



**Introduction to**

# **Mammalian**

# **Reproduction**

edited by  
Daulat Tulsiani

---

# INTRODUCTION TO MAMMALIAN REPRODUCTION

*edited by*

**Daulat Tulsiani, Ph.D.**  
*Vanderbilt University School of Medicine*  
*Nashville, Tennessee*



**KLUWER ACADEMIC PUBLISHERS**  
**Boston / Dordrecht / London**

---

**Distributors for North, Central and South America:**

Kluwer Academic Publishers  
101 Philip Drive  
Assinippi Park  
Norwell, Massachusetts 02061 USA  
Telephone (781) 871-6600  
Fax (781) 681-9045  
E-Mail: [kluwer@wkap.com](mailto:kluwer@wkap.com)

**Distributors for all other countries:**

Kluwer Academic Publishers Group  
Post Office Box 322  
3300 AH Dordrecht, THE NETHERLANDS  
Telephone 31 786 576 000  
Fax 31 786 576 254  
E-Mail: [services@wkap.nl](mailto:services@wkap.nl)



Electronic Services < <http://www.wkap.nl>>

---

**Library of Congress Cataloging-in-Publication Data**

A C.I.P. Catalogue record for this book is available  
from the Library of Congress.

Introduction to Mammalian Reproduction edited by Daulat Tulsiani  
ISBN 1-4020-7283-X

---

**Copyright** © 2003 by Kluwer Academic Publishers

All rights reserved. No part of this work may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, microfilming, recording, or otherwise, without the written permission from the Publisher, with the exception of any material supplied specifically for the purpose of being entered and executed on a computer system, for exclusive use by the purchaser of the work.

Permission for books published in Europe: [permissions@wkap.nl](mailto:permissions@wkap.nl)

Permissions for books published in the United States of America: [permissions@wkap.com](mailto:permissions@wkap.com)

*Printed on acid-free paper.*

Printed in the United States of America.

***The Publisher offers discounts on this book for course use and bulk purchases.  
For further information, send email to <[joanne.tracy@wkap.com](mailto:joanne.tracy@wkap.com)> .***

## CONTRIBUTORS

Aida Abou-Haila, D.Sc.  
UFR Biomedicale  
Universite Rene Descartes Paris V  
45, rue des Saints-Peres  
F-75270 Paris, Cedex 6  
France  
[aida.abou-haila@biomedicale.univ-paris5.fr](mailto:aida.abou-haila@biomedicale.univ-paris5.fr)

Yoshihiko Araki, M.D., D.Med.  
Sci  
Department of Immunology and  
Parasitology  
Yamagata University School of  
Medicine  
2-2-2 Iida-Nishi  
Yamagata City 990-9585  
Japan  
[yaraki@med.id.yamagata-u.ac.jp](mailto:yaraki@med.id.yamagata-u.ac.jp)

Gail A. Cornwall, Ph.D.  
Departments of Cell Biology &  
Biochemistry  
Texas Tech. University Health  
Science Center  
3601 4<sup>th</sup> Street  
Lubbock, TX 79430  
USA  
[gail.cornwall@ttmc.ttuhsu.edu](mailto:gail.cornwall@ttmc.ttuhsu.edu)

Benjamin J. Danzo, Ph.D.  
Department of Obstetrics &  
Gynecology  
Vanderbilt University School of  
Medicine  
Room B-1100 MCN  
Nashville, TN 37232-2519  
USA  
[ben.danzo@vanderbilt.edu](mailto:ben.danzo@vanderbilt.edu)

M. Deng, Ph.D.  
Department of Animal Science  
University of Connecticut  
3636 Horsebarn Road Ext U-40  
Storrs, CT 06269  
USA

Luis Dettin, Ph.D.  
Department of Cell Biology  
Georgetown University Medical  
Center  
SE 216 Medical-Dental Building  
3900 Reservoir Road, NW  
Washington, DC 20007  
USA

Alan B. Diekmann, Ph.D.  
Ctr. for Research in Contraceptive  
and Reproductive Health  
Department of Cell Biology  
Univ. of Virginia Health System  
Box 800732  
Charlottesville, VA 22908-0732  
USA

Bonnie S. Dunbar, Ph.D.  
Department of Cell Biology  
Baylor College of Medicine  
One Baylor Plaza  
Houston, TX 77030  
USA  
[bdunbar@bcm.tmc.edu](mailto:bdunbar@bcm.tmc.edu)

Martin Dym, Ph.D.  
Department of Cell Biology  
Georgetown Univ. Medical Ctr.  
3900 Reservoir Road NW  
Washington, DC 20007  
USA  
[dymm@gunet.georgetown.edu](mailto:dymm@gunet.georgetown.edu)

Janice P. Evans, Ph.D.  
 Department of Biochemistry &  
 Molecular Biology  
 Division of Reproductive Biology  
 Johns Hopkins University  
 Bloomberg School of Public  
 Health  
 615 N. Wolfe St., Room 3606  
 Baltimore, MD 21205  
 USA  
[jpevans@jhsp.h.edu](mailto:jpevans@jhsp.h.edu)

Asgerally T. Fazleabas, Ph.D.  
 Department of Obstetrics &  
 Gynecology  
 University of Illinois at Chicago  
 820 South Wood Street  
 Chicago, IL 60612-7313  
 USA  
[asgi@uic.edu](mailto:asgi@uic.edu)

Chhanda Gupta, Ph.D.  
 Department of Pediatrics  
 University of Pittsburgh  
 Pittsburgh, PA 15213  
 USA  
[gchhanda@hotmail.com](mailto:gchhanda@hotmail.com)

John C. Herr, Ph.D.  
 Ctr. for Research in Contraceptive  
 and Reproductive Health  
 Department of Cell Biology  
 Univ. of Virginia Health System  
 Charlottesville, VA 22908-0732  
 USA

Barry T. Hinton, Ph.D.  
 Department of Cell Biology  
 Univ. of Virginia Health System  
 School of Medicine  
 P.O. Box 800732  
 Charlottesville, VA 22908-0732  
 USA  
[bth7c@virginia.edu](mailto:bth7c@virginia.edu)

Gautam Kaul, Ph.D.  
 Senior Scientist  
 Department of Biochemistry  
 National Dairy Research Institute  
 Karnal 132001  
 Haryana, India

Firyal S. Khan-Dawood, Ph.D.  
 Department of Pathology  
 Morehouse School of Medicine  
 720 West View Drive SE  
 Atlanta, GA 30310 USA  
[fkhan@msm.edu](mailto:fkhan@msm.edu)

Gary Killian, Ph.D.  
 Dairy Breeding Research Center  
 Penn State University  
 University Park, PA 16802  
 USA  
[lwj@psu.edu](mailto:lwj@psu.edu)

Lin Liu, Ph.D.  
 Department of Animal Science  
 University of Connecticut  
 3636 Horsebarn Road Ext U-40  
 Storrs, CT 06269  
 USA  
[lliu@canr.uconn.edu](mailto:lliu@canr.uconn.edu)

Christoph R. Loeser, M.D.  
 Center for Dermatology and  
 Andrology  
 Justus-Liebig-University  
 Gaffkystr. 14  
 35385 Giessen  
 Germany  
[christoph.loeser@derma.med.uni-giessen.de](mailto:christoph.loeser@derma.med.uni-giessen.de)

Ben M.J. Pereira, Ph.D.  
 Department of Biosciences &  
 Biotechnology  
 Indian Institute of Technology  
 Roorkee  
 Roorkee-247 667,  
 India  
[benmjfbs@rurkiu.ernet.in](mailto:benmjfbs@rurkiu.ernet.in)

Ramasare Prasad, Ph.D.  
 Department of Biosciences &  
 Biotechnology  
 Indian Institute of Technology  
 Roorkee  
 Roorkee-247 667  
 India

Sarvamangala V. Prasad, Ph.D.  
 Department of Cell Biology  
 Baylor College of Medicine  
 One Baylor Plaza  
 Houston, TX 77030  
 USA  
[prasad@bcm.tmc.edu](mailto:prasad@bcm.tmc.edu)

Parul Pruthi, Ph.D.  
 Department of Biosciences &  
 Biotechnology  
 Indian Institute of Technology  
 Roorkee  
 Roorkee-247 667  
 India

Neelakanta Ravidranath, Ph.D.  
 Department of Cell Biology  
 Georgetown Univ. Medical Center  
 SE216 Medical-Dental Building  
 3900 Reservoir Road, NW  
 Washington, DC 20007  
 USA  
[ravindr@gunet.georgetown.edu](mailto:ravindr@gunet.georgetown.edu)

Carmen M. Rodriguez, Ph.D.  
 Department of Cell Biology  
 Univ. of Virginia Health System  
 School of Medicine  
 P. O. Box 800732  
 Charlottesville, VA 22908-0732  
 USA  
[cmr4z@virginia.edu](mailto:cmr4z@virginia.edu)

Wolf-Bernhard Schill, M.D.  
 Center for Dermatology and  
 Andrology  
 Justus-Liebig-University  
 Gaffkystr. 14  
 35385 Giessen  
 Germany

Hans-Christian Schuppe, M.D.  
 Center for Dermatology and  
 Andrology  
 Justus-Liebig-University  
 Gaffkystr. 14  
 35385 Giessen  
 Germany

Thomas Stalf, Ph.D.  
 Institute of Reproductive  
 Medicine  
 Frankfurter Str. 52  
 35392 Giessen  
 Germany

Susan S. Suarez, M.S., Ph.D.  
 Department of Biomedical  
 Sciences  
 College of Veterinary Medicine  
 Cornell University  
 T5-006 Veterinary Research  
 Tower  
 Ithaca, NY 14853  
 USA  
[sss7@cornell.edu](mailto:sss7@cornell.edu)

X. C. Tian, Ph.D.  
Department of Animal Science  
University of Connecticut  
3636 Horsebarn Road Ext U-40  
Storrs, CT 06269  
USA

Kiyotaka Toshimori, M.D.  
Miyazaki Medical College  
Kihara 5200, Kiyotake  
Miyazaki, 889-1692  
Japan  
[ktoshi@post.miyazaki-med.ac.jp](mailto:ktoshi@post.miyazaki-med.ac.jp)

Daulat R.P. Tulsiani  
Departments of Obstetrics &  
Gynecology  
and Cell Biology  
Vanderbilt University School of  
Medicine  
Room D-3243 MCN  
Nashville, TN 37232-2633  
[daulat.tulsiani@vanderbilt.edu](mailto:daulat.tulsiani@vanderbilt.edu)

Srinivasan Vijayaraghvan, Ph.D.  
Department of Biological Sciences  
Kent State University  
Kent, OH 44242  
USA  
[svijayar@kent.edu](mailto:svijayar@kent.edu)

Pablo E. Visconti, Ph.D.  
Center for Research in  
Contraceptive  
and Reproductive Health  
Department of Cell Biology  
Univ. of Virginia Health System  
Charlottesville, VA 22908-0732  
USA  
[pv6j@virginia.edu](mailto:pv6j@virginia.edu)

V. Anne Westbrook, Ph.D.  
Center for Research in  
Contraceptive  
and Reproductive Health  
Department of Cell Biology  
Univ. of Virginia Health System  
Box 800732  
Charlottesville, VA 22908-0732  
USA  
[aw2p@virginia.edu](mailto:aw2p@virginia.edu)

Xiangzhong Yang, Ph.D.  
Department of Animal Science  
University of Connecticut  
3636 Horsebarn Road Ext U-40  
Storrs, CT 06269  
USA  
[tyang@canr.uconn.edu](mailto:tyang@canr.uconn.edu)

<sup>1</sup>Email address of corresponding  
authors

## PREFACE

One of the goals of reproductive (gamete) biologists is to understand the biochemical processes and molecular mechanisms that regulate the formation and maturation of male and female gametes, and their ultimate union to form a zygote, a cell with somatic chromosome numbers. Development of the zygote begins immediately after sperm and egg haploid pronuclei come together, pooling their chromosomes to form a single diploid nucleus with the parental genes. The major difference between the reproductive and non-reproductive processes is that many events including interaction of the opposite gametes are species specific, and the knowledge gained in a given species may be applicable only in a few closely related species. Thus, the progress in understanding many aspects of gamete biology have been painfully slow. Despite slow advancement, many fascinating discoveries have been made. Recent successes of *in vitro* fertilization (IVF), and intracytoplasmic sperm injection (ICSI) techniques are noteworthy, and have helped many couples experience joy of parenthood. The assisted reproductive procedures are now being routinely used to increase the numbers of farm animals and endangered species. Many of these advances, in conjunction with recent successes in the cloning of laboratory and farm animals, were some of the factors behind my decision to undertake the task of organizing this book on mammalian reproduction.

So far as I know, there is no other book that systematically describes the formation and maturation of male and female gametes, and factors that regulate their union during the fertilization process, activation and implantation of fertilized egg, manipulation of the gametes for assisted reproduction, and environmental toxicants. That such book was needed became apparent to me when teaching a course on reproduction to the graduate and medical students at the Vanderbilt School of Medicine. Every attempt has been made to include a wide spectrum of topics (chapters) on morphological and physiological aspects of male and female gametes. These chapters are contributed by investigators currently engaged in "cutting-edge" research in the area of reproductive biology. Needless to say, I am very grateful to all the contributors, whose expertise, willingness to contribute, and hard work have made this book possible. My sincere hope is that the book will succeed in giving pertinent information to most of its readers, which are likely to include undergraduate, graduate and medical students, and perhaps their mentors. If my attempts generate a reasonable interest and stimulate a few young minds to expect exciting possibilities in the area of gamete biology, this book has fulfilled its purpose.

Daulat R.P. Tulsiani, Ph.D.



## ACKNOWLEDGEMENTS

I was born in Village Gucherow of the District of Karachi in former British India and moved with my family to the independent India during the partition in 1947. I must acknowledge the help of many kind souls who assisted my refugee family to get settled in India. I am grateful to all my teachers in India for their continuous help and encouragement.

My first encounter in the U.S.A. came when I joined the laboratory of Professor Raul Carubelli in the Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma, in 1968 as a postdoctoral fellow. I greatly benefited from the advice and encouragement of my mentor and his colleagues during my young years in Oklahoma.

I joined the research team of Professor and Chairman Oscar Touster at Vanderbilt University, Nashville, in 1972. The university provided a rich academic environment for my professional growth. It is not possible to list the names of all the colleagues, research fellows, and students who have been a continuing inspiration during my stay at Vanderbilt. However, I must acknowledge the following colleagues for their collaborations and discussions throughout my tenure in the area of reproductive biology: Drs. Marjorie D. Skudlarek, Marie-Claire Orgebin-Crist, Benjamin J. Danzo, and Michael K. Holland. I am grateful to Professor Stephen S. Entman, Chairman of the Department of Obstetrics & Gynecology, and Vanderbilt University for providing me with the space and facilities for editing this book.

My sincere thanks to the contributors who graciously sent their assigned chapters in a timely manner. Many chapters needed very little editing; however, there were some that needed extensive editing and formatting. I learned more by editing these chapters than from any other text book or research article.

I am deeply indebted to Loreita Little and Lynne Black for editorial assistance, and to Lynne Black for final preparation of the chapters for the camera-ready format. Without this assistance, the publication of this book would have been considerably delayed. Finally, I am grateful to Joanne Tracy, Editor of Biosciences, at the Kluwer Academic Publishers, for her faith in this project. The research in my laboratory is supported in part by research grants HD25869 and HD34041 from the National Institute of Child Health and Human Development.

# TABLE OF CONTENTS

<b>List of Contributors .....</b>	<b>ix</b>
<b>Preface.....</b>	<b>xiii</b>
<b>Acknowledgements .....</b>	<b>xv</b>

## Part 1. Male Gamete

1. Mammalian Testes: Structure and Function Neelakanta Ravidranath, Luis Dettin, and Martin Dym .....	1
2. The Sperm Acrosome: Formation and Contents Aida Abou-Haila and Daulat R.P. Tulsiani .....	21
3. Ductus Epididymis Gail A. Cornwall.....	41
4. The Testicular and Epididymal Luminal Fluid Microenvironment Carmen M. Rodriguez and Barry T. Hinton .....	61
5. Sperm Motility: Patterns and Regulation Srinivasan Vijayaraghvan.....	79
6. Testicular and Epididymal Maturation of Mammalian Spermatozoa Kiyotaka Toshimori.....	93
7. Glycosyl Phosphatidyl Inositol (GPI) Anchored Molecules on Mammalian Spermatozoa Ben M.J. Pereira, Parul Pruthi, and Ramasare Prasad .....	113
8. Male Accessory Glands: Molecular Mechanisms of Development Chhanda Gupta .....	127

## Part II. Female Gamete

9. Formation and Structure of Mammalian Ovaries Yoshihiko Araki .....	141
--	-----

10. The Ovarian Cycle  
Firyal S. Khan-Dawood ..... 155
11. Estrogen-Associated Glycoproteins in Oviduct Secretions: Structure and Evidence for a Role in Fertilization  
Gary Killian ..... 187
12. Structure and Function of Mammalian Zonae Pellucidae  
Sarvamangala V. Prasad, Gautam Kaul, and Bonnie S. Dunbar ..... 203

### **Part III. Early Events of Fertilization**

13. Transport of Spermatozoa in the Female Genital Tract  
Susan S. Suarez..... 227
14. Capacitation: Signaling Pathways Involved in Sperm Acquisition of Fertilizing Capacity ..... 237  
V. Anne Westbrook, Alan B. Diekman, John C. Herr, and Pablo E. Visconti
15. Sperm-Egg Interaction and Exocytosis of Acrosomal Contents  
Daulat R.P. Tulsiani and Aida Abou-Haila ..... 257

### **Part IV. Fusion of Gametes**

16. Gamete Membrane Interactions. The Cell-Cell Interactions between Sperm and Egg during Fertilization  
Janice P. Evans ..... 289
17. Activation of Mammalian Oocytes: Principles and Practice  
L. Liu, M. Deng, X.C. Tian, and X. Yang..... 319
18. Implantation: Lessons from A Primate Model  
Asgerally T. Fazleabas ..... 347

### **Part V. Medical Implications**

19. Assisted Reproduction: Techniques and Participants  
Christoph R. Loeser, Thomas Stalf, Hans-Christian Schuppe, and Wolf-Bernhard Schill ..... 357

20. The Reproductive Effects of Hormonally Active Environmental Agents Benjamin J. Danzo.....	377
--	-----

<b>Subject Index .....</b>	<b>401</b>
----------------------------	------------

## Chapter 1

# **MAMMALIAN TESTES: STRUCTURE AND FUNCTION**

Neelakanta Ravindranath, Luis Dettin, and Martin Dym  
*Georgetown University School of Medicine, Washington, DC, USA*

## **INTRODUCTION**

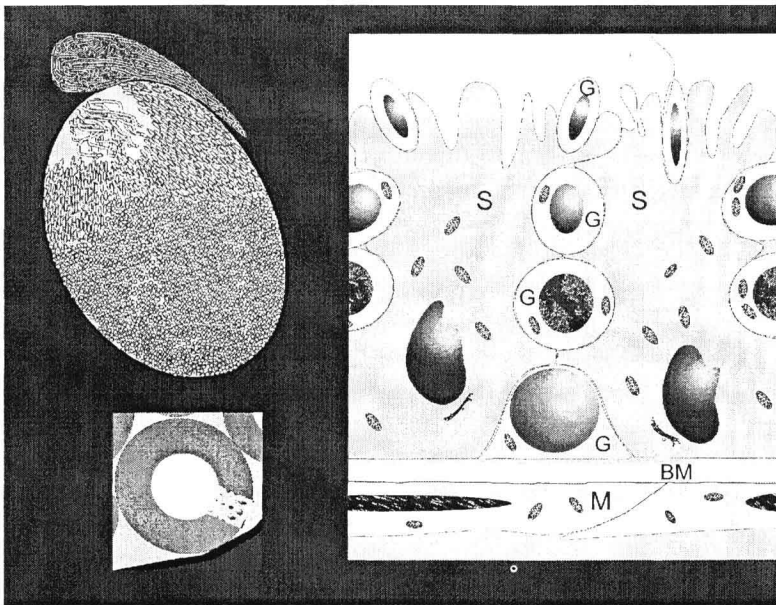
The male reproductive system consists of the primary sex organs, the two testes and a set of accessory sexual structures. The adult mammalian testis performs two important functions, spermatogenesis and male sex hormone production. It is an organ structurally designed to produce the haploid male gametes from diploid postnatal germ-line stem cells, i.e. type A spermatogonia. The process of morphological and functional differentiation of type A spermatogonia into the haploid male gamete, the spermatozoon, is termed spermatogenesis. In addition, the testis elaborates a steroid hormone, testosterone, that is responsible for maintaining the spermatogenic process as well as the secondary male sexual characteristics. Furthermore, testosterone is important for several different functions in various organ systems including the maintenance of muscle mass and bone density. The process of testosterone formation from its precursor, cholesterol, is termed steroidogenesis. In this chapter, we will discuss how the structure and form of the testis contributes to the processes of spermatogenesis and steroidogenesis.

## **MORPHOLOGY OF THE ADULT TESTIS**

Each testis is covered with a thick fibrous capsule, the tunica albuginea. The thick infolding of the tunica albuginea at the posterior margin of the testis forms the mediastinum of the testis. Connective tissue septae originate from the mediastinum and pass into the interior of the testis, and subdivide it into several lobules. Within these lobules lie the convoluted folds of the seminiferous tubule. The space surrounding the seminiferous folds is occupied by the interstitial tissue. The seminiferous tubules form coiled

loops that terminate at both ends into the rete testes located within the mediastinum. Spermatozoa and testicular fluid produced within the seminiferous tubule pass through the rete testes into the ductuli efferentes and epididymis.

Histologically, the adult testis can be divided into two compartments, a seminiferous tubular compartment and an interstitial compartment (Fig. 1). The tubular compartment consists of an outer layer (s) of peritubular myoid cells and an inner layer of seminiferous epithelium separated by an intermediate layer of acellular matrix or basement membrane. The interstitial compartment consists of Leydig cells, immune cells (macrophages and lymphocytes), and fibroblasts. In addition, it also contains blood and lymph vessels, nerves, and loose connective tissue. The tubular and interstitial compartments of the testis perform the defined functions of spermatogenesis and steroidogenesis, respectively.



*Figure 1.* Schematic representation of a mammalian testis (top left), a cross section of a seminiferous tubule (bottom left), and the seminiferous epithelium (right) showing myoid cells (M), the basement membrane (BM), Sertoli cells (S), and germ cells (G).

## SEMINIFEROUS EPITHELIUM AND SPERMATOGENESIS

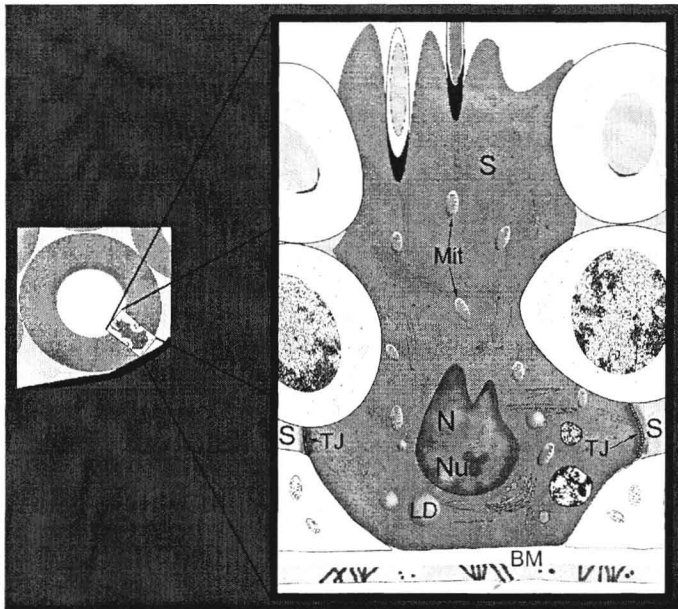
The seminiferous epithelium rests on the acellular basement membrane and contains two types of cells, Sertoli cells and germ cells (Fig. 1). At the

time of birth, the seminiferous epithelium consists of Sertoli cells and only one type of germ cell, i.e., the gonocyte, located in the central part of the seminiferous cord. Gonocytes migrate to the basement membrane during the early postnatal period and are now called type A spermatogonia. Type A spermatogonia could be called 'male germ-line stem cells' as they renew themselves and also differentiate into spermatozoa (1). During the process of differentiation into spermatozoa, the type A spermatogonia undergo several mitotic divisions to yield type B spermatogonia. Type B spermatogonia mitotically divide to yield primary spermatocytes. Primary spermatocyte through two successive meiotic divisions form haploid spermatids. The haploid spermatids morphologically differentiate into spermatozoa. Thus, in the adult testis, the seminiferous epithelium consists of various germ cell types with the stem cells resting on the basement membrane and more differentiated germ cell types arranged progressively towards the lumen. The germ cells at different stages of differentiation are in close anatomical and functional contact with the Sertoli cells. However, tight junctional complexes between adjoining Sertoli cells compartmentalize the seminiferous epithelium into a basal compartment and an adluminal compartment (2). The junctional complexes separate young germ cells, i.e. spermatogonia and the preleptotene spermatocytes, from later spermatocytes, spermatids, and spermatozoa. In addition, they form the morphological basis of blood-testis barrier. This barrier creates a unique microenvironment in the adluminal compartment. The germ cells in the basal compartment communicate with the neighbouring Sertoli cells, the basement membrane, the peritubular myoid cells, and the blood and lymphatic vessels. More advanced germ cells in the adluminal compartment derive substances in blood or lymph through the Sertoli cell (3). Thus, Sertoli cells interact with all types of germ cells via desmosomes and gap junctions (4, 5). In addition, Sertoli cells develop ectoplasmic specializations (actin-rich filaments sandwiched between plasma membrane and endoplasmic reticulum) and tubulobulbar complexes with spermatids (6). The development and degradation of these structural complexes between neighbouring Sertoli cells at the base of the seminiferous epithelium and between elongating spermatids and the Sertoli cell at the apical end of the seminiferous epithelium has been correlated with the movement of spermatocytes from basal to adluminal compartment and the release of sperm to the lumen, respectively (6).

## Sertoli Cell

Generally, the Sertoli cells exhibit an infolded nuclear envelope with pores, a homogeneous nucleoplasm, and a single tripartite nucleolus (Fig. 2). Within the cytoplasm of Sertoli cells, a large Golgi apparatus, numerous mitochondria, lysosomes, multivesicular bodies, lipid droplets, and residual

bodies have been described. Sertoli cells present a profuse network of both rough and smooth endoplasmic reticulum suggesting their capability for both protein and steroid synthesis and secretion. Sertoli cells lack secretory granules, large vacuoles, and exocytotic vesicles (7, 8). Lack of these structures indicate that the synthesized proteins may be transferred to the plasma membrane where they are either secreted after cleavage or remain membrane-bound for interaction with the corresponding receptor on germ cell types. A classic example of the growth factor that is expressed by Sertoli cells in both membrane-bound and secretory form is stem cell factor (9). The corresponding receptor, c-kit, is expressed on the surface of type A spermatogonia (10, 11). This concept appears to be true as Sertoli cells have been shown to extend cytoplasmic processes (conical at the base, sheet-like in the middle, and tapered apical towards the lumen) that interact with the plasma membranes of spermatogonia, spermatocytes, spermatids, and spermatozoa (12). The shape of the Sertoli cells in 3-dimension



*Figure 2.* A schematic representation of a Sertoli cell. Morphological details of the Sertoli cell is shown in the magnified image of the portion of the seminiferous tubule from the drawing on the left. Note that the Sertoli cell (S) is placed perpendicular to the basement membrane (BM). It possesses an infolded nucleus (N) with cytoplasm containing numerous mitochondria (Mit) and lipid droplets (LD). A tripartite nucleolus (Nu) is apparent within the nucleus. A tight junction (TJ) between adjoining Sertoli cells is also shown.

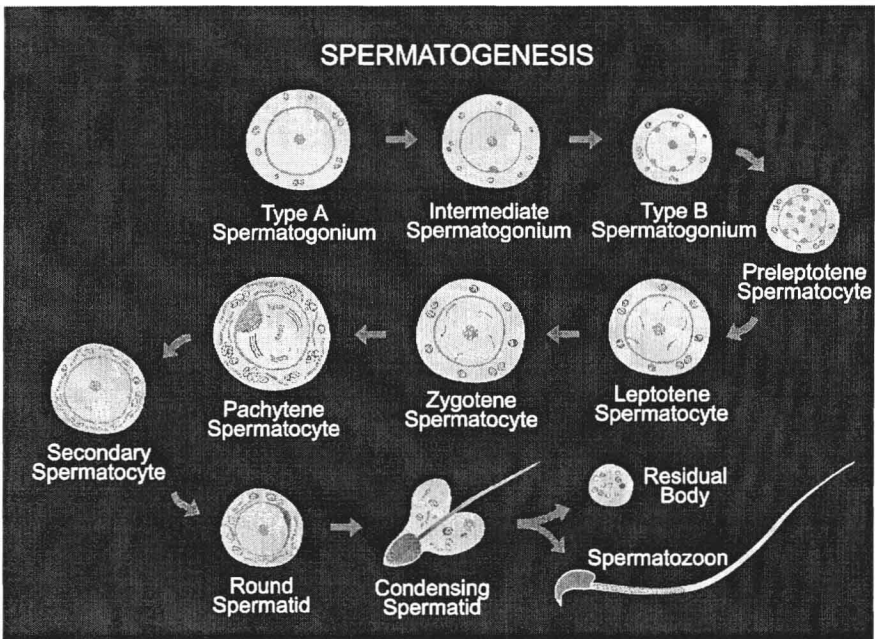
changes continuously to accommodate the developing and differentiating germ cells and their mobilization from the base to the lumen (13). Apart from the above mentioned structures, Sertoli cells possess elaborate



cytoskeleton consisting of microtubules and filaments that may be involved in transport of spermatids through the seminiferous epithelium (6).

## Germ Cells

In the seminiferous epithelium of the adult testis where spermatogenesis is progressing actively, germ cell types beginning with the most primitive germ cell, i.e., type A spermatogonia, to the most differentiated type, i.e., spermatozoa, are observed. The intermediary cell types during this differentiation pathway are type B spermatogonia, preleptotene spermatocytes, spermatocytes in different phases prior to meiotic division (leptotene, zygotene, and pachytene), secondary spermatocytes, and spermatids (round and elongating). A schematic representation of the stages of differentiation of type A spermatogonia into spermatozoa is shown in Fig. 3.



*Figure 3.* A schematic representation of the process of spermatogenesis. Type A spermatogonia that are present at the base of the seminiferous epithelium undergo a series of mitotic divisions to yield intermediate and type B spermatogonia. Further mitotic divisions of type B result in the formation of preleptotene spermatocytes. The preleptotene spermatocytes through leptotene, zygotene, and pachytene stages undergo the first meiotic division. The resultant secondary spermatocytes proceed through the second meiotic division to yield round spermatids. Round spermatids morphologically differentiate into spermatozoa.