Enzymes in Industry

Production and Applications

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Edited by Wolfgang Gerhartz





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The cover illustration shows the Stuart-Briegleb model of lysozyme. It was produced by E. E. Polymeropoulos of ASTA Pharma AG, Frankfurt, Federal Republic of Germany. The data were taken from the Cambridge Crystallographic Data System (F. H. Allen, O. Kennard and R. Taylor, Acc. Chem. Res. 16, 146–153 (1983)). The picture was generated on a Silicon Graphics workstation using the molecular modelling program MOLCAD of the Darmstadt Institute of Technology, Darmstadt, Federal Republic of Germany.

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Foreword

"Enzymes" was the last major manuscript I edited for "Ullmann's Encyclopedia of Industrial Chemistry" – and it was the most rewarding article in my ten-year period as executive editor of Ullmann's.

Its careful planning was headed by Professor Günter Schmidt-Kastner at Bayer AG in Wuppertal, Federal Republic of Germany, who invited two other German enzyme specialists to a small symposium with the sole purpose of determining the contents of the article and proposing potential authors. The two other specialists were Professor Maria-Regina Kula of Düsseldorf University and Dr. Georg-Burkhard Kreße of Boehringer Mannheim, Tutzing. The help and advice given by these three experts are gratefully acknowledged.

The response of the suggested authors was very enthusiastic. The list of authors (page XIII) reads like an international "Who is Who" of industrial enzymology.

I accepted the publisher's invitation to make a monograph from the Ullmann article with a great deal of pleasure and cooperation with the authors has once again been perfect. All chapters have been updated, and the literature is covered until late 1989. Even the latest nomenclature of DNA modification methyltransferases has been included.

The book is recommended to all those non-enzymologists who want to know what is really happening as regards enzymes in industry. The term "industry" is used in its broadest sense: it includes the production of enzymes as well as their use in the production of bulk products, such as detergents, glucose, or fructose; in fine chemicals synthesis; in food processing and food analysis; in clinical diagnosis and therapy; and – last but obviously not least – in genetic engineering.

I hope that the reader will derive as much pleasure from reading this book as I did from producing it.

The Editor

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Abbreviations

GDP

adenosine A acetamidocinnamic acid ACA α-amino-ε-aprolactam ACL alcohol dehydrogenase ADH acceptable daily intake ADI adenosine 5'-diphosphate ADP alanine Ala Arginine Arg adenosine 5'-monophosphate AMP D,L-2-amino-\(\alpha^2\)-thiazoline-4-carboxylic acid **ATC** adenosine 5'-triphosphate ATP cytidine C copy DNA cDNA citrate lyase CL cytidine 5'-monophosphate **CMP** coenzyme A CoA citrate synthetase CS cytidine 5'-triphosphate CTP deoxy d gene locus for E. coli DNA adenine methylase (N^6 methyladenine) dam gene locus for E. coli DNA cytosine methylase (5-methylcytosine) dcmIdideoxy dd dideoxynucleoside 5'-triphosphate ddNTP diethylaminoethyl DEAE deoxyribonucleic acid DNA **DNase** deoxyribonuclease deoxynucleoside 5'-triphosphate **dNTP** 3-(3,4-dihydroxyphenylalanine)[3-hydroxy-L-tyrosine] **DOPA** decays per minute dpm double-stranded ds **Enzyme Commission** E.C. fructose 6-phosphate F₆P N-formylmethionine fMet. flavin mononucleotide FMN flavin mononucleotide, reduced FMNH, quanosine G guanosine 5'-diphosphate

Glu glutamic acid

Glv glycine

GMP guanosine 5'-monophosphate

GOD glucose oxidase

GOT glutamate - oxaloacetate transaminase

G6P glucose 6-phosphate

GPT glutamate - pyruvate transaminase

GTP guanosine 5'-triphosphate

3-HBDH 3-hydroxybutyrate dehydrogenase

HFCS high-fructose corn syrup

hsdME. coli gene locus for methylation hsdR E. coli gene locus for restriction

hsdS E. coli gene locus for sequence specificity

IDP inosine 5'-diphosphate

Tle isoleucine

INT iodonitrotetrazolium chloride

ITP inosine 5'-triphosphate LDH lactate dehydrogenase

Lvs lysine }

m(superscript) methylated

MDH malate dehydrogenase

Met methionine

M₆P mannose 6-phosphate mRNA messenger RNA

3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide MTT

any nucleotide N

NAD nicotinamide - adenine dinucleotide

NADH nicotinamide - adenine dinucleotide, reduced NADP nicotinamide – adenine dinucleotide phosphate

NADPH nicotinamide - adenine dinucleotide phosphate, reduced

NMN nicotinamide mononucleotide NTP nucleoside 5'-triphosphate

phosphate groups

р ³²Р phosphate groups containing ³²P phosphorus atoms

inorganic phosphate p_i PEP phosphoenolpyruvate

6-phosphogluconate dehydrogenase 6-PGDH

Phe phenylalanine

PMS 5-methylphenazinium methyl sulfate poly(deoxyadenosine 5'-monophosphate) poly(dA)

inorganic pyrophosphate pp;

Pro proline

PRPP phosphoribosyl pyrophosphate

Pu purine pyrimidine Py

ribo F

ribonucleic acid RNA RNase ribonuclease

S-adenosylmethionine SAM

serine hydroxymethyltransferase **SMHT**

single-stranded SS thymidine T

thymidine 5'-monophosphate TMP

transfer RNA tRNA

thymidine 5'-triphosphate TTP

uridine H

uridine 5'-monophosphate **UMP** uridine 5'-triphosphate UTP

Val valine

Bacteriophages:

fd ghl M13 N4

PBS₁

PBS2 SPO₁

SP6

SP15

T3

T4

T5 T7

XP12

à

λgt11

ΦSM11 ΦX174

Plasmids:

pBR322 pBR328 pSM1 pSP64

pSP65

pSPT18, pSPT19 pT7-1, pT7-2 pUC 18, pUC 19

pUR222

Eukaryotic viruses:

Ad2 **SV40**

Contents

	Foreword	V
	Authors	(1)
	Abbreviations x	(V
1.	Introduction	1
1.1.	History	1
1.2. 1.2.1. 1.2.2.	Enzyme Nomenclature General Principles of Nomenclature Classification and Numbering of Enzymes	5 5
1.3. 1.3.1. 1.3.2. 1.3.3. 1.3.4.	Structure of Enzymes Primary Structure Three-Dimensional Structure Quaternary Structure, Folding, and Domains	7 7 7 8
1.4. 1.4.1. 1.4.2. 1.4.3.	Biosynthesis of Enzymes	11 11 11
2.	Catalytic Activity of Enzymes	13
2.1.1. 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6.	Factors Governing Catalytic Activity Temperature Value of pH Activation Inhibition Allostery	14 14 15 16 18
2.2.1. 2.2.1. 2.2.2. 2.2.3. 2.2.4. 2.2.5. 2.2.6.	Enzyme Assays Reaction Rate as a Measure of Catalytic Activity Definition of Units Absorption Photometry Fluorometry Luminometry	20 20 20 21 23 23

Contents	
Potentiometry Conductometry Calorimetry Polarimetry Manometry Viscosimetry Immobilized Enzymes Electrophoresis	24 25 25 25 25 25 25 26 26
Quality Evaluation of Enzyme Preparations Quality Criteria Specific Activity Protein Determination Contaminating Activities Electrophoretic Purity Performance Test Stability Formulation of Enzyme Preparations	28 28 28 28 29 30 30 30
General Production Methods	33
Fermentation Organism and Enzyme Synthesis Strain Improvement Physiological Optimization The Fermentor and Its Limitations Process Design Modeling and Optimization Instrumentation and Control	33 34 35 37 39 41 42 43
Isolation and Purification Preparation of Biological Starting Materials Cell Disruption by Mechanical Methods Cell Disruption by Nonmechanical Methods Separation of Solid Matter Filtration Centrifugation Extraction Flocculation and Flotation Concentration Thermal Methods Precipitation Ultrafiltration	43 44 45 46 46 46 48 50 50 50 50 50 52 53
	Potentiometry Conductometry Calorimetry Polarimetry Manometry Viscosimetry Immobilized Enzymes Electrophoresis Quality Evaluation of Enzyme Preparations Quality Criteria Specific Activity Protein Determination Contaminating Activities Electrophoretic Purity Performance Test Stability Formulation of Enzyme Preparations General Production Methods Fermentation Organism and Enzyme Synthesis Strain Improvement Physiological Optimization The Fermentor and Its Limitations Process Design Modeling and Optimization Instrumentation and Control Isolation and Purification Preparation of Biological Starting Materials Cell Disruption by Mechanical Methods Cell Disruption by Nonmechanical Methods Separation of Solid Matter Filtration Centrifugation Extraction Flocculation and Flotation Concentration Thermal Methods Precipitation

			Contents	IX
	3.2.5. 3.2.6.	Product Formulation		62 62
	3.3. 3.3.1. 3.3.2. 3.3.3. 3.3.3.1. 3.3.3.2. 3.3.3.3. 3.3.4. 3.3.5.	Immobilization Definitions History Methods Carrier Binding Cross-linking Entrapment Characterization Application		63 64 65 65 67 70 70 73 74
Ag.	4.	Industrial Uses of Enzymes		77
	4.1. 4.1.1. 4.1.2. 4.1.3. 4.1.4. 4.1.5. 4.1.6. 4.1.7. 4.1.8. 4.1.9.	Survey of Industrial Enzymes Amylases Cellulases Hemicellulases Pectinases Proteinases Lipases Glucose Isomerase Lactases Oxidoreductases		777 78 81 83 84 85 89 91 91
	4.2.1. 4.2.2. 4.2.3. 4.2.4. 4.2.5. 4.2.6.	Enzymes in Starch Processing and Baking Syrups and Sweeteners Fuel Alcohol Baking Characterization Analysis Trade Names and Manufacturers		93 97 98 99 101
	4.3. 4.4. 4.4.1. 4.4.2. 4.4.3. 4.4.4.	Proteolytic Enzymes Detergent Enzymes Enzymes in Leather Production Enzymatic Synthesis of Aspartame Synthesis of Human Insulin from Hog Insulin		108 110 115
	4.5.	Meat Processing		118
	4.6. 4.6.1. 4.6.2. 4.6.3. 4.6.4	Dairy Products Enzymes from Rennet and Rennet Substitutes β-1,4-Galactosidases Sulfhydryl Oxidase Lysozymes		120 123 124

X	Contents	
4.6.5. 4.6.6. 4.6.7. 4.6.8.	Production of Aroma and Texture Membrane Cleansing Phosphatases Catalase	125 125
4.7.	Processing of Fruit, Vegetables, and Wine	126
4.8.	Hydrolysis of Protein, Fat, and Cellulose, and Inversion of Sucrose .	129
4.9. 4.9.1. 4.9.2. 4.9.3. 4.9.4.	Amino Acids and Hydroxycarboxylic Acids L-Amino Acids from Racemic Precursors L-Amino Acids from Prochiral Precursors L-Amino Acids from Chiral Precursors Synthesis of L-Amino Acids by Enzymatic Carbon – Carbon Bonding	130 135 138
4.9.5.	L-Hydroxycarboxylic Acids	
4.10.2.3. 4.10.3. 4.10.4. 4.10.4.1. 4.10.4.2.	Enzymes in Organic Synthesis General Considerations Enzymes Not Requiring Coenzymes Esterases, Lipases, and Amidases Aldolases Lyases, Hydrolases, and Isomerases Enzymes Requiring Coenzymes, but Not Cofactor Regeneration Systems Enzymes Requiring Added Coenzymes Enzymes Requiring Nucleoside Triphosphates Enzymes Requiring Nicotinamide Coenzymes Enzymes Requiring Other Cofactors Synthesis with Multienzyme Systems Outlook	142 142 142 144 144 145 145 145 146 148 148
5.	Enzymes in Analysis and Medicine	151
5.1. 5.1.1. 5.1.2.	Survey Enzymes in Clinical Diagnosis and Food Analysis Enzymes in Therapy	151
5.2. 5.2.1. 5.2.2. 5.2.3.	•	162 164 164
5.3. 5.3.1.	Enzymes for Food Analysis Carbohydrates	

5.3.2.

5.3.3.

5.3.4.

		Contents	XI
5.4. 5.4.1. 5.4.2. 5.4.3. 5.4.4. 5.4.5. 5.4.6. 5.4.7. 5.4.8.	Enzymes in Therapy Digestive Enzymes Debridement of Wounds Improvement of Blood Rheology Thrombolysis Support of Blood Clotting Therapy of Malignancies Chemonucleolysis in Intervertebral Disk Herniation Treatment of Inflammation and Reperfusion Injury		179 180 180 181 182 183 183
6.	Enzymes in Genetic Engineering		185
6.1. 6.1.1. 6.1.2. 6.1.2.1. 6.1.2.2. 6.1.3. 6.1.3.1. 6.1.3.2. 6.1.3.3. 6.1.4. 6.1.5.	Restriction Endonucleases and Methylases Classification Activity of Class II Restriction Endonucleases Reaction Parameters Additional Structural Requirements Influencing Activity Specificity of Class II Restriction Endonucleases Palindromic Recognition Sequences Nonpalindromic Recognition Sequences Isomers Changes in Sequence Specificity Novel Class II Restriction Endonucleases	ty	189 199 199 200 201 202 203 204 205
6.2.1. 6.2.2. 6.2.3. 6.2.4. 6.2.5.	DNA Polymerases Escherichia coli DNA Polymerase I Klenow Enzyme T4 DNA Polymerase Reverse Transcriptase Terminal Transferase		209 209 212 213
6.3. 6.3.1. 6.3.2.	RNA Polymerases SP6 RNA Polymerase T7 RNA Polymerase		216
6.4. 6.4.1. 6.4.2. 6.4.3. 6.4.4.	DNA Nucleases DNase I Exonuclease III Nuclease S1 Nuclease Bal 31		219 220 221 221
6.5.1. 6.5.2. 6.5.2.1. 6.5.2.2. 6.5.2.3. 6.5.2.4.	RNA Nucleases RNase H Site-Specific RNases RNase A RNase CL3 RNase T ₁ RNase U ₂		223 224 224 225 225

7411	Contents	
6.5.2.5. 6.5.2.6.	Nuclease S7 Site-Specific RNases in RNA Sequence Analysis	
6.6. 6.6.1. 6.6.2. 6.6.3. 6.6.4. 6.6.5. 6.6.6.	Modifying Enzymes Alkaline Phosphatase T4 DNA Ligase Escherichia coli DNA Ligase T4 Polynucleotide Kinase T4 Polynucleotide Kinase, 3'-Phosphatase-Free DNA Modification Methyltransferase (M · HpaII)	229 230 231 232. 234
7.	Economic Aspects	249
8.	Safety and Environmental Aspects	253
9.	References	255
10.	Index	299

1. Introduction

Enzymes are the catalysts of biological processes. Like any other catalyst, an enzyme brings the reaction catalyzed to its equilibrium position more quickly than would occur otherwise; an enzyme cannot bring about a reaction with an unfavorable change in free energy unless that reaction can be coupled to one whose free energy change is more favorable. This situation is not uncommon in biological systems, but the true role of the enzymes involved should not be mistaken.

The activities of enzymes have been recognized for thousands of years; the fermentation of sugar to alcohol by yeast is among the earliest examples of a biotechnological process. However, only recently have the properties of enzymes been understood properly. Indeed, research on enzymes has now entered a new phase with the fusion of ideas from protein chemistry, molecular biophysics, and molecular biology. Full accounts of the chemistry of enzymes, their structure, kinetics, and technological potential can be found in many books and series devoted to these topics [1.1]-[1.5]. This chapter reviews some aspects of the history of enzymes, their nomenclature, their structure, and their relationship to recent developments in molecular biology.

1.1. History

Detailed histories of the study of enzymes can be found in the literature [1.6], [1.7].

Early Concepts of Enzymes. The term "enzyme" (literally "in yeast") was coined by KÜHNE in 1876. Yeast, because of the acknowledged importance of fermentation, was a favorite subject of research. A major controversy at that time, associated most memorably with LIEBIG and PASTEUR, was whether or not the process of fermentation was separable from the living cell. No belief in the necessity of vital forces, however, survived the demonstration by BUCHNER (1897) that alcoholic fermentation could by carried out by a cell-free yeast extract. The existence of extracellular enzymes had, for reasons of experimental accessibility, already been recognized. For example, as early as 1783, SPALLANZANI had demonstrated that gastric juice could digest meat in vitro, and SCHWANN (1836) called the active substance pepsin. KÜHNE himself appears to have given trypsin its present name, although its existence in the intestine had been suspected since the early 19th century.

Enzymes as Proteins. By the beginning of the 20th century, the protein nature of enzymes had been recognized. Knowledge of the chemistry of proteins drew heavily on the improving techniques and concepts of organic chemistry in the second half of the 19th century; it culminated in the peptide theory of protein structure, usually credited to Fischer und Hofmeister. However, methods that had permitted the separation and synthesis of small peptides were unequal to the task of purifying enzymes. Indeed, there was no consensus that enzymes were proteins. Then, in 1926, Sumner crystallized urease from jack bean meal and announced it to be a simple protein. Against this, Willstätter argued that enzymes were not proteins but "colloidal carriers" with "active prosthetic groups." However, with the conclusive work by Northrop and his colleagues who isolated a series of crystalline proteolytic enzymes, beginning with pepsin in 1930, the protein nature of enzymes was established.

The isolation and characterization of intracellular enzymes was naturally more complicated and, once again, significant improvements were necessary in the separation techniques applicable to proteins before, in the late 1940s, any such enzyme became available in reasonable quantities. Because of the large amounts of accessible starting material and the historical importance of fermentation experiments, most of the first pure intracellular enzymes came from yeast and skeletal muscle. However, as purification methods were improved, the number of enzymes obtained in pure form increased tremendously and still continues to grow. Methods of protein purification are so sophisticated today that, with sufficient effort, any desired enzyme can probably be purified completely, even though very small amounts will be obtained if the source is poor.

Primary Structure. After the protein nature of enzymes had been accepted, the way was clear for more precise analysis of their composition and structure. Most amino acids had been identified by the early 20th century. The methods of amino acid analysis then available, such as gravimetric analysis or microbiological assay, were quite accurate but very slow and required large amounts of material. The breakthrough came with the work of MOORE and STEIN on ion-exchange chromatography of amino acids, which culminated in 1958 in the introduction of the first automated amino acid analyzer [1.8]. Modern machines have lowered the time required for an analysis to less than 1 h and the amount of protein required to $< 1 \mu g$ [1.9].

The more complex question – the arrangement of the constituent amino acids in a given protein, generally referred to as its primary structure – was solved in the late 1940 s. The determination in 1951 of the amino acid sequence of the β -chain of insulin by Sanger and Tuppy [1.10] demonstrated for the first time that a given protein does indeed have a unique primary structure. The genetic implications of this were enormous. The introduction by Edman of the phenyl isothiocyanate degradation of proteins stepwise from the N-terminus, in manual form in 1950 and subsequently automated in 1967 [1.11], provided the principal chemical method for determining the amino acid sequences of proteins. The primary structures of pancreatic ribonuclease [1.12] and egg-white lysozyme [1.13] were published in 1963. Both of