol. Again, since there are only two different kinds of carbon atom in the molecule (the three CH₃ carbons and the C-OH carbon) each comes to resonance, not at 25 MHz, but chemically shifted so that the CH₃ carbons give a signal about 750 Hz higher in frequency, and the C-OH carbon signal appears around 1750 Hz higher in frequency. Note again that the signals are of different intensity; for reasons that we shall meet later, it is not as easy in ¹³C NMR to treat these intensities quantitatively.

The carbon-12 isotopes in the sample of tert-butyl alcohol make no contribution to this spectrum since they are non-magnetic; the fact that only 1.1 per cent of the carbon atoms are ¹³C (see table 1.1) makes the carbon-13 NMR spectrum more difficult to record than the proton spectrum. A glance at table 1.1 shows that

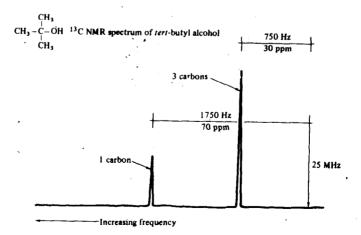


Figure 1.6 Chemical shift. In the routine carbon-13 NMR spectrum of tert-butyl alcohol, the three methyl carbons give a signal 30 ppm higher in frequency than 25 MHz. The C-O signal is 70 ppm higher than 25 MHz.

The field strength for this 25 MHz ¹³C NMR spectrum is 2.35 T

¹⁵N spectra are even more difficult to obtain, with a natural abundance of only 0.37 per cent, and that ¹⁷O is exceedingly rare; ¹⁹F and ³¹P NMR spectra in contrast are relatively easily recorded.

Worked Example 1.1

For the molecule of methyl acetate (methyl ethanoate) CH₃COOCH₃, (a) state how many different chemical shift environments exist for the hydrogen, carbon and oxygen nuclei, (b) note from table 1.1 the natural abundance of each principal magnetic isotope and (c) state the frequency at which each NMR spectrum would be recorded in a spectrometer with a 2.35 T magnet.

Answer 1.1

There are two chemically distinctive hydrogen environments: the natural abundance of the most important hydrogen isotope (¹H) is 99.98 per cent, and the spectrum would be recorded at 100 MHz and 2.35 T. There are three carbon environments: carbon-13 abundance is only 1.1 per cent, and the required frequency (for 2.35 T) is 25 MHz. (Carbon-12 is non-magnetic and does not exhibit NMR.) There are also two oxygen environments; oxygen-17 NMR at natural abundance is handicapped by the very low natural abundance (0.037 per cent) but oxygen-17 NMR spectra are acquired at 13.5 MHz in a 2.35 T instrument. (Oxygen-16 is non-magnetic and does not exhibit NMR.)

Problem Example 1.1

Formula 1.1 is an imaginary 'isotope isomer' or isotopomer of acetamide. In a 2.35 T magnet, state the frequencies at which the NMR spectrum of each element would be recorded. For each spectrum, state how many signals (that is, chemical shift positions) would be seen. (Detail: because of restricted rotation around the C-N bond, consider the protons on nitrogen to be in different environments, one being nearer to oxygen than the other.)

$$^{17}O$$
 $^{13}CH_3 - ^{13}C$
 $^{15}N - ^{1}H$
 ^{1}H
 ^{1}H
 ^{17}O
 ^{11}O
 $^$

1.4 FIELD STRENGTH AND FREQUENCY

Figure 1.2 and table 1.1 show that magnetic nuclei can occupy various magnetic sublevels, and that the frequency necessary to cause nuclear transitions is different for each element or isotope. This resonance frequency is found to vary in direct proportion to the applied field (for all magnetic nuclei); thus the larger the magnetic field the higher the frequency necessary to achieve resonance. That is

For the proton we can represent this fact as in figure 1.7.

Although the arithmetic of these field-frequency relationships is simple, a few examples will help to emphasise their crucial importance.

Worked Example 1.2

What is the frequency needed to induce transitions between the ¹³C nuclear energy levels, when the field strength is (a) 4.7 T and (b) 1.88 T?

Answer 1.2

Since from table 1.1 the ¹³C NMR frequency is 25 MHz at 2.35 T, then by simple proportion it is 50 MHz at 4.7 T and 20 MHz at 1.88 T.

Problem Example 1.2

What field strength is necessary in an instrument designed for studying proton NMR at (a) 60 MHz, (b) 200 MHz, (c) 600 MHz?

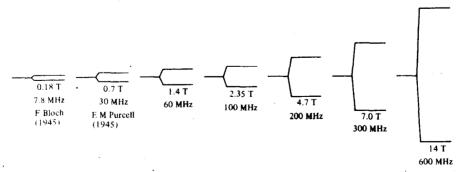


Figure 1.7 Field-field relationship. The energy gaps ($\Delta E = h\nu$) are shown for protons in different field strengths. For other elements the same simple proportion holds; table 1.1 lists frequencies for a field strength of 2.35 T