## The World of the Cell

Seventh Edition

# 细胞世界。

Wayne M. Becker Lewis J. Kleinsmith Jeff Hardin Gregory Paul Bertoni

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国外生命科学优秀教材

# 细胞世界

(影印版)

### The World of the Cell

seventh edition

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### 内容简介

本书由美国威斯康星大学、密歇根大学 4 位教授合作编写,在世界上享有盛誉,是细胞生物学学科经典教材之一。本书在亚马逊专业教材销售排行榜长期名列前茅,读者评价较高,并被许多北美、欧洲高校教学选用。

本书编写内容全面、理念先进,并具有鲜明的教学使用特色——适当的深度与简明性、艺术化教学、多层次解答问题、力求精准的概念阐述、为提高教学与学习效率而设计的诸多辅助学习内容。

本书适合生命科学相关专业教学选用,也可供从业人员参考使用。

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### **Guide to Techniques and Methods**

The following techniques are important to cell biologists. Each technique is described in the text at the indicated location.

#### CELL CYCLE

Cell fusion: Chapter 19 (pp. 584–85; Fig. 19-32); Chapter 24 (pp. 775–76; Fig. 24-16)

Flow cytometry: Chapter 19 (p. 553)

Micropipetting: Chapter 19 (Fig. 19-33 on p. 586, cytoplasm transfer); Chapter 23 (Box 23A on p. 725, nuclear transfer)

Temperature-sensitive mutants: Chapter 19 (p. 559)

### **CELL SIGNALING**

Antagonists and agonists for studying receptor-ligand interactions: Chapter 14 (p. 394)

Calcium indicators (to measure cytosolic calcium concentrations): Chapter 14 (pp. 401–2; Fig. 14-11)

Calcium ionophores (to induce ion movement): Chapter 14 (pp. 401–2) Dominant negative mutations (to study growth factor receptor function):

Chapter 14 (p. 412; Fig. 14-20)
Fluorescence resonance energy transfer (FRET) for studying Ras: Appendix (Fig. A-22c)

Using genetic analysis to identify components of signaling pathways: Chapter 14 (Box 14A on pp. 410–11)

#### **CLINICAL TECHNIQUES**

Administration of ethanol as an antidote for methanol poisoning: Chapter 9 (pp. 248–49; Prob. 9-6)

#### CYTOSKELETON

Drugs and mutations as tools for studying cytoskeletal function: Chapter 15 (pp. 431–32 and 438–39; Table 15-3 on p. 432)

Myosin S1 decoration to determine actin polarity: Chapter 15 (p. 438; Fig. 15-14)

Use of microscopy techniques in studying the cytoskeleton: Chapter 15 (Table 15-2 on p. 428)

#### **ENERGETICS**

Calculation of  $\Delta E$  (change in internal energy) and  $\Delta H$  (change in enthalpy): Chapter 5 (p. 113)

Calculation of  $\Delta G'$  (free energy change) and  $\Delta G^{\circ}$  (standard free energy change) for chemical reactions: Chapter 5 (pp. 120–24; Table 5-1 on p. 123)

Calculation of  $\Delta G'$  (free energy change) for transport of charged and uncharged solutes across membranes: Chapter 8 (pp. 217–18; Table 8-4 on p. 218)

Calculation of pmf (proton motive force): Chapter 10 (pp. 278–79) Disruption and reconstitution of mitochondria (to demonstrate the presence of a mitochondrial ATP synthase): Chapter 10 (p. 279; Fig. 10-17)

#### **ENZYME KINETICS**

Analysis of competitive and noncompetitive inhibition: Chapter 6 (p. 145; Fig. 6-14)

Determination of  $K_{\rm m}$  (Michaelis constant) and  $V_{\rm max}$  (maximum velocity): Chapter 6 (pp. 143–44; Figs. 6-12 and 6-13)

Use of the Lineweaver-Burk, Eadie-Hofstee, and Hanes-Woolf plots to linearize enzyme kinetics data: Chapter 6 (pp. 142–43; Figs. 6-10 and 6-11; Prob. 6-12)

#### **GENETICS**

Cis-trans test with partially diploid bacteria: Chapter 23 (pp. 715–16; Table 23-2)

Cotransductional mapping: Chapter 20 (p. 621)

Genetic analysis by Mendelian crosses: Chapter 20 (pp. 612–15; Figs. 20-10, 20-11, and 20-12)

Genetic mapping by recombinant frequencies: Chapter 20

Gene therapy: Chapter 8 (Box 8B on pp. 208–9); Chapter 20 (p. 639) Knockout mice (for studying gene function): Chapter 20 (pp. 628–29)

Using genetic analysis to identify components of signaling pathways: Chapter 14 (Box 14A on pp. 410–11)

#### LABELING AND STAINING

Affinity labeling of proteins: Chapter 7 (Box 7A on pp. 182-84)

Autoradiography: Chapter 12 (pp. 339–40; Fig. 12-10); Chapter 18 (p. 522); Appendix (p. A-17)

Cytochemical localization of enzyme activity: Chapter 4 (Fig. 4-18); Chapter 12 (p. 352; Fig. 12-20)

Electron-opaque tracers (to demonstrate that tight junctions create a permeability barrier): Chapter 17 (p. 487; Fig. 17-10)

Fluorescent antibodies (to identify specific membrane proteins): Chapter 7 (pp. 187–88; Fig. 7-28); Appendix (pp. A-9–10; Fig. A-12)

Fluorescent dyes (to demonstrate gap junction permeability): Chapter 17 (pp. 505–6; Prob. 17-9 and Fig. 17-27)

Green fluorescent protein (GFP): Appendix (p. A-10; Fig. A-14)

Immunochemical staining: Chapter 12 (p. 334; Fig. 12-5)

Pulse-chase experiments: Chapter 23 (p. 747)

Radioisotopes (to trace metabolic fates of atoms): Chapter 1 (p. 9); (to distinguish between proteins on the inner and outer surfaces of membranes): Chapter 7 (p. 184; Fig. 7-24; Probs. 7-11 and 7-12)

#### MEMBRANES AND MEMBRANE TRANSPORT

Affinity labeling of membrane proteins: Chapter 7 (Box 7A on pp. 182–84)

Antibodies against specific cell-surface molecules (to identify specific proteins involved in cell-cell adhesion): Chapter 17 (p. 481)

Cell fusion (to study mobility of membrane proteins): Chapter 7 (pp. 187–88; Fig. 7-28)

Differential scanning calorimetry (to determine transition temperatures of membranes): Chapter 7 (pp. 169–70; Fig. 7-12)

Experimental evidence that discredited Davson-Danielli model: Chapter 7 (pp. 160–61)

Fluorescence recovery after photobleaching (to study mobility of membrane lipids and proteins): Chapter 7 (p. 169; Fig. 7-11)

Fluorescence microscopy (to study passage of molecules from one cell to another via gap junctions): Chapter 17 (pp. 505–6; Prob. 17-9 and Fig. 17-27)

Freeze-fracture microscopy (to study membrane proteins and their mobility and to visualize tight junctions): Chapter 7 (pp. 173–75; Figs. 7-16, 7-17, and 7-18; p. 188; Fig. 7-29); Chapter 17 (pp. 487–88; Fig. 17-9)

Hydropathic analysis of membrane proteins (to identify possible transmembrane segments): Chapter 7 (pp. 180 and 192; Figs. 7-23 and 7-30; Prob. 7-10)

Inverted membrane vesicles (to study membrane transport): Chapter 7 (p. 193; Prob. 7-12); Chapter 8 (p. 222; Prob. 8-11)

Liposomes (to study membrane proteins and membrane permeability): Chapter 8 (p. 199)

Microelectrodes (to demonstrate flow of electric current from one cell to another via gap junctions): Chapter 17 (pp. 505–6; Prob. 17-9 and Fig. 17-27)

Patch clamping: Chapter 13 (pp. 371-72; Fig. 13-5)

Radioisotope labeling procedures (to distinguish between proteins exposed on the inner and outer surfaces of membranes): Chapter 7 (p. 184; Fig. 7-24; pp. 192–93; Probs. 7-11 and 7-12)

SDS-polyacrylamide gel electrophoresis, including two-dimensional (to separate proteins): Chapter 7 (pp. 178–79; Fig. 7-22)

Site-specific mutagenesis (to modify membrane proteins genetically): Chapter 7 (Box 7A on pp. 182–84; Fig. 7A-1)

Thin-layer chromatography (TLC) for analysis of membrane lipids: Chapter 7 (pp. 166–67; Fig. 7-9)

Tonicity changes (effects of hypertonicity and hypotonicity on osmotic movement of water in cells): Chapter 8 (Box 8A on pp. 200–1; Fig. 8A-1)

Voltage recording: Chapter 13 (p. 375; Fig. 13-9)

#### MICROSCOPY

Electron microscopy: Chapter 1 (pp. 8–9); Appendix (pp. A-17–26) Autoradiography: Chapter 12 (pp. 339–40; Fig. 12-10); Appendix (pp. A-17 and A-21)

CryoEM: Appendix (pp. A-28-29)

Earliest use by biologists: Chapter 1 (p. 6)

Electron tomography (3-D): Appendix (pp. A-25-26)

Freeze etching: Appendix (pp. A-24-25)

Freeze fracturing: Chapter 1 (p. 8); Chapter 7 (pp. 173–75; Figs. 7-16, 7-17, and 7-18; p. 188; Fig. 7-29): Appendix (pp. A-24–25; Figs. A-34 and A-35)
High-voltage electron microscopy: Appendix (p. A-19)
Immunoelectron microscopy (immunoEM): Appendix (p. A-21;

Fig. A-29)

Negative staining: Chapter 1 (p. 8); Chapter 10 (pp. 257–58; Fig. 10-5); Appendix (p. A-22; Fig. A-31)

Sample preparation techniques for electron microscopy: Appendix (pp. A-20–26)

Sectioning: Appendix (pp. A-17 and A-20–21; Figs. A-24 and A-28) Shadowing: Appendix (pp. A-23–24; Figs. A-32 and A-33) Staining: Appendix (p. A-17)

Scanning electron microscopy: Chapter 1 (p. 8; Fig. 1-4); Appendix (pp. A-19–20; Figs. A-26b and A-27)

Stereo electron microscopy: Chapter 1 (pp. 8–9); Appendix (pp. A-25–26; Fig. A-36a)

Transmission electron microscopy: Chapter 1 (p. 8); Appendix (pp. A-18–19; Figs. A-25 and A-26a)

Light microscopy: Chapter 1 (pp. 6–8; Table 1-1); Appendix (pp. A-5–17; Fig. A-5)

Autoradiography: Appendix (p. A-17)

Brightfield microscopy: Chapter 1 (p. 7; Table 1-1); Appendix (p. A-6)

Confocal microscopy: Chapter 1 (p. 7; Table 1-1); Appendix (pp. A-11–13; Figs. A-15 and A-17)

Digital deconvolution microscopy: Chapter 1 (p. 8); Appendix (pp. A-13–14; Fig. A-19)

Digital microscopy: Chapter 1 (p. 8); Appendix (pp. A-14–15; Fig. A-20)

Differential interference contrast (DIC) microscopy: Chapter 1 (p. 7; Table 1-1); Appendix (pp. A-7–8; Figs. A-8 and A-9)

Earliest use by biologists: Chapter 1 (pp. 1–3) Fixation: Appendix (p. A-17)

Fluorescence microscopy: Chapter 1 (p. 7; Table 1-1); Appendix (pp. 8–11; Figs. A-11–15)

Fluorescence recovery after photobleaching (FRAP): Chapter 7 (p. 169; Fig. 7-11); Appendix (p. A-15)

Fluorescence resonance energy transfer (FRET): Appendix (p. A-16, Fig. A-22)

Fluorescent probes: Appendix (pp. A-8-11)

Green fluorescent protein (GFP): Appendix (p. A-10; Fig. A-14) Immunofluorescence microscopy: Appendix (pp. A-9–10; Fig. A-12) Multiphoton excitation microscopy: Appendix (p. A-13; Fig. A-18) Optical "tweezers": Appendix (pp. A-16–17; Fig. A-23)

Phase-contrast microscopy: Chapter 1 (p. 7; Table 1-1); Appendix (pp. A-6–7; Figs. A-6, A-7)

Photoactivation: Appendix (p. A-15; Fig. A-21b)

Sectioning: Appendix (p. A-17; Fig. A-24)

Staining: Appendix (p. A-17)

Total internal reflection fluorescence microscopy: Appendix (p. A-15; Fig. A-21c)

Optical principles of microscopy: Appendix (pp. A-1–5; Fig. A-1) Scanning probe microscopy: Appendix (pp. A-26–27)

Atomic force microscope: Appendix (p. A-27)

Scanning tunneling microscope: Appendix (pp. A-26–27; Fig. A-37)

#### NUCLEIC ACIDS AND RECOMBINANT DNA

Bacterial artificial chromosomes (BACs): Chapter 20 (p. 636)

BLAST searching: Chapter 18 (p. 527)

Cloning of genes: Chapter 20 (pp. 629–36; Figs. 20-26, 20-27, 20-28, 20-31) Chromatin immunoprecipitation (ChIP) assay: Chapter 21 (Box 21B on p. 658)

Cloning of organisms: Chapter 23 (Box 23A on p. 725)

Colony hybridization with nucleic acid probe: Chapter 20 (p. 633; Fig. 20-29)

cDNA molecules for transcription studies: Chapter 23 (p. 733; Fig. 23-20) cDNA preparation (reverse transcription): Chapter 20 (p. 634; Fig. 20-30) DNA denaturation and renaturation: Chapter 18 (pp. 518–20; Fig. 18-10)

DNA fingerprinting: Chapter 18 (Box 18C on pp. 530-31)

DNA microarrays: Chapter 23 (p. 733, Fig. 23-20)

DNA sequencing: Chapter 18 (pp. 524–25; Fig. 18-14)

DNase sensitivity of active genes in chromatin: Chapter 23 (pp. 729–30; Fig. 23-17)

Electrophoresis of DNA: Chapter 18 (pp. 521-22; Fig. 18-12)

Equilibrium density centrifugation of DNA: Chapter 19 (pp. 553–54; Fig. 19-3)

Footprinting technique for detecting protein-binding sites on DNA: Chapter 21 (Box 21B on p. 658)

Hybridization of nucleic acids: Chapter 18 (pp. 519-20)

Northern blotting: Chapter 23 (p. 733)

Nuclease digestion of chromatin (to isolate nucleosomes): Chapter 18 (pp. 533–34; Fig. 18-19)

Polymerase chain reaction (PCR): Chapter 19 (Box 19A on pp. 562–63); use in gene cloning and gene modification by epitope or polyhistidine tagging: Chapter 20 (p. 636)

Restriction enzymes: Chapter 18 (Box 18B on pp. 522-23)

Restriction mapping of DNA: Chapter 18 (pp. 522-24; Fig. 18-13)

RNA interference (for silencing genes to study gene function or treat diseases): Chapter 23 (p. 749)

Run-on transcription assay: Chapter 23 (pp. 732–33; Fig. 23-19)

Southern blotting: Chapter 18 (p. 531)

Temperature-sensitive mutants (DNA replication): Chapter 19 (p. 559)

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Transgenic plants: Chapter 20 (p. 637; Fig. 20-32)

Yeast artificial chromosomes (YACs): Chapter 20 (pp. 635–36; Fig. 20-31) X-ray crystallography: Appendix (pp. A-27–28; Fig. A-39)

#### **PROTEINS**

Affinity labeling of proteins: Chapter 7 (Box 7A on pp. 182–84)
Antibodies against specific cell-surface molecules (to identify specific proteins involved in cell-cell adhesion): Chapter 17 (p. 481)
BLAST searching: Chapter 18 (p. 527)

Hydropathic analysis of proteins (to identify possible transmembrane segments): Chapter 7 (pp. 180 and 192; Figs. 7-23 and 7-30; Prob. 7-10)

Monoclonal antibodies: Chapter 24 (Box 24B on p. 787)

Protein denaturation and renaturation: Chapter 2 (pp. 32–33; Fig. 2-18a) Protein sequencing: Chapter 3 (pp. 47–48)

SDS-polyacrylamide gel electrophoresis, including two-dimensional: Chapter 7 (pp. 178–79; Fig. 7-22)

Western blotting: Chapter 7 (pp. 178-179)

X-ray crystallography: Chapter 6 (p. 137; Fig. 6-6); Chapter 7 (p. 180); Chapter 11 (p. 302; Fig. 11B-1); Appendix (pp. A-27–28; Fig. A-39)

#### SCIENTIFIC METHOD

Use of scientific method: Chapter 1 (Box 1B on p. 12)

### SEPARATION OF CELLS, ORGANELLES, AND MOLECULES

Density gradient centrifugation: Chapter 12 (pp. 330–31; Fig. 12A-5) Differential centrifugation: Chapter 12 (pp. 329–30; Figs. 12A-2 and 12A-4); used to isolate lysosomes: Chapter 4 (Box 4B on pp. 92–93)

Equilibrium density centrifugation: Chapter 12 (pp. 331 and 356); Chapter 19 (pp. 553–54; Fig. 19-3)

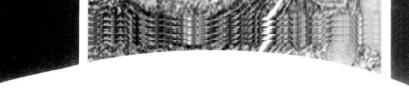
Gel electrophoresis of DNA: Chapter 18 (p. 521-22; Fig. 18-12)

SDS-polyacrylamide gel electrophoresis as a means of separating proteins: Chapter 7 (pp. 178–79; Fig. 7-22)

Subcellular fractionation: Chapter 12 (Box 12A on pp. 328-31)

Thin-layer chromatography (TLC) for analysis of membrane lipids: Chapter 7 (pp. 166–67; Fig. 7-9)

Ultracentrifugation, earliest use by biologists: Chapter 1 (p. 9); used to achieve high g forces: Chapter 12 (p. 328)



### About the Authors



WAYNE M. BECKER taught cell biology at the University of Wisconsin-Madison, for 30 years until his recent retirement. His interest in text-book writing grew out of notes, outlines, and problem sets that he assembled for his students, culminating in *Energy and the Living Cell*, a paperback text on bioenergetics published in 1977, and *The World of the Cell*, the first edition of which appeared in 1986. He earned all his

degrees at the University of Wisconsin-Madison. All three degrees are in biochemistry, an orientation that is readily discernible in his textbooks. His research interests have been in plant molecular biology, focused specifically on the regulation of the expression of genes that encode enzymes of the photorespiratory pathway. His interests in teaching, learning, and research have taken him on sabbatical leaves at Harvard University, Edinburgh University, the University of Indonesia, the University of Puerto Rico, Canterbury University in Christchurch, New Zealand, the Chinese University of Hong Kong, and the Charles University in Prague. His honors include a Chancellor's Award for Distinguished Teaching, Guggenheim and Fulbright Fellowships, and a Visiting Scholar Award from the Royal Society of London.



**LEWIS J. KLEINSMITH** is an Arthur F. Thurnau Professor Emeritus of Molecular, Cellular, and Developmental biology at the University of Michigan, where he has served on the faculty since receiving his Ph.D. from Rockefeller University in 1968. His teaching experiences have involved courses in introductory biology, cell biology, and cancer biology, and his research interests have

included studies of growth control in cancer cells, the role of protein phosphorylation in eukaryotic gene regulation, and the control of gene expression during development. Among his numerous publications, he is the author of *Principles of Cancer Biology* as well as several award-winning educational software programs. His honors include a Guggenheim Fellowship, the Henry Russell Award, a Michigan Distinguished Service Award, citations for outstanding teaching from the Michigan Students Association, an NIH Plain Language Award, and a Best Curriculum Innovation Award from the EDUCOM Higher Education Software Awards Competition.

JEFF HARDIN is a Professor in the Zoology Department at the University of Wisconsin-Madison. His research interests center on how cells migrate and adhere to one another to change the shape of animal embryos. Dr. Hardin's teaching is enhanced by his extensive use of videomicroscopy and his Web-based teaching materials, which are used on many campuses in the United



States and other countries. As part of his interest in teaching biology, Dr. Hardin has been involved in several teaching initiatives. He was a founding member of the University of Wisconsin Teaching Academy and a cofounder of a University of Wisconsin system-wide instructional technology initiative known as BioWeb. He is currently faculty director of the Biology Core Curriculum, a four-semester honors biology sequence for undergraduates. His teaching awards include a Lily Teaching Fellowship and a National Science Foundation Young Investigator Award. He is also on the editorial board of *CBE: Life Sciences Education*.

GREGORY PAUL BERTONI, the newest member of the author team, has been active in teaching and research for over 20 years. He earned a Ph.D. in Cellular and Molecular Biology from the University of Wisconsin-Madison, where his teaching experiences included introductory and graduate-level biochemistry, sophomore cell biology, and plant physiology. He also



helped to develop a new course entitled "Ways of Knowing" designed to introduce entering freshmen to the learning process itself. His published research includes studies in bacterial pathogenesis, plant-microbe interactions, and plant gene expression. He is currently teaching biology and medical microbiology at Columbus State Community College in Columbus, Ohio, where he has just been nominated for a Distinguished Teaching Award. In addition, Dr. Bertoni is a freelance scientific writer who has contributed to text- and web-based projects in biology, physics, and microbiology. He is also a science editor for *The Plant Cell*, a leading research journal in plant biology and molecular biology.

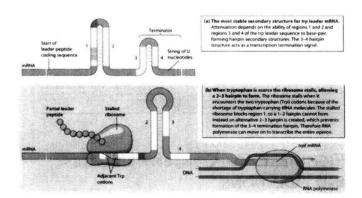
### **Preface**

ecause we enjoy interacting with biology undergrads and think that they should have biology textbooks that are clearly written, make the subject matter relevant to the reader, and help them appreciate not only how much we already know about biology—cell biology, in our case—but also how much more remains to be investigated and discovered." That's how any of the authors of this text would likely respond if asked why we've invested so much time in writing and revising The World of the Cell. Each of us has an extensive history of teaching undergraduate courses in cell biology and related areas, and each of us treasures our contact with students as one of the most rewarding aspects of being a faculty member.

As we reflect on the changes we've seen in our courses over the years, we realize that the past several decades have seen an explosive growth in our understanding of the properties and functions of living cells. This enormous profusion of information presents us with a daunting challenge as we confront the task of keeping *The World of the Cell* up to date while simultaneously ensuring that it remains both manageable in length and readily comprehensible to students encountering the field of cell and molecular biology for the first time. This seventh edition represents our most recent attempt to rise to that challenge. As with the previous editions, each of us has brought our own teaching and writing experience to the venture in ways that we have found mutually beneficial—a view that we hope our readers will share.

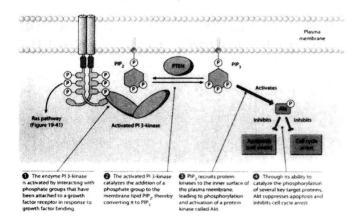
One major objective for this edition has been to update the content of the text, especially in areas where the pace of research is especially brisk and recent findings are particularly significant. At the same time, we have remained committed to the three central goals that have characterized each preceding edition. As always, our primary goal is to introduce students to the fundamental principles that guide cellular organization and function. Second, we think it is important for students to understand some of the critical scientific evidence that has led to the formulation of these central concepts. And finally, we have sought to accomplish these goals in a book of manageable length that can be easily read and understood by beginning cell biology students—and that still fits in their backpacks! To accomplish this third goal, we have necessarily been selective both in the types of examples chosen to illustrate key concepts and in the quantity of scientific evidence included. We have, in other words, attempted to remain faithful to the overall purpose of each previous edition: to present the essential principles, processes, and methodology of molecular and cell biology as lucidly as possible. We have also given careful attention to accuracy, consistency, vocabulary, and readability to minimize confusion and maximize understanding for our readers.

Another objective for the seventh edition has been to enhance the effectiveness of the art program through the widespread introduction of *minicaptions* to help students understand technically complex diagrams. A minicaption is a pedagogical tool that takes detailed textual descriptions previously found in lengthy figure legends and incorporates the information directly into the illustration itself. With diagrams that use minicaptions, students no longer need to look back and forth between the figure legend and the illustration to understand what is taking place. Two types of minicaptions have been widely introduced into this edition of *The World of the Cell:* **Text Box Minicaptions** are colored text boxes that describe events taking place in various locations within a diagram.

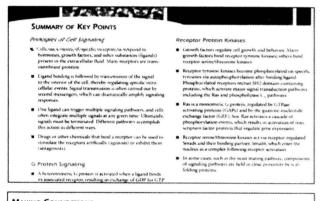


Step Circle Minicaptions are consecutively numbered, brief blocks of colored text that point to various steps within a sequential pathway and describe the events occurring at each step. These devices are designed to help students grasp concepts more quickly by drawing their focus into the body of an illustration rather than

depending solely on a separate figure legend to describe what is taking place.



Two other features new to the seventh edition provide an overview of the key points covered in each chapter and highlight connections between material covered in different chapters. These two pedagogical tools, which appear at the end of each chapter, are a bulleted Summary of Key **Points** that briefly describes the main points covered in each section of the chapter and a Making Connections paragraph that highlights connections between the content of the current chapter and topics covered elsewhere in the book.



### MAKING CONNECTIONS

In this chapter, you learned that a crucial feature of cells in a mul-ticellular organism is the ability to send and receive ignals. Some of these ignals, such as growth fisters, as at very short range, much like the neutotransmitters we examined in Chapter 13. Others, such as hortimones, ast a long range Jeneum most received softers much as hortimones, and stop range Jeneum most received starface must be transmitted deep insule the cell. In some cases, the relay system involves cascades of phosphorylation, a topic you first learned about in Chapter Seel liquiding controls a vast array of cellular processes—from glucose utilization, which feed-

into the metabolic pathways you learned about in Chapters 9 and 10, to endo- and exocytors, which you learned about in Chapter 12, to embryonic development and cell death. One tight controlled process regulated by cell inpating in the cell cycle, which you will learn much more about in Chapter 19. When such signaling gost eavers, cancer an easil, in Chapter 14 you will learn more about this process. The assembly and disassembly of the cytosleckton are a set of crossel processes regulated by cell signaling, we turn to these processes in the next two chapters.

### HIGHLIGHTS OF CONTENT UPDATES AND ADDITIONS

Updated material and new information has been added throughout the book. Topics that have been altered, updated, or added include the following:

CHAPTER 1: diversity of cell types, cell biology timeline, deconvolution microscopy, green fluorescent protein

CHAPTER 3: trans fats and health, hair structure

CHAPTER 4: relationship between archaea, bacteria, and eukaryotes, actin-like and tubulin-like cytoskeletal proteins in bacteria, bacterial cell wall composition

CHAPTER 5: oxidation-reduction, autotrophs and heterotrophs, coupled reactions

CHAPTER 6: quantum tunneling mechanism of certain enzymes, relationship of enzyme cofactors to dietary needs for vitamins and metals, use of substrate analogs to fight diseases such as AIDS

CHAPTER 7: involvement of adhesive surface proteins and glycocalyx in bacterial pathogenesis, effect of dietary trans fats on membrane structure, two-dimensional electrophoresis, Western blotting, antifungal antibiotics

CHAPTER 8: antibiotic resistance arising from mutations in porins or from acquisition of multiple drug resistance proteins, effect of cellular pH on transport of oxygen by hemoglobin

CHAPTER 9: novel roles of glycolytic enzymes, entropy contribution to the exergonic nature of ATP hydrolysis

CHAPTER 10: mitochondrial structure and cristae junctions, ATP synthase structure and mechanism of action, Q cycle, respirasomes

CHAPTER 11: mobility of light harvesting complexes, chloroplast Q cycle, similarities between chloroplast and mitochondrial electron transport, chloroplast ATP synthase, overview of Calvin cycle

CHAPTER 12: endomembrane trafficking of proteins and lipids, pharmacogenetics, steroid biosynthesis, ER retention tags on NMDA receptor, autophagy and cancer, desensitization and tumor formation, tolerance to barbiturates, caveolae and cholesterol uptake, phagocytosis by white blood cells to educate the immune system

CHAPTER 13: Goldman equation relationship to the Nernst equation, direct (ionotropic) and indirect (metabotropic) mechanism of neurotransmitter action, nicotinic acetylcholine receptors

CHAPTER 14: signal integration, opioid receptors, G protein-linked receptor kinases,  $\beta$ -arrestin, protein kinase A, regulators of G protein signaling, G<sub>By</sub> subunit signaling, metabotropic neurotransmitter receptors, yeast mating pathway and scaffolding complexes, insulin signaling

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CHAPTER 16: kinesin and myosin families, primary ("9 + 0") cilia, cell crawling and focal adhesions, cyclosis

CHAPTER 17: tissue organization, human disorders linked to the extracellular matrix, cell-cell adhesion, or junctions

CHAPTER 18: bioinformatics, transcriptomes, BLAST searching, haplotypes, copy number variants, transposable elements (LINEs and SINEs), plasmid functions, nuclear substructures

CHAPTER 19: flow cytometry, bacterial cell division, telomere shortening and aging, spindle assembly in cells lacking centrosomes, role of Ran-GTP in spindle assembly, Rho and the contractile ring, TOR signaling and cell growth

CHAPTER 20: parthenogenesis in Komodo dragons, cytostatic factor (CSF), pairing sites in synapsis, knockout mice, use of PCR for gene cloning, epitope and polyhistidine tagging

CHAPTER 21: RNA proofreading, transcription factories, archaeal promoters, exon-junction complex, alternative splicing, the "one gene-many polypeptides" concept

CHAPTER 22: initiation factors for eukaryotic translation, hybrid states during translocation step of protein synthesis, peptide anticodons, similarities between archaeal and eukaryotic translation, unfolded protein response, ER-associated degradation (ERAD)

CHAPTER 23: regulons and stimulons, cloning of carrot plants, genomic imprinting, DNA methylation, histone methylation, DNA insulators, translational control by eIF2 and eIF4F, pulse-chase technique, mRNA degradation, P bodies, RNA interference-based disease treatments, developmental role of microRNAs

CHAPTER 24: epigenetic changes in cancer cells, cancer stem cells, immune surveillance, BRCA genes, HPV vaccine, microRNAs in cancer, genetic basis of aneuploidy

APPENDIX: 3-D electron tomography, multichannel fluorescent imaging

### BUILDING ON THE STRENGTHS OF PREVIOUS EDITIONS

We have retained and built upon the strengths of prior editions in four key areas:

### 1. The chapter organization focuses on main concepts.

- Each chapter is divided into sections that begin with a *concept statement heading*, which summarizes the material and helps students focus on the main points to study and review.
- Chapters are written and organized in ways that allow instructors to assign the chapters and chapter sections in different sequences to make the book adaptable to a wide variety of course plans.

- **NEW:** Each chapter culminates with a bulleted *Summary of Key Points* that briefly describes the main points covered in each section of the chapter.
- NEW: Each Summary of Key Points is followed by a Making Connections paragraph that highlights some of the interrelationships that connect the content of the current chapter to topics covered elsewhere in the book.

### 2. The illustrations teach concepts at an appropriate level of detail.

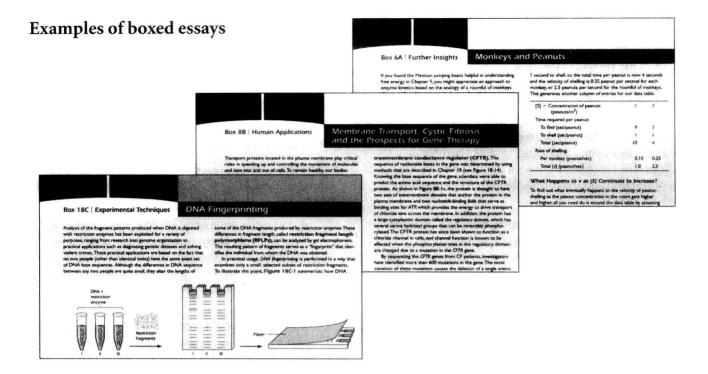
- **NEW**: Many of the more complex figures now incorporate *minicaptions* (described earlier) to help students grasp concepts more quickly by drawing their focus into the body of an illustration rather than depending solely on a separate Figure Legend to describe what is taking place.
- Overview figures outline complicated structures or processes in broad strokes and are followed by text and figures that present supporting details.
- Carefully-selected micrographs are usually accompanied by size bars to indicate magnification.

### 3. Important terminology is highlighted and defined in several ways.

- **Boldface type** is used to highlight the most important terms in each chapter, all of which are defined in the Glossary.
- *Italics* are employed to identify additional technical terms that are less important than boldfaced terms but significant in their own right. Occasionally, italics are also used to highlight important phrases or sentences.
- The Glossary includes definitions and page references for all bold-faced key terms and acronyms in every chapter—more than 1500 terms in all, a veritable "dictionary of cell biology" in its own right.

### 4. Each chapter helps students learn the process of science, not just facts.

- Text discussions emphasize the experimental evidence that underlies our understanding of cell structure and function, to remind readers that advances in cell biology, as in all branches of science, come not from lecturers in their classrooms or textbook authors at their computers but from researchers in their laboratories.
- The inclusion of a *Problem Set* at the end of each chapter reflects our conviction that we learn science not just by reading or hearing about it, but by working with it. The problems are designed to emphasize understanding and application, rather than rote recall. Many of the problems are classtested, having been selected from problem sets



and exams we have used in our own courses. To maximize the usefulness of the problem sets, detailed answers for all problems are available for students in a *Solutions Manual* that is available for purchase separately.

- Each chapter contains one or more *Boxed Essays* to aid students in their understanding of particularly important or intriguing aspects of cell biology (see samples at top of page). Some of the essays provide Further Insights into potentially difficult principles, such as the essay that uses the analogy of monkeys shelling peanuts to explain enzyme kinetics (Box 6A). Other essays describe some of the important Experimental Techniques used by cell biologists, as exemplified by the description of DNA fingerprinting in Box 18C. And yet another role of the boxed essays is to describe Human Applications of research findings in cell biology, as illustrated by the discussion of cystic fibrosis and the prospects for gene therapy in Box 8B.
- A Suggested Reading list is included at the end of each chapter, with an emphasis on review articles and carefully selected research publications that motivated students are likely to understand. We have tried to avoid overwhelming readers with lengthy bibliographies of the original literature but have referenced articles that are especially relevant to the topics of the chapter. In most chapters, we have included a few citations of especially important historical publications, which are marked with blue dots to alert the reader to their historical significance.

### TECHNIQUES AND METHODS

Throughout the text, we have tried to explain not only what we know about cells but also how we know what we know. Toward that end, we have included descriptions of experimental techniques and findings in every chapter, almost always in the context of the questions they address and in anticipation of the answers they provide. For example, polyacrylamide gel electrophoresis is introduced not in a chapter that simply catalogues a variety of methods for studying cells but in Chapter 7, where it becomes important to our understanding of how membrane proteins can be separated from one another. Similarly, equilibrium density centrifugation is described in Chapter 12, where it is essential to our understanding of how lysosomes were originally distinguished from mitochondria and subsequently from peroxisomes as well.

To help readers locate techniques out of context, an alphabetical Guide to Techniques and Methods appears on the inside of the front cover, with references to chapters, pages, tables, figures, and boxed essays, as appropriate. To enhance its usefulness, the Guide to Techniques and Methods includes references not just to laboratory techniques but also to the mathematical determination of values such as  $\Delta G$  (free energy change) and  $\Delta E_0'$  (standard reduction potential), bioinformatics techniques such as BLAST searching, and even to clinical procedures such as the treatment of methanol poisoning.

Microscopy is the only exception to our general approach of introducing techniques in context. The techniques of light and electron microscopy are so pervasively relevant to contemporary cell biology that they warrant special consideration as a self-contained unit, which is

included as an Appendix entitled Visualizing Cells and Molecules. This Appendix gives students ready access to detailed information on a variety of microscopy techniques, including cutting-edge uses of light microscopy for imaging and manipulating molecular processes.

### SUPPLEMENTARY LEARNING AIDS

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Written by the authors, this is a collection of complete, detailed answers for all of the end-of-chapter questions and problems.

### The Cell Place for The World of the Cell • 0-321-52744-5

The book's companion website, www.thecellplace.com, helps students explore a variety of cell biology topics in depth, and includes interactive tutorials, simulations, animations, and step-by-step problems. Practice quizzes contain 20 multiple-choice questions for each chapter, with instant feedback for correct and incorrect answers.

### WE WELCOME YOUR COMMENTS AND SUGGESTIONS

The ultimate test of any textbook is how effectively it helps instructors teach and students learn. We welcome feedback and suggestions from readers and will try to acknowledge all correspondence. Please send your comments, criticisms, and suggestions to the appropriate authors listed below.

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The many reviewers listed below provided helpful criticisms and suggestions at various stages of manuscript development and revision. Their words of appraisal and counsel were gratefully received and greatly appreciated. Indeed, the extensive review process to which this and the prior editions of the book have been exposed should be considered a significant feature of the book. Nonetheless, the final responsibility for what you read here remains ours, and you may confidently attribute to us any errors of omission or commission encountered in these pages.

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