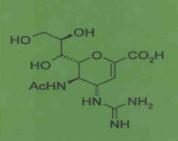
# PRACTICAL METHODS FOR

# BIOCATALYSIS AND BIOTRANSFORMATIONS



Editors JOHN WHITTALL | PETER W. SUTTON



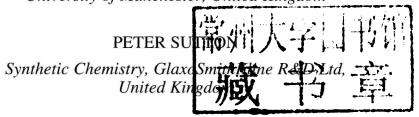


# Practical Methods for Biocatalysis and Biotransformations

### **Editors**

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# Practical Methods for Biocatalysis and Biotransformations

# **Preface**

During the early to mid 1990s Professor Stan Roberts was chief editor of a series of loose-leaf laboratory protocols detailing the use of biotransformations in synthetic organic chemistry that were collected together and published in book form (*Preparative Biotransformations*, Wiley, Chichester, 1999). This led to the publication of the series of books *Catalysts for Fine Chemical Synthesis*, volumes 1–5, by the same publisher which covered the application of chemo- and bio-catalytic procedures for the synthesis of fine chemicals; for this series, Dr John Whittall became co-editor on the homogeneous catalysis volumes. Following the format of this series, *Practical Methods in Biocatalysis and Biotransformations* has been prepared. In keeping with these earlier formats, we aim to provide the readership with enough information to understand when a biocatalytic or biotransformation method would be a suitable practical method to carry out their synthetic transformation.

In recent times, the employment of enzymes and whole cells to perform a range of organic reactions has become much more commonplace, and biotransformation has become accepted as a powerful method for application in synthetic organic chemistry. However, for chemists developing synthetic methods for a particular target molecule, the understanding of the advantages and limitations of biocatalysis and biotransformation is not always clear. Therefore, this book intends to review the industrial background to when biotransformations are used and introduce the nonmicrobiologist to the background of how biocatalysts are discovered and developed and then give detailed experimental procedures for a comprehensive range of useful biotransformation methods.

In order to place the later chapters in proper context, Chapter 1 offers a comprehensive review of biotransformation from the perspective of a large pharmaceutical company (GSK) and Chapter 2 gives an introduction that allows an appreciation of molecular biology for scientists with no formal training in this area.

In the remaining chapters, key biotransformations have been identified from the recent primary literature (learned journals) and the respective authors have amplified the disclosure of their methodologies in this volume. These disclosures often contain additional equipment and experimental details to those found in the experimental section of most journals, allowing the reader to decide whether these methods are suitable for addressing their needs.

Chapter 3 describes the application of lipases, proteases and sulfatases for the kinetic resolution of a range of interesting molecules. A selection of dynamic kinetic resolution (DKR) procedures is disclosed in Chapter 4. DKRs are attracting a significant amount of

interest as they allow access to >50 % yields of single enantiopure products from racemates. Other useful synthetic applications of hydrolase enzymes are covered in Chapter 5, including desymmetrization and regio- and chemo-selective transformations.

Chapters 6 and 7 cover sugar-type chemistry, focusing on aldol and glycosylation methods which can offer substantial advantages over traditional chemical approaches.

Chapter 8 describes the application of hydroxyl nitrile lyases to the synthesis of new chiral cyanohydrins and  $\alpha$ -hydroxy acids and includes new approaches to the transformation of 'difficult' aldehyde and ketone substrates using substrate engineering and immobilization techniques.

The latter part of the book is dedicated to redox biotransformation application, with Chapter 9 disclosing several methods for the synthesis of chiral secondary alcohols using a range of commercially available ketoreductases (alcohol dehydrogenases) which are now being applied regularly on a large scale.

Chapter 10 covers reductive enzymes with an emphasis on transaminase enzymes, which are enjoying widespread application in the synthesis of nonnatural amino acids which are key building blocks for several products of industrial importance.

The use of a range of oxidative enzymes in synthesis is covered in Chapter 11, whilst the very powerful technique of regio- and stereo-specific biohydroxylation of even complex molecules by fermenting whole-cell methods is covered in Chapter 12.

The Editors are most grateful to the authors who have submitted details of their procedures in the prescribed format for inclusion in this book. We hope that this book will increase the exposure of these methods to the chemical community and contribute to the expanded employment of biocatalysis in organic synthesis.

> John Whittall, Manchester Peter Sutton, Stevenage 2009

# **Abbreviations**

A adenine

ABTS 2,2'-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid

7-ACA 7-aminocephalosporanic acid

ACN acetonitrile AcOH acetic acid

ACS GCI American Chemical Society Green Chemistry Institute

7-ADCA 7-aminodesacetoxycephalosporanic acid

ADH alcohol dehydrogenase (alternative name for a ketoreductases or KREDs)

ADH-RE alcohol dehydrogenase from *Rhodococcus erythropolis* 

AIBN 2,2'-azobis(2-methylpropionitrile)

6-APA 6-aminopenicillanic acid

API active pharmaceutical ingredient Ara-G 9- $\beta$ -D-arabinofuranosylguanidine Ara-U 9- $\beta$ -D-arabinofuranosyluridine AspAT aspartate aminotransferase

AT aminotransferases

AZT 3'-azido-2',3'-dideoxythymidine (zidovudine)

BEHP bis(2-ethylhexyl)phthalate

BES N,N-bis(2-hydroxyethyl)-2-aminoethanesulfonic acid

BLAST basic local alignment search tool BREP butanol-rinsed enzyme preparation

BSA bovine serum albumin
BSA N-bromosuccinimide

BVMO Baeyer-Villiger monooxygenase

C cytosine

CAL-A lipase A from *Candida antarctica* CAL-B lipase B from *Candida antarctica* 

Car carboxylic acid reductase

CASTing combinatorial active site saturation test

Cbz Benzyloxycarbonyl

CCL lipase from Candida cylindracea (now known as lipase from

Candida rugosa or CRL)

#### xiv Abbreviations

CDI 1,1'-carbonyldiimidazole

CDW cell dry weight

cGMP current good manufacturing practice
CHMO cyclohexanone monooxygenase
CINV chemotherapy-induced nausea
CLEA cross-linked enzyme aggregate
CLEC cross-linked enzyme crystal
CNS Central nervous system

CPDMO cyclopentadecanone monooxygenase

CPO chloroperoxidase

CRL lipase from Candida rugosa

CSA cysteine sulfinic acid CYP cytochrome P450

DBDMH N,N'-dibromodimethylhydantoin

DBE di-n-butylether
DCM dichloromethane
DCW dry cell weight
DDI drug-drug interaction

DERA 2-deoxyribose-5-phosphate aldolase

dGTP deoxyguanosine triphosphate

DHA dihydroxyacetone

DHAP dihydroxyacetone phosphate

DHF dihydrofolate DIPE diisopropylether

DKR dynamic kinetic resolution
DMAP 4-dimethylaminopyridine
DMF dimethylformamide
DMSO dimethylsulfoxide
DNA deoxyribonucleic acid
DNAse deoxyribonuclease

dNTP deoxyribonucleotide triphosphate

DOE design of experiment
DOT dissolved oxygen tension

DSMZ Deutsche Sammlung von Mikroorganismen und Zellkulturen

dsDNA double-stranded DNA D4T dideoxydidehydrothymidine

DTT dithiothreitol

dUDP 2'-deoxyuridine-5'-diphosphate dUMP 2'-deoxyuridine-5'-monophosphate

E enantiomeric ratio

EDC 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride

EDTA ethylenediaminetetraacetic acid

EHS environmental health and safety

epPCR error-prone PCR EtOAc Ethyl acetate

FACS fluorescence-activated cell sorting

FAD flavin adenine dinucleotide

FADH<sub>2</sub> flavin adenine dinucleotide, reduced form

FASTA FAST ALL (a programme for fast protein comparison or fast

nucleotide sequence comparison)

FDA Food and Drug Administration (United States)

FDH formate dehydrogenase

FMN flavin mononucleotide (riboflavin-5'-phosphate)

FPLC fast protein liquid chromatography
FruA fructose-1,6-bisphosphate aldolase
FSA D-fructose-6-phosphate aldolase

FTIR Fourier-transform infrared spectroscopy

GABA γ-aminobutyric acid

G guanine

GC gas chromatography
GDH glucose dehydrogenase
GlcI glucose isomerase

GMO genetically modified organism
GMM genetically modified microorganism

G6P glucose-6-phosphate

G6PDH glucose-6-phosphate dehydrogenase GPC gel permeation chromatography GPO L-glycerol-3-phosphate oxidase

GR glucocorticoid receptor GRAS generally recognized as safe

GSK GlaxoSmithKline HBV hepatitis B virus

HEPES 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid

HIV human immunodeficiency virus

HMQC heteronuclear multiple quantum coherence

HNL hydroxynitrile lyase HOBt 1-hydroxybenzotriazole HOPhPr hydroxyphenylpropanol

HOTYR hydroxytyrosol HPA hydroxypyruvate HPI N-hydroxyphthalimide

HPLC high-performance liquid chromatography

HTS high-throughput screening
HVAlc Homovanillic alcohol

#### xvi Abbreviations

IMI imidacloprid

Indels insertions and deletions
IP intellectual property

IPTG isopropyl-β-D-thiogalactopyranoside

ISPR in situ product removal KPB potassium phosphate buffer

KR kinetic resolution

KRED ketoreductase (alternative name for an alcohol dehydrogenase or ADH)

LAS lovastatin ammonium salt

LB Luria-Bertani LCA life cycle analysis

LovD acyltransferase from the lovastatin biosynthetic pathway

Mab monoclonal antibody MAO-N monoamine oxidase MEA malt extract agar

MES 2-morpholino ethansulfonic acid monohydrate

MGF minimum genome factories MML lipase from *Mucor* sp.

MOPS 3-morpholino propane sulfonic acid

m.p. melting point
MPA mycophenolic acid

MPLC medium-pressure chromatography

mRNA messenger RNA
MS molecular sieves
MTBE tert-butylmethylether

MTQ methyl-tetrahydroisoquinoline

MYB malt yeast broth

NAD<sup>+</sup> β-nicotinamide adenine dinucleotide

NADH β-nicotinamide adenine dinucleotide, reduced form

NADPH β-nicotinamide adenine dinucleotide 2'-phosphate, reduced form

NADP<sup>+</sup> β-nicotinamide adenine dinucleotide 2'-phosphate

NAG N-acetyl-D-glucosamine
NAM N-acetyl-D-mannosamine
NANA N-acetyl-D-neuraminic acid

NCE new chemical entity

NK-1 neurokinin-1

NME new molecular entity

NMR nuclear magnetic resonance
NP nucleoside phosphorylase
OCH 6-oxo camphor hydrolase
ORI origin of replication
P450 cytochrome P450

P450 BM-3 cytochrome P450 BM-3 from Bacillus megaterium

PAMO phenylacetone monooxygenase

Pase acid phosphatase

PAT process analytical technology

PCL lipase from Pseudomonas cepacia (now renamed to Burkholderia

cepacia)

PCR polymerase chain reaction
PDCB potato-dextrose-carrot broth
PEP phosphoenolpyruvic acid

PFL lipase from *Pseudomonas fluorescens* 

PGA penicillin G acylase

Pip2C  $\Delta^1$ -piperideine-2-carboxylate reductase

PLE pig liver esterase

pNPG p-nitrophenyl-β-D-glucopyranosidePNP purine nucleoside phosphorylase

PPL porcine pancreatic lipase

ProSAR protein sequence–activity relationship

QbD quality by design

QSAR quantitative structure–activity relationship

RAMA rabbit muscle aldolase (fructose-1,6-bis-phosphate aldolase)

R&D research and development

rDNA recombinant DNA rRNA ribosomal RNA retention factor

RhaD rhamnulose-1-phosphate aldolase

RNA ribonucleic acid ROH generic alcohol R<sub>t</sub> retention time

SAS simvastatin ammonium salt

SCR Saccharomyces cerevisiae carbonyl reductase
SIGEX substrate-induced gene-expression screening
SMB simulated moving bed chromatography

SOT Spirulina-Ogawa-Terui ssDNA single-stranded DNA

T thymine

Taq a thermostable DNA polymerase from *Thermus aquaticus* 

TBDMSCl *tert*-butyldimethylsilyl chloride

TBME tert-butylmethylether
TCA trichloroacetic acid
TDP thymidine 5'-phosphate

TdR thymidine

TEMPO 2,2,6,6-tetramethyl-1-piperidinyloxy

#### xviii Abbreviations

**TFA** trifluoroacetic acid THF tetrahydrofuran tetrahydrofolate **THFo** 

thiamine pyrophosphate ThDP

TK transketolase

TLC thin-layer chromatography thymidine 5'-monophosphate **TMP** 

**TMS** tetramethyl silane

**TMOS** tetramethyl orthosilicate **TMSOTf** trimethylsilyl triflate TP

thymidine-5'-phosphorylase

tris-HCl tris(hydroxymethyl)aminomethane HCl

TTN total turnover number

**TYR** tyrosol

unit of enzyme activity (µmol min<sup>-1</sup>) U

U uracil

UdR 2'-deoxyuridine uridine-5'-diphosphate UDP UTP uridine-5'-triphosphate

**UDPGA** uridine-5'-diphosphoglucuronic acid **UDPGT** uridine-5'-diphosphoglucuronyl transferase

uridine-5'-phosphorylase **URDP** 

UV ultraviolet

**VVM** gas volume flow per unit of liquid volume per minute

WFCC World Federation for Culture Collections

**YPG** yeast extract-peptone-glucose

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