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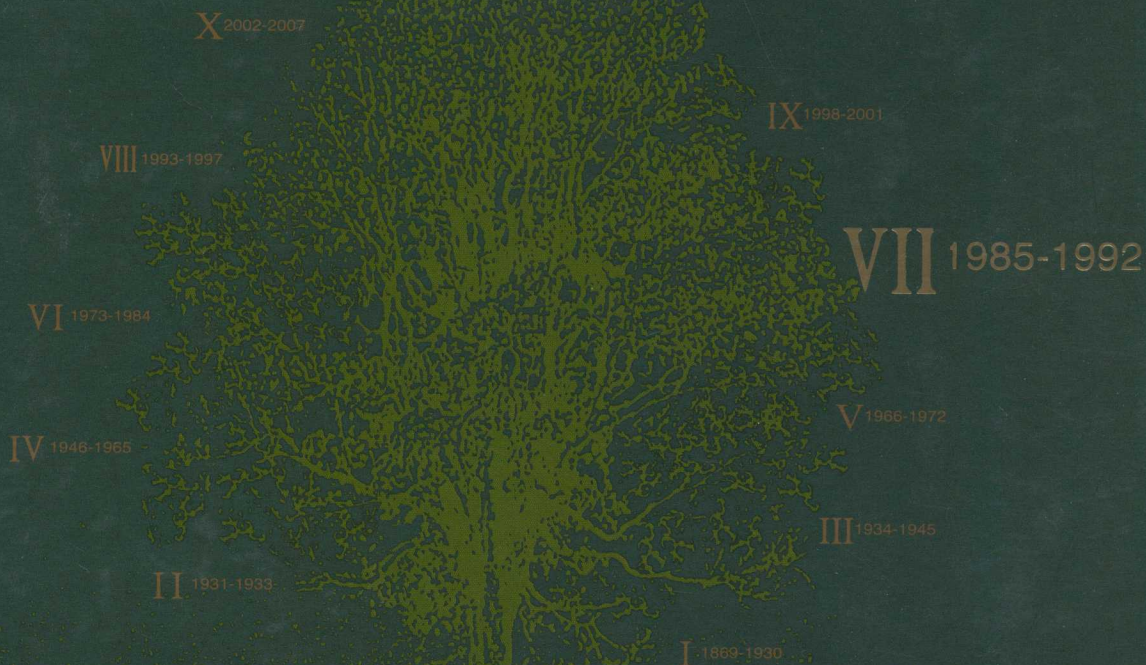
(英汉对照版)

第七卷

总顾问：李政道 (Tsung-Dao Lee)

英方主编：Sir John Maddox
Philip Campbell

中方主编：路甬祥



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Complete Nucleotide Sequence of the AIDS Virus, HTLV-III

L. Ratner *et al.*

Editor's Note

The early 1980s was a period of intense AIDS-related research and discovery. The AIDS-causing virus, then called human T-cell leukaemia virus III (HTLV-III), was identified and isolated, as were its modes of transmission. Here US biologist Robert Gallo and colleagues describe the complete nucleotide sequences of the AIDS virus, a milestone in AIDS research. Overall, the sequence resembles that of other RNA-encoded "retroviruses" containing the three hallmark genes *gag*, *pol* and *env*. But it also contains anomalies, not least the immense heterogeneity between clones. The genome has enabled researchers to define key viral genes and proteins, has shed light on the origins, nature and spread of HIV, and continues to influence diagnostic and drug development.

The complete nucleotide sequence of two human T-cell leukaemia type III (HTLV-III) proviral DNAs each have four long open reading frames, the first two corresponding to the *gag* and *pol* genes. The fourth open reading frame encodes two functional polypeptides, a large precursor of the major envelope glycoprotein and a smaller protein derived from the 3'-terminus long open reading frame analogous to the long open reading frame (*lor*) product of HTLV-I and -II.

HUMAN T-cell leukaemia (lymphotropic) viruses HTLV-I, -II and -III, a family of Hexogenous retroviruses, are associated with T-cell disorders including adult T-cell leukaemia lymphoma (ATLL) and the acquired immune deficiency syndrome (AIDS)¹⁻¹⁴. These viruses share a number of biological and structural features: tropism for OKT4⁺ lymphocytes^{10,15-21}, the ability to produce giant multinucleated cells in culture^{10,17,22-24}, immunological cross-reactivity of some virally encoded proteins^{12,13}, distant nucleic acid sequence similarities^{8,9,25-28} and the preference for magnesium of the viral reverse transcriptase²⁹⁻³¹. Moreover, the genome of HTLV-I and -II as well as the related bovine leukaemia virus (BLV) is somewhat longer than that of other retroviruses^{8,9,25,32-34}. A sequence of 1,600–1,800 nucleotides is interposed between the 3' end of the *env* gene and the long terminal repeat (LTR) sequences of these viruses^{25,32-37}, the 3' portion of which is a long open reading frame (*lor*)^{32,35,37}. Another feature that distinguishes HTLV-I, -II, -III and BLV from other retroviruses is the marked increase in the rate of transcription initiated within the viral LTR sequences in infected compared with uninfected cells^{38,39}. This phenomenon, *trans*-acting transcriptional regulation, is not observed for other retroviruses⁴⁰ and is probably mediated by the *lor* gene product of these retroviruses³⁸.

艾滋病病毒HTLV-III的完整核苷酸序列

拉特纳等

编者按

20世纪80年代早期是艾滋病相关研究和发现的密集时期。人们鉴定并分离出了引发艾滋病的病毒——后被称为T细胞白血病病毒III (HTLV-III) 并弄清了它的传播模式。在本文中, 美国生物学家罗伯特·加洛及其同事描述了艾滋病病毒的完整核苷酸序列, 这是艾滋病研究领域的一块里程碑。这个序列和其他RNA编码的“逆转录病毒”一样, 都具有 *gag*、*pol* 和 *env* 这三个标志基因。但它也有一些不同, 这些不同不仅仅局限于克隆间巨大的遗传异质性。基因组使得研究人员能够定义关键的病毒基因和蛋白, 解释艾滋病的起源、性质和传播, 进而影响诊断和药物的发展。

人类两种T细胞白血病III (HTLV-III) 的前病毒DNA的完整序列各有四个长的开放阅读框, 前两个与 *gag* 和 *pol* 基因相关。第四个开放阅读框编码两个功能多肽, 主要包膜糖蛋白的大前体和来源于3'末端长开放阅读框的小蛋白, 类似于HTLV-I和-II长开放阅读框 (*lor*) 的产物。

人类T细胞白血病(淋巴性)病毒HTLV-I, -II和-III, 属于外源逆转录病毒家族, 它们与T细胞紊乱有关, 包括成人T淋巴细胞白血病(ATLL)和获得性免疫综合征(AIDS) [1-14]。这些病毒具有许多共同的生物学和结构特征: 对OKT4⁺淋巴细胞的嗜性 [10,15-21]、在培养基中产生巨型多核细胞的能力 [10,17,22-24]、与一些病毒编码蛋白的免疫学交叉反应 [12,13]、远亲核苷酸序列之间的相似性 [8,9,25-28] 和病毒逆转录酶对镁的偏好 [29-31]。此外, HTLV-I和-II与相关的牛白血病病毒(BLV)一样比其他的逆转录病毒的基因组都略长 [8,9,25,32-34]。在 *env* 基因的3'末端和这些病毒的长末端重复序列(LTR)之间得到一段1,600~1,800个核苷酸的序列 [25,32-37], 该序列3'部分是一个长的开放阅读框 (*lor*) [32,35,37]。HTLV-I、-II、-III和BLV区别于其他逆转录病毒的另一个特点是, 被感染的细胞中LTR序列区域起始转录的速率比未被感染的细胞有显著增长 [38,39]。这种反式转录调控的现象在其他逆转录病毒中 [40] 没有观察到, 可能是由这些逆转录病毒的 *lor* 基因的产物所介导的 [38]。

Despite the similarity between HTLV-III and the other members of the HTLV-BLV family of viruses, the biology and pathology of HTLV-III differ substantially. Infection with HTLV-III results often in profound immunosuppression (AIDS), consequent on the depletion of the OKT4⁺ cell population^{10-14,41-43}. This effect is mirrored by a pronounced cytopathic rather than transforming effect of HTLV-III infection upon the OKT4⁺ cells in lymphocyte cultures *in vitro*^{10,11,20}. In contrast, infection with HTLV-I results in a low incidence of T-cell leukaemia lymphoma (an OKT4⁺-cell malignancy)¹⁻⁶. There is evidence also for some degree of immunodeficiency in HTLV-I patients^{6,44}. Infection of primary lymphocytes in culture by HTLV-I and -II results in *in vitro* transformation of predominantly OKT4⁺ cells^{45,46}. A cytopathic effect of HTLV-I infection on lymphocytes is apparent, but the effect is not as pronounced as that in HTLV-III^{16,17,45-48}. HTLV-III differs also from HTLV-I and -II in the extent of infectious virion production *in vivo* and *in vitro*. High titres of cell-free infectious virions can be obtained from AIDS patient semen and saliva and from the supernatant of cultures infected with HTLV-III^{10,49-51}. Very few, if any, cell-free infectious virions can be recovered from ATLL patients or from cultures infected with HTLV-I or -II⁵².

To investigate the biological activity of these viruses *in vitro* and *in vivo* and to provide information useful for the development of diagnostic and therapeutic reagents for AIDS, we have determined the complete nucleotide sequence of the HTLV-III provirus.

Genomic Structure of HTLV-III

Several closely related clones of HTLV-III DNA were obtained from the H9 cell line infected with HTLV-III present in the blood pooled from several American AIDS patients¹⁰. The complete primary nucleotide sequence of three unintegrated viral clones of 8.9, 5.3 and 3.6 kilobases (kb) in length²⁷ was determined with the partial sequence from an integrated proviral clone⁵³ (Fig. 1).

The HTLV-III provirus is 9,749 base pairs (bp) long. The overall structure of the provirus resembles that of other retroviruses. The sequences that encode viral proteins are flanked by LTR sequences. The LTR itself is flanked by inverted repeated sequences two nucleotides long (Fig. 1). Four long open reading frames are identified in the viral DNA (Fig. 2).

Long Terminal Repeat

A detailed analysis of the HTLV-III LTR is presented elsewhere⁵⁴; it is 634 nucleotides long with U3, R and U5 regions of 453, 98 and 83 nucleotides, respectively. The boundaries of these regions of the LTR were defined by localization of the 5'-cap site by S₁ nuclease mapping and by measurement of the length of the strong stop DNA transcript, as well as by determination of the 3'-terminus of the viral RNA by sequence analysis of cDNA clones. A TATAA sequence typical of eukaryotic promoters, as well as the consensus sequence for polyadenylation, are indicated in Fig. 1. A transfer RNA binding site complementary to the 3' end of tRNA^{Lys} is located 3' to the 5' LTR. DNA sequence homologies to the LTR of HTLV-I³², -II⁵⁵ and BLV⁵⁶ are indicated in Fig. 3.

尽管 HTLV-III 与 HTLV-BLV 病毒家族其他成员之间有相似性，但是 HTLV-III 与它们的生物学和病理学特征有着根本上的不同。HTLV-III 的感染常常导致完全的免疫抑制 (AIDS)，随后就是 OKT4⁺ 细胞数减少^[10-14,41-43]。这一效应是通过 HTLV-III 感染体外淋巴细胞培养物 OKT4⁺ 造成的细胞病变反映出来的，而非转染效应^[10,11,20]。相反地，细胞感染 HTLV-I 后仅导致一种低发病率的 T 淋巴细胞白血病（一种 OKT4⁺ 细胞恶性肿瘤）^[1-6]，而且也有 HTLV-I 病人存在不同程度的免疫缺陷的例子^[6,44]。HTLV-I 和 -II 体外感染导致培养的原代淋巴细胞大部分变为 OKT4⁺ 细胞^[45,46]。HTLV-I 感染淋巴细胞引起的细胞病变效应是明显的，但是这一效应不如 HTLV-III 引起的显著^[16,17,45-48]。HTLV-III 与 HTLV-I 和 -II 的不同之处还在于它们在体内和体外产生的具有感染能力的病毒颗粒的数量不同。从艾滋病患者的精液和唾液以及 HTLV-III 感染的细胞培养液上清中能够获得高效价的感染性病毒颗粒^[10,49-51]。从 ATLL 病人或者从 HTLV-I 和 -II 感染的细胞培养基中，很少能够获得游离的病毒粒子^[52]。

为了研究这些病毒在体内和体外的生物学活性并为艾滋病的诊断和治疗药物的开发提供有用的信息，我们完成了 HTLV-III 前病毒完整核苷酸序列的测序。

HTLV-III 的基因组结构

从几个美国艾滋病患者血液中提取 HTLV-III，然后感染 H9 细胞系，从中获得了几个密切相关的 HTLV-III DNA 克隆^[10]。利用一个完整前病毒克隆的部分序列^[53] (图 1)，我们测定了长度^[27] 分别为 8,900、5,300 和 3,600 个碱基的三个不完整病毒克隆的全部初级序列。

HTLV-III 前病毒长 9,749 个碱基对。前病毒的总体结构类似于其他的逆转录病毒。编码病毒蛋白的序列两侧为 LTR 序列。LTR 自身的两侧是 2 个核苷酸长度的反向重复序列 (图 1)。在病毒 DNA 中发现了四个长的开放阅读框 (图 2)。

长末端重复序列

HTLV-III LTR 的详细分析已经被报道过^[54]；该序列长 634 个核苷酸，有 U3、R 和 U5 三个区域，长度分别是 453、98 和 83 个核苷酸。这些区域在 LTR 中的边界是通过 S₁ 核酸酶谱和测量强制终止 DNA 转录产物的长度定位 5' 端帽子位点，以及通过 cDNA 克隆的序列分析来确定病毒 RNA 的 3' 末端，这些方法来确定的。在图 1 中标明了 TATAA 序列，它是真核启动子的典型序列，保守的多腺嘌呤序列也被标出。与 tRNA^{Lys} 的 3' 末端互补的转运 RNA 结合位点位于 5' LTR 的 3' 末端。在图 3 中标出了与 HTLV-I^[32]、-II^[55] 和 BLV^[56] 的 LTR 同源的序列。

Complete Nucleotide Sequence of the AIDS Virus, HTLV-III

CLONE	NUCLEOTIDE POSITION	AMINO ACID RESIDUE
BH10	59	277.1
BH5	345	38.1
BH10	345	38.1
BH5	270	29.6
BH10	195	406
BH5	120	4.51
BH10	120	4.51
BH5	296	4.51
BH10	45	296
BH5	120	4.51
BH10	39	514
BH5	39	506
BH10	75	322.1
BH5	75	53.1
BH10	221	329.6
BH5	221	55.6
BH10	296	37.1
BH5	296	13
BH10	371	33.1
BH5	371	38.1
BH10	446	60.6
BH5	446	38
BH10	521	63.1
BH5	521	6.3
BH10	596	35.6
BH5	596	8.8
BH10	671	36.1
BH5	671	11.3
BH10	746	37.6
BH5	746	13.8
BH10	821	38.1
BH5	821	16.3
BH10	896	38.6
BH5	896	18.8
BH10	971	39.1
BH5	971	21.3
BH10	1046	40.6
BH5	1046	23.8
BH10	1121	41.1
BH5	1121	26.5
BH10	1196	41.6
BH5	1196	28.8
BH10	1271	42.1
BH5	1271	31.3
BH10	1346	42.6
BH5	1346	33.8
BH10	1421	43.1
BH5	1421	36.5
BH10	1496	43.6
BH5	1496	38.8
BH10	1571	44.1
BH5	1571	41.5
BH10	1646	44.6
BH5	1646	43.8
BH10	1721	45.1
BH5	1721	46.5
BH10	1796	45.6
BH5	1796	48.8
BH10	1871	46.1
BH5	1871	51.2
BH10	1946	46.6
BH5	1946	53.8
BH10	2021	47.1
BH5	2021	56.1
BH10	2096	47.6
BH5	2096	58.6
BH10	2171	48.1
BH5	2171	61.1
BH10	2246	48.6
BH5	2246	63.6
BH10	2321	49.1
BH5	2321	66.1
BH10	2396	49.6
BH5	2396	68.6
BH10	2471	50.1
BH5	2471	71.1
BH10	2546	50.6
BH5	2546	73.6
BH10	2621	51.1
BH5	2621	76.1
BH10	2696	51.6
BH5	2696	78.6
BH10	2771	52.1
BH5	2771	81.1
BH10	2846	52.6
BH5	2846	83.6
BH10	2921	53.1
BH5	2921	86.1
BH10	2996	53.6
BH5	2996	88.6
BH10	3071	54.1
BH5	3071	91.1
BH10	3146	54.6
BH5	3146	93.6
BH10	3221	55.1
BH5	3221	96.1
BH10	3296	55.6
BH5	3296	98.6
BH10	3371	56.1
BH5	3371	101.1
BH10	3446	56.6
BH5	3446	103.6
BH10	3521	57.1
BH5	3521	106.1
BH10	3596	57.6
BH5	3596	108.6
BH10	3671	58.1
BH5	3671	111.1
BH10	3746	58.6
BH5	3746	113.6
BH10	3821	59.1
BH5	3821	116.1
BH10	3896	59.6
BH5	3896	118.6
BH10	3971	60.1
BH5	3971	121.1
BH10	4046	60.6
BH5	4046	123.6
BH10	4121	61.1
BH5	4121	126.1
BH10	4196	61.6
BH5	4196	128.6
BH10	4271	62.1
BH5	4271	131.1
BH10	4346	62.6
BH5	4346	133.6
BH10	4421	63.1
BH5	4421	136.1
BH10	4496	63.6
BH5	4496	138.6
BH10	4571	64.1
BH5	4571	141.1
BH10	4646	64.6
BH5	4646	143.6
BH10	4721	65.1
BH5	4721	146.1
BH10	4796	65.6
BH5	4796	148.6
BH10	4871	66.1
BH5	4871	151.1
BH10	4946	66.6
BH5	4946	153.6
BH10	5021	67.1
BH5	5021	156.1
BH10	5096	67.6
BH5	5096	158.6
BH10	5171	68.1
BH5	5171	161.1
BH10	5246	68.6
BH5	5246	163.6
BH10	5321	69.1
BH5	5321	166.1
BH10	5396	69.6
BH5	5396	168.6
BH10	5471	70.1
BH5	5471	171.1
BH10	5546	70.6
BH5	5546	173.6
BH10	5621	71.1
BH5	5621	176.1
BH10	5696	71.6
BH5	5696	178.6
BH10	5771	72.1
BH5	5771	181.1
BH10	5846	72.6
BH5	5846	183.6
BH10	5921	73.1
BH5	5921	186.1
BH10	5996	73.6
BH5	5996	188.6

CLONE

核苷酸位置 氨基酸残基

Table with 3 columns: CLONE, Nucleotide position, and Amino acid residue. It contains a detailed alignment of HTLV-III sequences with various restriction enzyme sites (Hind III, Eco RI, Pvu II, etc.) and reading frame markers (ORF I, ORF II, ORF III) indicated above the sequences.

9 12 15 17 19 22 24 27 29 32 34 37 39 42 44 47 49 51 52 54 57 59 62 64 67 71 74 77 79 82 84 86 88 91 94 97 100 103 106 109 112 115 118 121 124 127 130 133 136 139 142 145 148 151 154 157 160 163 166 169 172 175 178 181 184 187 190 193 196 199 202 205 208 211 214 217 220 223 226 229 232 235 238 241 244 247 250 253 256 259 262 265 268 271 274 277 280 283 286 289 292 295 298 301 304 307 310 313 316 319 322 325 328 331 334 337 340 343 346 349 352 355 358 361 364 367 370 373 376 379 382 385 388 391 394 397 400 403 406 409 412 415 418 421 424 427 430 433 436 439 442 445 448 451 454 457 460 463 466 469 472 475 478 481 484 487 490 493 496 499 502 505 508 511 514 517 520 523 526 529 532 535 538 541 544 547 550 553 556 559 562 565 568 571 574 577 580 583 586 589 592 595 598 601 604 607 610 613 616 619 622 625 628 631 634 637 640 643 646 649 652 655 658 661 664 667 670 673 676 679 682 685 688 691 694 697 700 703 706 709 712 715 718 721 724 727 730 733 736 739 742 745 748 751 754 757 760 763 766 769 772 775 778 781 784 787 790 793 796 799 802 805 808 811 814 817 820 823 826 829 832 835 838 841 844 847 850 853 856 859 862 865 868 871 874 877 880 883 886 889 892 895 898 901 904 907 910 913 916 919 922 925 928 931 934 937 940 943 946 949 952 955 958 961 964 967 970 973 976 979 982 985 988 991 994 997 1000

Fig. 1. Nucleotide sequence of HTLV-III. The complete nucleotide sequence of clone BH10 is shown together with the predicted amino acid sequence of the four largest open reading frames. The position of sequences encoding gag protein p17, the N-terminus of gag p24 and the C-terminus of gag p15 (which overlaps with the N-terminus of the pol protein) are indicated. The open reading frames for pol, sor and env-lor are indicated. The sequence of the remaining 182 bp of the HTLV-III provirus not present in clone BH10 (including a portion of R, U5, the tRNA primer binding site and a portion of the leader sequence) was derived from clone HXB2. The boundaries of R, U5 and U3, the positions of the polypurine tract, inverted repeated sequences (IR) and the transcriptional initiation (TATA) and termination (AATAA) signals are shown. The sequences of BH8 and BH5 are illustrated also; nucleotide and predicted amino acid differences compared with BH10 are listed and dashes are shown for identical sequences. Restriction enzyme sites are listed above the nucleotide sequence and sites present in clone BH8 but not BH10 are in parentheses. Deletions are also noted ([]) at nucleotides 251, 254, 5,671 and 6,987-7,001. The nucleotide positions (to the right of each line) start with the nucleotide initiation site and end with the viral RNA transcriptional termination site. The amino acid sequence is numbered (to the right of each line) for the four largest open reading frames starting after the preceding termination codon in each case except gag which is enumerated from the first methionine codon. A proposed peptide cleavage site (v) and possible asparagine-linked glycosylation sites (77) are shown (*) for the env-lor open reading frame. The sequences in the LTR derived from clones BH8 and BH10 listed in the beginning of the figure are derived from the 3' portion of each clone and are assumed to be identical to those present in the 5' LTR of the integrated copies of these viral genomes. Clone HXB2 was derived from a recombinant phage library of XbaI-digested DNA from HTLV-III-infected H9 cell cloned in λJ1 (ref. 53). Clones BH10, BH8 and BH5 were derived from a library of SstI-digested DNA from the Hirt supernatant fraction of HTLV-III-infected H9 cells cloned in λgtWes-λB (ref. 27). Both libraries were screened with a cDNA probe synthesized from virion RNA using oligo(dT) as a primer²⁶. Clones BH8, BH5 and a portion of HXB2 were sequenced as described previously⁹¹. Clone BH10 was sequenced by the method of Sanger⁹² modified by the use of oligonucleotides complementary to the M13 insert sequence as primers and using Klenow fragment of DNA polymerase I or reverse transcriptase as the polymerase.