

IMMUNOLOGICAL DISEASES

Third Edition

Max Samter, M.D., Editor

Volume II

Section Editors:

David W. Talmage, M.D.

Bram Rose, M.D.

K. Frank Austen, M.D.

John H. Vaughan, M.D.

IMMUNOLOGICAL DISEASES

Third Edition

Max Samter, M.D., Editor

Volume II

Section Editors:

David W. Talmage, M.D.

Bram Rose, M.D.

K. Frank Austen, M.D.

John H. Vaughan, M.D.



Little, Brown and Company

Copyright © 1978 by Little, Brown and Company (Inc.)

Third Edition

Previous editions copyright © 1965, 1971 by Little, Brown and Company (Inc.)

All rights reserved. No part of this book may be reproduced in any form or by any electronic or mechanical means, including information storage and retrieval systems, without permission in writing from the publisher, except by a reviewer who may quote brief passages in a review.

Library of Congress Catalog Card No. 78-69906

ISBN 0-316-76985-1

Printed in the United States of America

Contents

Preface

xi

Contributing Authors

xv

A Note on Abbreviations

xxvii

Volume I

PART ONE. BASIC IMMUNOLOGY

David W. Talmage, Editor

SECTION I. ANTIGENS AND THE IMMUNE SYSTEM

1. Antigens, Haptens, and Antigenic Determinants

Hugh J. Callaban and Paul H. Maurer

3

2. Localization and Fate of Foreign Antigens in Tissues

Judith Mitchell and Gustav J. V. Nossal

22

3. The Cellular Basis of Immune Responses

J. F. A. P. Miller

35

SECTION II. THE HUMORAL RESPONSE TO ANTIGEN

4. The Structure, Genetics, and Biological Properties of Immunoglobulins

Joseph A. Gally

49

5. Histology of Immune Responses

Frank W. Fitch and Robert L. Hunter, Jr.

81

6. Regulation of Antibody Formation

David W. Thomas and David W. Talmage

105

7. Antibody Specificity

Frank F. Richards, Robert W. Rosenstein, Janos M. Varga, and William H. Konigsberg

121

8. The Detection and Measurement of Antigen-Antibody Interaction

Alfred J. Crowle

139

9. Experimental Anaphylaxis and Reaginic Hypersensitivity

Kimishige Ishizaka

164

10. The Chemical Mediators of Immediate Hypersensitivity Reactions

K. Frank Austen

183

11. Immune Complex Injury

Charles G. Cochrane and Frank J. Dixon

210

12. Eosinophils

Max Samter and Stephen I. Wasserman

230

13. Human Complement System

Peter F. Kohler

244

SECTION III. CELLULAR RESPONSES TO ANTIGEN

14. Delayed Hypersensitivity	<i>George Mackaness</i>	281
15. The Immune Granulomas	<i>Emil R. Unanue</i>	297
16. Lymphocyte Mediators: The "Lymphokines"	<i>John R. David and Ross E. Rocklin</i>	307
17. The Major Histocompatibility Complex: Genetic Control of Antigens Eliciting Cell-Mediated Immune Reactions	<i>Fritz H. Bach and Marilyn L. Bach</i>	325
18. Tissue Transplantation	<i>David W. Scott and D. B. Amos</i>	341
19. The Immunology of Animal Tumors	<i>Robert C. Bast, Jr., and Herbert J. Rapp</i>	359

SECTION IV. IMMUNOLOGICAL UNRESPONSIVENESS

20. Immunological Tolerance	<i>William O. Weigle</i>	389
-----------------------------	--------------------------	-----

PART TWO. NONATOPIC IMMUNOLOGICAL DISORDERS

SECTION I. RESPONSES TO THERAPEUTIC AGENTS AND MEASURES

21. Drug Reactions	<i>Alain L. de Weck</i>	413
22. Transfusion Reactions and Blood Group Substances	<i>Scott N. Swisher and Jacob Nusbacher</i>	440
23. Allograft Rejection	<i>John P. Merrill</i>	460

SECTION II. DISEASES WITH ABNORMAL IMMUNE RESPONSES

24. The Immunodeficiency Syndromes	<i>Fred S. Rosen</i>	472
25. Multiple Myeloma and Related Plasma Cell Dyscrasias	<i>Elliott F. Osserman</i>	499
26. Lymphoma, Leukemia, and Hodgkin's Disease	<i>Alan C. Aisenberg</i>	530
27. Sarcoidosis	<i>Louis E. Siltzbach and Robert N. Taub</i>	548
23. The Cryopathies	<i>Leonard A. Moroz and Bram Rose</i>	570

SECTION III. HYPERSENSITIVITY TO
INFECTIOUS AGENTS

29. Streptococcal Infections and Nonsuppurative Complications
Berno Heymer and Richard M. Krause 592
30. Tuberculosis
Maurice J. Lefford 613
31. Leprosy
J. L. Turk and M. F. R. Waters 627
32. Syphilis
Daniel M. Musher and Robert E. Baughn 639
33. Brucellosis
Margaret E. Meyer 651
34. Histoplasmosis and Cryptococcosis
John P. Utz 660
35. Toxoplasmosis
John A. Robinson 669
36. Coccidioidomycosis
Demosthenes Pappagianis 681
37. Pulmonary Aspergillosis, Farmer's Lung, and Related Diseases
Jack Pepys 692
38. Worms
Kenneth S. Warren 718
39. Protozoa
Adel A. F. Mahmoud 738
40. Virus and Virus Vaccine-Induced Immunological Injury
Vincent A. Fulginiti 752
41. Cancer Antigens in Man
Samuel O. Freedman and Phil Gold 762

Index to Volumes I and II

Volume II

PART THREE. THE ATOPIC DISEASES

42. The Atopic Diseases: Definition and Comment
K. Frank Austen, Editor 781
43. Antigens That Cause Atopic Disease
Te P. King and Philip S. Norman 787
44. IgE and Atopic Disease
Lawrence M. Lichtenstein and Robert N. Hamburger 804
45. The Genetics of Atopic Allergy
David G. Marsh and Wilma B. Bias 819
46. Allergic Rhinitis
Philip S. Norman and Lawrence M. Lichtenstein 832

47. The Pathogenesis of Bronchial Asthma	<i>Bram Rose, James C. Hogg, and Peter Macklem</i>	852
48. Bronchial Asthma: Clinical Course and Treatment	<i>Robert P. Orange and Bram Rose</i>	868
49. The Anaphylactic Syndrome	<i>K. Frank Austen</i>	885
50. The Aspirin Triad and the Prostaglandins	<i>Max Samter and Howard J. Zeitz</i>	900
51. Insect Venom Allergy	<i>Martin D. Valentine</i>	917
PART FOUR. ALLERGIC REACTION PATTERNS OF THE SKIN	<i>K. Frank Austen, Editor</i>	
52. Urticaria	<i>Robert P. Warin and Robert H. Champion</i>	929
53. Angioedema	<i>Irma Gigli, Albert L. Sheffer, and K. Frank Austen</i>	941
54. Atopic Dermatitis and Infantile Eczema	<i>Lawrence M. Solomon and Adolph Rostenberg, Jr.</i>	953
55. Allergic Eczematous Contact Dermatitis	<i>Rudolf L. Baer and Leonard C. Harber</i>	966
56. Erythema Nodosum	<i>William L. Epstein</i>	975
57. Erythema Multiforme and Toxic Epidermal Necrolysis	<i>Nicholas A. Soter and Irwin M. Freedberg</i>	984
58. Cutaneous Necrotizing Angiitis	<i>Nicholas A. Soter and K. Frank Austen</i>	993
59. Bullous Diseases of the Skin	<i>Robert E. Jordon and Thomas T. Provost</i>	1002
60. Reactions to Light, Heat, and Trauma	<i>Leonard C. Harber and Rudolf L. Baer</i>	1014
PART FIVE. DISEASES WITH IMMUNOLOGICAL FEATURES	<i>John H. Vaughan, Editor</i>	
61. Autoimmune and Histocompatibility (HLA)-Associated Diseases: General Considerations	<i>John H. Vaughan</i>	1029
SECTION I. CONNECTIVE TISSUE DISEASES		
62. Systemic Lupus Erythematosus	<i>Eng M. Tan and Naomi F. Rothfeld</i>	1038
63. Rheumatoid Arthritis	<i>Charles L. Christian and Stephen A. Paget</i>	1061

64. Polyarteritis and Other Primary Vasculitides	<i>David J. Gocke and L. A. Healey</i>	1077
65. Polymyositis and Dermatomyositis	<i>Carl M. Pearson</i>	1091
66. Progressive Systemic Sclerosis (Scleroderma)	<i>Gerald P. Rodnan</i>	1109
67. Mixed Connective Tissue Disease	<i>Gordon C. Sharp</i>	1142
68. Sjögren's Syndrome	<i>John A. Hardin and Kurt J. Bloch</i>	1151
69. Amyloidosis	<i>Edward C. Franklin and Evan Calkins</i>	1158
70. The Seronegative Spondyloarthropathies	<i>Rodney Bluestone</i>	1175
SECTION II. HEMATOLOGICAL DISEASES		
71. Acquired Immune Hemolytic Disorders (Including Drug-Induced Immune Hemolytic Anemia)	<i>John P. Leddy and Scott N. Swisher</i>	1187
72. Autoimmune Thrombocytopenic Purpura	<i>Simon Karparkin</i>	1228
73. Hemostatic Mechanisms in Immunological Processes	<i>Oscar D. Ratnoff</i>	1244
SECTION III. ENDOCRINE AND ASSOCIATED ORGAN DISEASES		
74. Hashimoto's Disease and Graves' Disease	<i>G. N. Beall and D. H. Solomon</i>	1261
75. Adrenalitis, Hypoparathyroidism, and Associated Diseases	<i>W. J. Irvine</i>	1278
76. Pernicious Anemia and Atrophic Gastritis	<i>Graham H. Jeffries</i>	1296
77. Immunological Factors in Human Male and Female Infertility	<i>R. E. Mancini and J. A. Andrada</i>	1308
78. Diabetes Mellitus	<i>Jørn Nerup, Morten Christy, Torsten Deckert, and Jørn Egeberg</i>	1330
SECTION IV. NONENDOCRINE ORGAN DISEASES		
79. Glomerulonephritis	<i>Curtis B. Wilson and Frank J. Dixon</i>	1348

80. Myasthenia Gravis	<i>Dale E. McFarlin</i>	1383
81. The Demyelinating Diseases	<i>Philip Y. Paterson</i>	1400
82. Uveitis	<i>Edward L. Howes, Jr., and Samuel B. Aronson</i>	1436
83. Chronic Active Hepatitis, Cirrhosis, and Other Diseases of the Liver	<i>Ian Reay Mackay</i>	1454
84. The Immunology of the Gastrointestinal Tract and Inflammatory Bowel Disease	<i>T. B. Tomasi, Jr., and R. G. Shorter</i>	1478
85. Myocardial Disease	<i>Melvin H. Kaplan and Louis Rakita</i>	1502
SECTION V. IMMUNOSUPPRESSION		
86. Pharmacological Immunosuppression	<i>Alfred D. Steinberg and Gary W. Williams</i>	1522
A Glossary of Immunological Terms		1541
Index to Volumes I and II		[3]

PART THREE: THE ATOPIC DISEASES

K. Frank Austen, Editor

"Atopy," in its original definition, described an altered state of reactivity which occurs in families and leads to the development of "reagins" and disease after "natural" exposure to specific allergens.

Like other terms which served a healthy purpose during the early part of the century because they emphasized the heterogeneity of altered reactivity, "atopy" has come to raise more questions than it answers.¹ Some of these questions attempt to clarify the genetic make-

up of atopic patients, clearly recognized by William Sherman in the second edition of this text, and brought up to date by Marsh and Bias in Chapter 45. Genetics, however, might not only refer to specific immune response genes, but also determine the level of total IgE, and even the affinity of IgE for allergen and for mast cell receptors. Consequently, investigations of atopy must extend to the regulation of mediator generation and release as well as to the extracellular controls of mediator function. While this brings the atopic diseases closer to the "diseases of immediate hypersensitivity," the atopic diseases have a sufficient number of characteristics of their own that, at least for the time being, the term *atopic diseases* should be retained.

¹Samter had previously questioned the legitimacy of the term "atopy" (*Med. Clin. North Am.* 58:233, 1974).

"For years, the definition remained unchallenged, even though it failed to explain common clinical observations. For instance, technicians in laboratories might become sensitive to dander of guinea pigs, rats, rabbits, and dogs, but not to ragweed pollen, even in areas in which hay fever is far more prevalent than sensitivity to animal dander. On the other hand, the hay fever patient, once pollen-sensitive, would remain pollen-sensitive after years of transfer to a pollen-free area, even though the half-life of IgE turned out to be days or weeks, not years.

"Even today, the patient who has life-long and often overwhelming sensitivity—associated with circulating IgE and positive skin reactions—to a single antigen, e.g. to crustacea, defies classification. Can he be atopic, in the absence of any sensitivity to common allergens? Some say "yes" and apply the term atopy to any clinical entity which can be traced to IgE, but most of us prefer to limit it to those conditions which have a familial component. Infestation with ascaris and trichinella spiralis might induce synthesis of IgE and massive eosinophilia, but they are non-atopic diseases: the point, however, has not been settled, and our terminology must be tightened."

42. The Atopic Diseases: Definition and Comment

K. Frank Austen

Early studies of allergy focused on the relationships of hay fever and asthma in humans to experimental anaphylaxis in animals [4] on the assumption that the observed species differences were attributable to the structure of the antigens, the route and form of their administration, and the nature of the subject's subsequent encounter with the antibody [43]. Indeed, "naturally" sensitized humans who developed hay fever or asthma upon exposure to horses, and humans sensitized by prior injections with horse serum containing diphtheria antitoxin exhibited the same anaphylactic response to administration of a therapeutic dose of horse serum [23]. Both routes of sensitization were associated with the presence of skin-sensitizing antibodies which could be passively transferred to nonsensitive recipients, but the injection of horse serum produced an additional specific antibody of the IgG class [13]. The subsequent recognition of spontaneous asthma due to environmental allergens in dogs and the demonstration that their anaphylactic response to allergen is indistinguishable from that of actively sensitized dogs challenged with specific proteins [52, 53] supported the early view that the immunological mechanisms recognized in human allergic rhinitis and asthma bore some relationship to those studied experimentally in animals. However, as Sherman has pointed out [68], the animal models lacked the genetic considerations central to the human diseases, failed to manifest the characteristics of atopic dermatitis, and generally involved heat-stable homocytotropic antibodies [6].

The association of allergic rhinitis and asthma in the same patient, often with a familial back-

ground, and the presence in the serum of passive transfer activity led Coca and Cooke in 1923 [12] to introduce the term "atopy" for the familial tendency of patients to become sensitive to "allergens" and to develop, alone or in combination, such clinical conditions as bronchial asthma, rhinitis, urticaria and eczematous dermatitis (atopic dermatitis). Specific IgE, however, can develop without an atopic setting (e.g., in certain forms of urticaria and in the anaphylactic syndrome), and heat-stable homocytotropic antibodies have now been recognized in humans with reversible airway disease [7].

Most terms are broad when they are coined and narrowed as our experience grows: "allergy"—as conceived by von Pirquet in 1906 [75]—described "altered reactivity" which could be either injurious or beneficial to the host. Common usage eventually made allergy synonymous with "hypersensitivity"; and the term "atopy," in general, was applied to the "classic" diseases of allergy. It seems likely that the unrestricted term "allergic diseases" or the restrictive connotation of "atopic diseases" might be replaced by a more precise definition which must account for the unexplained clinical course of atopic dermatitis [5] or the chronic hyperirritability of the airways demonstrable in an asymptomatic asthmatic subject [45].

Mast Cells and Mediators

The importance of the presence of specific IgE in atopic diseases by implication involves tissue mast cells as the target cells of the antigen challenge and mast cell-derived mediators as the effector principles of the symptom complex. Although reversible bronchospasm [67] of hyper-responsive airways [14, 18, 50, 73] is the hallmark of natural and experimentally induced bronchial asthma, the functional abnormalities are now understood to include not only increases in bronchomotor tone and gas trapping [57] but also a decrease in static compliance [42, 59]. Further, the characteristic pathological features include edema of the submucosa and mucosa; thickening of the basement membrane; infiltration with polymorphonuclear leukocytes of the eosinophilic and neutrophilic type; and a bronchiolar exudate of shed epithelial cells, viscid mucus, and a serous transudate [1, 19]. In order to implicate mast cell-mediated events in such a disorder it is necessary to establish acces-

From the Departments of Medicine, Harvard Medical School and Robert B. Brigham Hospital, Boston, Massachusetts 02120.

Supported by grants AI-07722, HL-17382, and AI-10356 from the National Institutes of Health.

sibility to environmental stimuli and capacity to elicit both acute *pathopharmacological* and subacute/chronic *inflammatory* tissue changes [2].

The presence of mast cells in the bronchial mucosa and lumen of rhesus monkeys and humans [48, 54], the antigen reactivity of comparable cells from the bronchial lumen of dogs [55], and the intraepithelial location of IgE-bearing mast cells in the tonsils, adenoids, and nasal polyps of humans [20] indicate that initial mediator release may occur without involvement of interstitial mast cells. Subsequent alterations in mucosal impermeability to allergens or in delivery of allergen after absorption might account for the decrease in total numbers of detectable mast cells and the prominence of disrupted cells with scattered granules throughout the bronchial connective tissue in acute allergic asthma [66]. Although measurements of systemic release of mediators in spontaneous and experimentally induced asthma are generally lacking, it appears that such measurements could be made, since recent studies [37, 70] have confirmed earlier findings [61] of the release into the venous effluent of chemical mediators during cold-induced urticaria-angioedema, as demonstrated with newly available specific and sensitive techniques for analyzing complex biological fluids.

Mast Cell Regulation

Mast cell-dependent phenomena are regulated at many levels: by the intensity of the mast cell-activating event, by the endogenous determinants of the ratio of preformed and newly generated mediators released, by the receptor binding and responsiveness of target cells to released mediators, and by the rate at which released mediators undergo biodegradation. An example of each of these control levels has been recognized in *in vitro* systems, which serves to emphasize that the pathobiological considerations of immediate type hypersensitivity reactions need not be limited to a focus on specific IgE alone. At the first level, a minimal IgE-mediated stimulus may cause intracellular SRS-A generation without release of SRS-A or histamine. At the second level, histamine, a primary preformed mediator, acting via an H₂ receptor to activate adenylate cyclase, increases cellular levels of cyclic 3',5'-adenosine monophosphate (cyclic AMP), thereby suppressing further

mediator release [39]. At the third level, the NH₂- and COOH-terminal tripeptides of ECF-A block the chemotactic response of eosinophils to ECF-A by reversible competition for the receptor and irreversible deactivation, respectively [26]; such tripeptides would be readily derived from the eosinophilotactic tetrapeptides [24] (ECF-A) by cleavage with carboxypeptidase A and aminopeptidase M. Conversely histamine, in minimally chemotactic doses [11], facilitates the response of the eosinophil to the eosinophilotactic tetrapeptides [25].¹ Finally eosinophils, concentrated by directed migration, inactivate released primary mediators such as SRS-A [77], histamine [78], and a platelet-activating factor [38] because of their content of arylsulfatase B, histaminase, and phospholipase D, respectively. Biodegradation is not dependent wholly on responding and infiltrating cells in that in the human lung arylsulfatase B, which predominates over arylsulfatase A in that organ, inactivates SRS-A in a reaction comparable to that achieved with the eosinophil-derived enzyme [76].

Level two is the point at which regulation has been extensively considered in atopic patients in regard to a possible functional impairment of the β -adrenergic receptor [71]. A selective defect in responsiveness of peripheral blood leukocytes to β -adrenergic agonists does appear to be acquired by some patients and to be more prominent with increased disease severity [49]; this might represent receptor shedding, as noted for lymphocytes cultured with insulin [22] and for human fibroblasts grown in the presence of various β -adrenergic agonists [21]. Even without administration of β -receptor agonists [46], an impaired response of peripheral leukocytes to β -agonists *in vitro* has been reported in studies of cells from untreated patients who have bronchial asthma [51] and atopic dermatitis without respiratory symptoms [60]. While endogenous catecholamine release from the adrenal glands by histamine [58] could be a factor, urinary excretion of catecholamines in atopic dermatitis has been normal [31].

¹Histamine has a positive chemokinetic effect on neutrophilic and eosinophilic polymorphonuclear leukocytes via their H₁ or H₂ receptors; and H₁ agonists augment the response to cell-specific chemotactic factors, whereas H₂ agonists block that same response (E. J. Goetzl and K. F. Austen, *Progress in Immunology III*. Australian Academy of Science, 1977. P. 439).

Mast Cell Function

The mast cell is the only cell which has a specific recognition unit, IgE, for a foreign substance and is located in the tissues in general rather than in a circulation, hematogenous or lymphatic. It is tempting to speculate that the physiological role of the mast cell in host defense is the recruitment of proteins and cells from the circulation to the reaction site. Once specific antibody and complement, followed in hours by polymorphonuclear leukocytes, have arrived at the reaction site to create the *cellular phase*, the initial or *humoral phase* mediated by substances such as histamine and SRS-A is terminated, not only by local controls but also by the constituents of the cellular phase attracted by such factors as ECF-A and HMW-NCF. Failure to limit the humoral phase creates a *pathopharmacological state* recognized clinically as urticaria, angioedema, exacerbations of rhinitis or asthma, or systemic anaphylaxis. Similarly, an inability to regulate the cellular phase would induce a local *inflammatory state* with attendant implications for long-term low-grade injury in nasal and bronchial tissues of atopic persons. Thus the mast cell and its mediators might be involved in diseases other than those initiated by IgE-dependent mechanisms.

That the mast cell and its constituent mediators have multiple capabilities is demonstrated by the dual, early and late, pulmonary and cutaneous responses to aerosol and intracutaneous administration, respectively, of *Aspergillus* [41, 56] and avian materials [29, 30]. Similar dual responses in sensitive subjects have been observed in skin with *Bacillus subtilis* enzyme preparations [17] and in both skin [72] and airways [10] with grass pollen. Even more telling has been the elicitation of dual responses* in normal subjects to crystalline *B. subtilis* α -amylase in the P-K reaction and to the F(ab)₂ fragment of sheep antihuman IgE [16]. These latter studies reveal that IgE-dependent mast cell activation is sufficient to achieve a clinical local inflammatory response characterized histologically by mast cell degranulation and infiltration with eosinophils and some neutrophils.

Immediate type hypersensitivity is present in patients experiencing metazoal parasite infections, as revealed by specific immediate type skin tests [33] and the release of histamine and ECF-A from peripheral leukocytes interacted with appropriate helminth antigen [64]. The

clinical course may include signs compatible with immediate type hypersensitivity reactions [65] and is characteristically marked by a profound elevation in IgE [32, 34, 62], of which only a fraction is specific [15]. In a particularly intriguing study, serum IgE levels and peripheral blood eosinophils were found to be elevated and fecal egg counts of *Necator americanus* substantially reduced in atopic as compared to nonatopic native inhabitants of a hookworm-infested region of New Guinea [27, 28]. The capacity of the tissue mast cell to recruit blood protein and cellular elements in the humoral phase of its response to specific antigen or to a parasitic degranulating factor [74] might be particularly pertinent to the control of the skin-penetrating or gut-attaching phase of a helminth cycle. For example, Ogilvie et al. [47] protected rats against invading cercariae of *Schistosoma mansoni* by intradermal injection of specific reagin-containing serum but not by intraperitoneal administration of much larger quantities of antiserum. *Nippostrongylus brasiliensis* expulsion from the gastrointestinal tract of the rat is associated with macromolecular leak as assessed with Evans blue dye [3] or horseradish peroxidase in electron microscopy [44], suggesting marked release of chemical mediators. Such an event might serve to clear worms already damaged by immune humoral or cellular events and might contribute directly to impairing the integrity of the parasite [63]. In schistosomiasis, killing of the schistosomula in vitro by human and peripheral blood leukocytes has been shown to be an IgG antibody-dependent function of the eosinophil [8, 9], and this has been confirmed in in vivo studies in mice [40].² In these circumstances both the humoral and cellular phases of the specific mast cell response might contribute to host resistance. Since both IgE and certain IgG subgroups mediate antigen-dependent mast cell reactions, there is ample opportunity for the mast cell activation to be closely linked to initiation of other effector pathways of host resistance. It is now critical to begin to assess tissue mast cells and their

²Non-antibody-dependent eosinophil cytotoxicity for schistosomula occurs via the capacity of the helminth surface to activate the alternative complement pathway (A. Sher, *Nature* 263:334, 1976; F. J. Ramalho-Pinto, D. J. McLaren, and S. R. Smithers, *J. Exp. Med.* 147:147, 1978), and the anaphylatoxic by-products would also recruit mast cell factors.

state of activation in experimental studies of host resistance and in human disease states by quantitative cell counts, ultrastructural techniques, and quantitative measurements of preformed and newly generated mediators [69].

References

1. Austen, K. F., and Blennerhassett, J. Bronchial Asthma. In P. A. Miescher and H. J. Müller-Eberhard (Eds.), *Textbook of Immunopathology*. New York: Grune & Stratton, 1969.
2. Austen, K. F., and Orange, R. P. Bronchial asthma: The possible role of the chemical mediators of immediate hypersensitivity in the pathogenesis of subacute chronic disease. *Am. Rev. Respir. Dis.* 112:423, 1975.
3. Barth, E. E. E., Jarrett, W. F. H., and Urquhart, G. M. Studies on the mechanism of the self-cure reaction in rats infected with *Nippostrongylus brasiliensis*. *Immunology* 10:459, 1966.
4. Becker, E. L. Nature and classification of immediate-type allergic reactions. *Adv. Immunol.* 13:267, 1971.
5. Blaylock, W. K. Atopic dermatitis: Diagnosis and pathobiology. *J. Allergy Clin. Immunol.* 57:62, 1976.
6. Bloch, K. J., and Ohman, J. L. The Stable Homocytotropic Antibodies of Guinea Pig, Mouse, and Rat; and Some Indirect Evidence for the In Vivo Interaction of Homocytotropic Antibodies of Two Different Rat Immunoglobulin Classes at a Common Receptor on Target Cells. In K. F. Austen and E. L. Becker (Eds.), *Biochemistry of the Acute Allergic Reactions*. Oxford: Blackwell, 1971.
7. Bryant, D. H., Burns, M. W., and Lazarus, L. Identification of IgG antibody as a carrier of reaginic activity in asthmatic patients. *J. Allergy Clin. Immunol.* 56:417, 1975.
8. Butterworth, A. E., Sturrock, R. F., Houba, V., Mahmoud, A. A. F., Sher, A., and Rees, P. H. Eosinophils as mediators of antibody-dependent damage to schistosomula. *Nature* 256:727, 1975.
9. Butterworth, A. E., Sturrock, R. F., Houba, V., and Rees, P. H. Antibody-dependent cell-mediated damage to schistosomula in vitro. *Nature* 252:503, 1974.
10. Citron, K. M., Frankland, A. W., and Sinclair, J. D. Inhalation tests of bronchial hypersensitivity in pollen asthma. *Thorax* 13:229, 1958.
11. Clark, R. A. F., Gallin, J. I., and Kaplan, A. P. The selective eosinophil chemotactic activity of histamine. *J. Exp. Med.* 142:1462, 1975.
12. Coca, A. F., and Cooke, R. A. On the classification of the phenomena of hypersensitiveness. *J. Immunol.* 8:163, 1923.
13. Cooke, R. A., and Spain, W. C. Studies in hypersensitiveness: A comparative study of antibodies occurring in anaphylaxis, serum disease, and the naturally sensitive man. *J. Immunol.* 17:295, 1929.
14. Curry, J. J. Comparative action of acetyl-beta-methylcholine and histamine on the respiratory tract in normals, patients with hay fever, and subjects with bronchial asthma. *J. Clin. Invest.* 26:430, 1947.
15. Dessaint, J. P., Capron, M., Bout, D., and Capron, A. Quantitative determination of specific IgE antibodies to schistosome antigens and serum IgE levels in patients with schistosomiasis (*S. mansoni* or *S. haematobium*). *Clin. Exp. Immunol.* 20:427, 1975.
16. Dolovich, J., Hargreave, F. E., Chalmers, R., Shier, K. J., Gauldie, J., and Bienenstock, J. Late cutaneous allergic responses in isolated IgE-dependent reactions. *J. Allergy Clin. Immunol.* 52:38, 1973.
17. Dolovich, J., and Little, D. C. Correlates of skin test reactions to *Bacillus subtilis* enzyme preparations. *J. Allergy Clin. Immunol.* 49:43, 1972.
18. Dubois, A. B., and Dautrebande, L. Acute effects of breathing inert dust particles and of carbachol aerosol on the mechanical characteristics of the lungs in man. Changes in response after inhaling sympathomimetic aerosols. *J. Clin. Invest.* 37:1746, 1958.
19. Dunnill, M. S. The Pathology of Asthma. In R. Porter and J. Birch (Eds.), *Identification of Asthma*. Edinburgh and London: Churchill Livingstone, 1971.
20. Feltkamp-Vroom, T. M., Stallman, P. J., Aalberse, R. C., and Reerink-Brongers, E. E. Immunofluorescence studies on renal tissue, tonsils, adenoids, nasal polyps, and skin of atopic and nonatopic patients, with specific reference to IgE. *Clin. Immunol. Immunopathol.* 4:392, 1975.
21. Franklin, T. J., Morris, W. P., and Twose, P. A. Desensitization of beta adrenergic receptors in human fibroblasts in tissue culture. *Mol. Pharmacol.* 11:485, 1975.
22. Gavin, J. R., Roth, J., Neville, D. M., deNeyts, P., and Buell, D. N. Insulin-dependent regulation of insulin receptor concentrations: A direct demonstration in cell culture. *Proc. Natl. Acad. Sci. U.S.A.* 71:84, 1974.
23. Gillette, H. F. Untoward results from diphtheria antitoxin, with special reference to its relation to asthma. *Ther. Gaz.* 33:159, 1909.
24. Goetzl, E. J., and Austen, K. F. Purification and synthesis of eosinophilotactic tetrapeptides of human lung: Identification as eosinophil chemotactic factor of anaphylaxis. *Proc. Natl. Acad. Sci. U.S.A.* 72:4123, 1975.
25. Goetzl, E. J., and Austen, K. F. Specificity and Modulation of the Eosinophil Polymorphonuc-

- lear Leukocyte Response to the Eosinophil Chemotactic Factor of Anaphylaxis (ECF-A). In S. G. O. Johansson, K. Strandberg, and B. Uvnäs (Eds.), *Molecular and Biological Aspects of the Acute Allergic Reaction*. London: Plenum, 1976.
26. Goetzel, E. J., and Austen, K. F. Structural determinants of the eosinophil chemotactic activity of the acidic tetrapeptides of ECF-A. *J. Exp. Med.* 144:1424, 1976.
27. Grove, D. I., Burston, T. O., and Forbes, I. J. Immunoglobulin E and eosinophil levels in atopic and non-atopic populations infested with hookworm. *Clin. Allergy* 4:295, 1974.
28. Grove, D. I., and Forbes, I. J. Increased resistance to helminth infestation in an atopic population. *Med. J. Aust.* 1:336, 1975.
29. Hargreave, F. E., and Pepys, J. Allergic respiratory reactions in bird fanciers provoked by allergen inhalation provocation tests. Relation to clinical features and allergic mechanism. *J. Allergy Clin. Immunol.* 50:157, 1972.
30. Hargreave, F. E., Pepys, J., Longbottom, J. L., and Wraith, D. G. Bird breeder's (fancier's) lung. *Lancet* 1:44, 1966.
31. Hingerty, D., and Meehan, F. O. C. Catecholamine excretion in children with atopic eczema. *Ir. J. Med. Sci.* 6:71, 1964.
32. Hogarth-Scott, R. S., Johansson, S. G. O., and Bennich, H. Antibodies to *Toxocara* in the sera of visceral larva migrans patients: The significance of raised levels of IgE. *Clin. Exp. Immunol.* 5:619, 1969.
33. Jarrett, E. E. E., and Urquhart, G. M. The immune response to nematode infections. *Int. Rev. Trop. Med.* 4:53, 1971.
34. Johansson, S. G. O., Mellbin, T., and Vahlquist, B. Immunoglobulin levels in Ethiopian preschool children with special reference to high concentrations of immunoglobulin E (IgND). *Lancet* 1:1118, 1968.
35. Jones, V. E., Edwards, A. J., and Ogilvie, B. M. The circulating immunoglobulins involved in protective immunity to the intestinal stage of *Nippostrongylus brasiliensis* in the rat. *Immunology* 18:621, 1970.
36. Kaliner, M., Wasserman, S. I., and Austen, K. F. Immunologic release of chemical mediators from human nasal polyps. *N. Engl. J. Med.* 289:277, 1973.
37. Kaplan, A. P., Gray, L., Shaff, R. E., Horakova, Z., and Beaven, M. A. In vivo studies of mediator release in cold urticaria and cholinergic urticaria. *J. Allergy Clin. Immunol.* 55:394, 1975.
38. Kater, L. A., Goetzel, E. J., and Austen, K. F. Isolation of human eosinophil phospholipase D. *J. Clin. Invest.* 57:1173, 1976.
39. Lichtenstein, L. M., and Gillespie, E. Inhibition of histamine release by histamine is controlled by an H₂ receptor. *Nature* 244:287, 1973.
40. Mahmoud, A. A. F., Warren, K. S., and Peters, P. A. A role for the eosinophil in acquired resistance to *Schistosoma mansoni* infection as determined by anti-eosinophil serum. *J. Exp. Med.* 142:805, 1975.
41. McCarthy, D. S., and Pepys, J. Allergic bronchopulmonary aspergillosis. Clinical immunology: (2) Skin, nasal and bronchial tests. *Clin. Allergy* 1:415, 1971.
42. McFadden, E. R., and Lyons, H. A. Serial factors influencing airway dynamics during recovery from acute asthma attacks. *J. Appl. Physiol.* 27:452, 1969.
43. Meltzer, S. T. Asthma, a phenomenon of anaphylaxis. *J.A.M.A.* 55:1021, 1910.
44. Murray, M., Jarrett, W. F. H., and Jennings, F. W. Mast cells and macromolecular leak in intestinal immunological reactions. The influence of sex of rats infected with *Nippostrongylus brasiliensis*. *Immunology* 21:17, 1971.
45. Nadel, J. A. Neurophysiologic Aspects of Asthma. In K. F. Austen and L. M. Lichtenstein (Eds.), *Asthma: Physiology, Immunopharmacology, and Treatment*. New York: Academic, 1973.
46. Nelson, H. R. The effect of ephedrine on the response to epinephrine in normal men. *J. Allergy Clin. Immunol.* 51:191, 1973.
47. Ogilvie, B. M., Smithers, S. R., and Terry, R. J. Reagin-like antibodies in experimental infections of *Schistosoma mansoni* and the passive transfer of resistance. *Nature* 209:1221, 1966.
48. Orange, R. P. Immunopharmacological aspects of bronchial asthma. *Clin. Allergy* 3(Suppl.):521, 1973.
49. Parker, C. W. Adrenergic Responsiveness in Asthma. In K. F. Austen and L. M. Lichtenstein (Eds.), *Asthma: Physiology, Immunopharmacology, and Treatment*. New York: Academic, 1973.
50. Parker, C. W., Bilbo, R. E., and Reed, C. E. Methacholine aerosol as a test for bronchial asthma. *Arch. Intern. Med.* 115:452, 1965.
51. Parker, C. W., and Smith, J. W. Alterations in cyclic adenosine monophosphate metabolism in human bronchial asthma: I. Leukocyte responsiveness to β -adrenergic agents. *J. Clin. Invest.* 52:48, 1973.
52. Patterson, R. Laboratory models of reaginic allergy. *Prog. Allergy* 13:332, 1969.
53. Patterson, R., Pruzansky, J. J., and Chang, W. W. Y. Spontaneous canine hypersensitivity to ragweed: Characterization of the serum factor transferring skin, bronchial, and anaphylactic sensitivity. *J. Immunol.* 90:35, 1963.
54. Patterson, R., Suszko, I. M., and Zeiss, C. R., Jr. Reactions of primate respiratory mast cells. *J. Allergy Clin. Immunol.* 50:7, 1972.
55. Patterson, R., Tomita, Y., Oh, S. H., Suszko,

- I. M., and Pruzansky, J. J. Respiratory mast cells and basophiloid cells: I. Evidence that they are secreted into the bronchial lumen, morphology, degranulation, and histamine release. *Clin. Exp. Immunol.* 16:223, 1974.
56. Pepys, J., Turner-Warwick, M., Dawson, P. L., and Hinson, K. R. W. Arthus (Type III) Skin Test Reactions in Man. Clinical and Immunopathological Features. In B. Rose, R. Richer, A. Sehon, and A. W. Frankland (Eds.), *Allergology* (No. 162). Amsterdam: Excerpta Medica, 1968.
57. Permutt, S. Some Physiological Aspects of Asthma: Bronchomuscular Contraction and Airways Calibre. In R. Porter and J. Birch (Eds.), *Identification of Asthma*. Edinburgh and London: Churchill Livingstone, 1971.
58. Piper, P. J., Collier, H. O. J., and Vane, J. R. Release of catecholamines in the guinea pig by substances involved in anaphylaxis. *Nature* 213:838, 1967.
59. Pride, N. B., Permutt, S., Riky, R. L., and Bromberger-Barnea, B. Determinants of maximal expiratory flow from the lungs. *J. Appl. Physiol.* 23:646, 1967.
60. Reed, C. E., Busse, W. W., and Lee, T. P. Adrenergic mechanisms and the adenylyl cyclase system in atopic dermatitis. *J. Invest. Dermatol.* 67:333, 1976.
61. Rose, B. Histamine, hormones, and hypersensitivity. *J. Allergy* 25:168, 1954.
62. Rosenberg, E. B., Whalen, G. E., Bennich, H., and Johansson, S. G. O. Increased circulating IgE in a new parasitic disease—Human intestinal capillariasis. *N. Engl. J. Med.* 283:1148, 1970.
63. Rothwell, T. L. W., Prichard, R. K., and Love, R. J. Studies on the role of histamine and 5-hydroxytryptamine in immunity against the nematode *Trichostrongylus colubriformis*: I. In vivo and in vitro effects of the amines. *Int. Arch. Allergy Appl. Immunol.* 46:1, 1974.
64. Rubin, R. H., Austen, K. F., and Goetzel, E. J. Studies of immediate hypersensitivity in a patient with *Acanthocheilonema perstans* filarial infection. *J. Infect. Dis.* 131 (Suppl.):S98, 1975.
65. Sadun, E. H. Homocytotropic Antibody Response to Parasitic Infection. In E. J. L. Soulsby (Ed.), *Immunity to Animal Parasites*. New York: Academic, 1972.
66. Salvato, G. Mast cells in bronchial connective tissue in man: Their modifications in asthma and after treatment with the histamine liberator 48/80. *Int. Arch. Allergy Appl. Immunol.* 18:348, 1961.
67. Scadding, J. G. The Definition of Asthma: General Introduction. In R. Porter and J. Birch (Eds.), *Identification of Asthma*. Edinburgh and London: Churchill Livingstone, 1971.
68. Sherman, W. B. The Atopic Diseases—Introduction. In M. Samter (Ed.), *Immunological Diseases* (2nd ed.). Boston: Little, Brown, 1971.
69. Soter, N. A., and Austen, K. F. The diversity of mast cell-derived mediators: Implications for acute, subacute and chronic cutaneous inflammatory disorders. *J. Invest. Dermatol.* 67:313, 1976.
70. Soter, N. A., Wasserman, S. I., and Austen, K. F. Cold urticaria: Release into the circulation of histamine and eosinophil chemotactic factor of anaphylaxis (ECF-A) during cold challenge. *N. Engl. J. Med.* 294:687, 1976.
71. Szentivanyi, A. The beta adrenergic theory of the atopic abnormality in bronchial asthma. *J. Allergy* 42:203, 1968.
72. Taylor, G., and Shivalkar, P. R. "Arthus-type" reactivity in the nasal airways and skin in pollen sensitive subjects. *Clin. Allergy* 1:407, 1971.
73. Townley, R. G., Ryo, U. Y., Kolotkin, B. M., and Kang, B. Bronchial sensitivity to methacholine in current and former asthmatic and allergic rhinitis patients and control subjects. *J. Allergy Clin. Immunol.* 56:429, 1975.
74. Uvnäs, B., Diamant, B., Hogberg, G., and Thon, I. L. Mechanisms of mast cell disruption induced by a principle extracted from *Ascaris suis*. *Am. J. Physiol.* 199:575, 1960.
75. von Pirquet, C. Allergie. *Munch. Med. Wochenschr.* 30:1457, 1906. [English translation: Allergy. In P. G. H. Gell and R. R. A. Coombs (Eds.), *Clinical Aspects of Immunology* (2nd ed.). Oxford: Blackwell, 1968.]
76. Wasserman, S. I., and Austen, K. F. Arylsulfatase B of human lung: Isolation, characterization, and interaction with slow reacting substance of anaphylaxis. *J. Clin. Invest.* 57:738, 1976.
77. Wasserman, S. I., Goetzel, E. J., and Austen, K. F. Inactivation of slow reacting substance of anaphylaxis by human eosinophil arylsulfatase. *J. Immunol.* 114:645, 1975.
78. Zeiger, R. S., Yurdin, D. L., and Colten, H. R. Histamine metabolism: II. Cellular and subcellular localization of catabolic enzymes. *J. Allergy Clin. Immunol.* 58:172, 1976.